

Peptides with anticancer use or potential

Review Article

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Received March 3, 2002

Accepted October 3, 2002

Published online January 20, 2003; © Springer-Verlag 2003

Summary. This review is an attempt to illustrate the diversity of peptides reported for a potential or an established use in cancer therapy. With 612 references, this work aims at covering the patents and publications up to year 2000 with many inroads in years 2001–2002. The peptides are classed according to four categories of effective (or plausible) biological mechanisms of action: receptor-interacting compounds; inhibitors of protein-protein interaction; enzymes inhibitors; nucleic acid-interacting compounds. The fifth group is made of the peptides for which no mechanism of action has been found yet. Incidentally this work provides an overview of many of the modern targets of anticancer research.

Keywords: Peptides – Antitumor

I Introduction

This review is an attempt to illustrate the diversity of characterised peptides reported for a potential or an established use in cancer therapy. The origin and *raison d'être* of this work stemmed from the coincidence described here:

In 1995, the co-crystallisation of Ras binding domain of Raf-1 and Rap-1 (a protein related to oncogenic protein Ras) enabled an X ray analysis of the interacting amino acids residues (Nassar et al., 1995). One interesting feature of protein Rap-1 binding domain is the sequence 37–39: **Glu-Asp-Ser**. These residues not only follow each other on the peptide backbone but the two carboxylic residues are also involved in very strong ionic interactions with Raf positively charged guanidinium residues; namely Arg-59 and Arg-89. Thus, it could be proposed that small peptides containing the motif Glu-Asp-Ser were potential Ras-Raf interaction inhibitors and hence

potential leads for an anticancer medicinal chemistry research project. This starting point was followed by some literature search which came out with the remarkable fact that the pentapeptide pGlu-**Glu-Asp-Ser-Gly** is an inhibitor of epidermal mitoses (Jensen et al., 1990). This fact probably led to the simultaneous development of two analogues by pharmaceutical companies as cell proliferation inhibitors (Balazs et al., 1992; Laerum, 1990). However, despite some trials, we could not confirm the inhibition of Ras-Raf interaction as the actual mechanism of this antiproliferative action. If nothing, this curiosity pointed out a possible need for a review of peptidic derivatives with an actual or a potential use in antitumor therapy; especially since this type of substance is more and more often a starting point in medicinal chemistry (Mizejewski, 2001).

Only a handful of peptides, such as the LH-RH agonists and somatostatin analogues, are actually used as anticancer drugs (Loffet, 2002). Even if more compounds are currently at various clinical trials stages (Jimeno, 2002), peptides even only reported for a cytotoxicity, an antiproliferative action or displaying a potentially useful effect were included in this review. Indeed, the low bioavailability of peptides can restrict their action to cell culture or even to molecular mechanism models and thus seriously hamper any pharmaceutical potential. However, we thought that enlarging our search to as many peptides as possible and thus to as many molecular mechanisms as possible would still be of interest. We excluded all the “large” proteins,

the main reason being an organic chemist point of view, since preparation and use of these cytokines for cancer therapy has become one of the many achievements of the molecular biochemists (Maini et al., 1997; Mueller, 1998; Oppenheim et al., 1997). The many peptidic substances involved in the stimulation of an immune response to cancer (Velders et al., 1998) were excluded as well as the peptide-containing prodrugs and the amphipatic peptides better known for their antimicrobial properties (Jacob and Zasloff, 1994) than for their antitumor potentials (Shin et al., 1999). The present manuscript aims at covering the patents and publications up to year 2000 with many inroads in years 2001–2002. The peptides are classed according to four categories of effective (or plausible) biological mechanisms of action: receptor-interacting compounds; inhibitors of protein-protein interaction; enzymes inhibitors; nucleic acid-interacting compounds. The fifth group is made of the peptides for which no mechanism of action has been found yet. Concerning this last group, the yearly reviews of D. J. Faulkner on marine natural products illustrate quite well the number of naturally-occurring cytotoxic peptides reported (Faulkner, 2002). However it was beyond the scope of this review to compare their respective level of cytotoxicity (if such thing is possible given the many different test used) or even to depict all the substances isolated.

II Receptor-interacting compounds

II.1 Luteinizing hormone-releasing hormone agonists and antagonists

In 1971 the decapeptidic structure of a luteinizing hormone-releasing hormone (LH-RH)/Gonadotrophin-releasing hormone (pGlu-His-Trp-Ser-Tyr-Gly-Leu-Arg-Pro-Gly-NH₂) was reported (Schally, 1999). This is one of the hormonal messages issued from the hypothalamus to the pituitary gland. Its action leads to the release of luteinizing hormone and follicle-stimulating hormone which acts on ovaries and testes, and thus, on the release of steroidal hormones. Synthesis of many analogues led to structure-activity relationships which were reviewed (Dutta, 1988a; Dutta, 1988b; Kutscher et al., 1997). Compounds currently used in hormonal therapies such as treatment of sex hormone-dependent malignant neoplasms lead to objective stable disease or partial remission (Hoffken and Kath, 2000; Schally, 1999). A noteworthy feature

is that the agonists currently used were found by “simply” replacing the glycine moiety in position 6 with various D-amino acids such as D-Trp, D-Leu or D-Ser. This change not only confers a stability toward the protease that quickly hydrolyses LH-RH but also a receptor affinity increase. A review (Kutscher et al., 1997) describes the remarkable and much more important structural changes necessary to obtain potentially useful (Huirne and Lambalk, 2001) antagonists such as cetrorelix (Fig. 1) (Reissmann et al., 1994). Other antagonist described are for example a Nal-Glu derivative (Rivier et al., 1986), ganirelix (Nestor et al., 1992) or abarelix (Molineaux et al., 1998). As described in a recent article, present research is focused on the design of long-acting antagonists (Jiang et al., 2001).

II.2 Mammalian gastrin-releasing peptide (bombesin) analogues

The human 27-mer peptide named Gastrin-releasing peptide (GRP), unlike the amphibian-originated tetradecapeptide bombesin (pGlu-Gln-Arg-Leu-Gly-Asn-Gln-Trp-Ala-Val-Gly-His-Leu-Met-NH₂), is a neuronal-originated hormone. Initially, both compounds were found to trigger the gastrin release *in vivo*. However, even if many other physiological actions were later found, the original names perpetuate. Bombesin and GRP share important sequence homologies and, based on immunology or gene cloning, searches for other human endogenous peptides were made. From this, emerged a picture (de Castiglione and Gozzini, 1996; Preston et al., 1996) involving, so far, two different peptides: the 27-mer GRP (Battey and Wada, 1991) and the 32-mer neuromedin B (Ohki-Hamazaki, 2000). Moreover, as three different mammalian receptors have been found (Nagalla et al., 1995; Preston et al., 1996), a specific

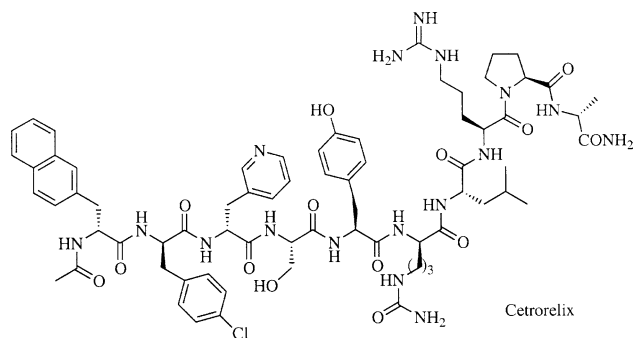


Fig. 1. Structure of Cetrorelix

endogenous ligand for the third type remain to be characterised (Ryan et al., 1998). The structural characteristics of the many known agonists were reviewed (Lin et al., 1995; Raynor et al., 1993) and recent works focus upon the determination of the receptor's residues involved in ligand binding (Sainz et al., 1998; Tokita et al., 2001). More than 10 years ago, it was recognised that the GRP secreted by malignant cells such as small cell lung cancer (SCLC) and medullary thyroid carcinoma could act as an autocrine/paracrine growth factor. This would explain some early results such as the mitogenic action of GRP on fibroblast cell line Swiss 3T3 or its *in vitro* growth action on SCLC (Cuttitta et al., 1985; Moody et al., 1981). More recently, neuromedin B was found to have a similar effect (Cardona et al., 1991). The search for antagonists with a potential use in anticancer treatment was thus rationalised. The first antagonists, which have peptidic structures based either on bombesin, litorin or substance P, have been shown (Thomas et al., 1992) to partially inhibit tumour growth in SCLC xenografted nude mice and were reviewed (de Castiglione and Gozzini, 1996). Since then, more compounds active *in vivo* have been reported (Burman et al., 2002; Burman et al., 2001a; Everard et al., 1993; Langdon et al., 1992; Matsumoto et al., 2000; Orosz et al., 1995; Reile et al., 1995). As an illustration, the short analogues D-(NMe)Phe-D-Trp-Phe-D-Trp-Leu Ψ (CH_2NH)Leu-NH₂ or derivatives of Phe-D-Trp-Phe-D-Trp-Leu-Leu were reported (Nyeki et al., 1998; Orosz, 2001; Orosz et al., 1994) and the antagonist RC-3095 (Radulovic et al., 1991), (Fig. 2) which bears a reduced bond between the two C-terminal leucines, was found active on a whole array of tumour models (de Castiglione and Gozzini, 1996).

Recent works and current hypothesis (Ohlsson et al., 1999; Petit et al., 2001), which were reviewed

recently (Heasley, 2001), point out the possible role of many other neuropeptides (i.e.: neurotensin, gastrin, cholecystokinin or arginine vasopressin) as autocrine/paracrine factors of tumour growth. Whether the use of finely tuned antagonists or the use of compounds active on a broad spectrum of neuropeptide receptors provide the best objective response will hopefully be determined in the future (Heasley, 2001; Sethi et al., 1992).

II.3 Somatostatin analogues

Somatostatin was initially isolated from ovine hypothalami and described as a regulator of growth hormone secretion (Brazeau et al., 1973; Krulich et al., 1968). It was later demonstrated that the multiple effects caused by somatostatin are mediated by at least five somatostatin receptors (Lamberts et al., 1996; Patel et al., 1995). Thus, actions of somatostatin are seen on neurotransmission in the central nervous system, on the regulation of growth hormone and on thyrotropin release (Reichlin, 1983). It has also multiform regulatory roles on the gastrointestinal tract and on the pancreas. Two important bioactive somatostatins exist, the sulfide-bridged tetradecapeptide somatostatin 14: Ala-Gly-Cys-Lys-Asn-Phe-Phe-Trp-Lys-Thr-Phe-Thr-Ser-Cys, and somatostatin 28, which is extended by 14 more residues from the aminated end of somatostatin 14. With the necessity to dispose of more stable somatostatin receptor agonists, the first "wave" of syntheses led to many analogues (Bauer et al., 1982; Cai et al., 1986; Kéri et al., 1993; Murphy et al., 1985; Raynor et al., 1993; Taylor et al., 1988; Veber et al., 1981). Amongst them are established anticancer drugs such as octreotide or somatuline (Fig. 3) (Bauer et al., 1982; Bogden et al., 1990; Lamberts et al., 1996). Both

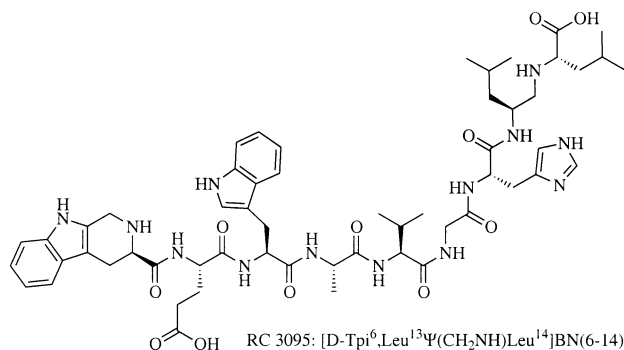


Fig. 2. Structure of the bombesin-derived antagonist RC 3095

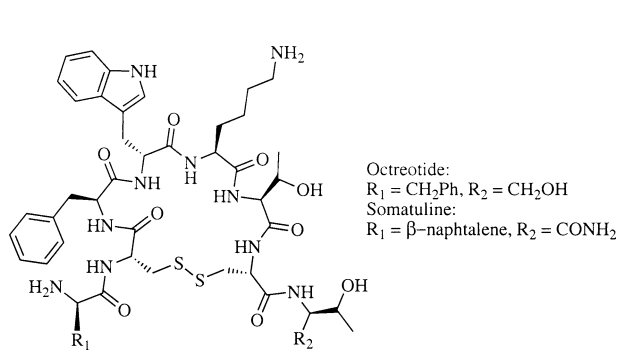


Fig. 3. Structure of octreotide and somatuline

compounds are bearing a β -turn feature, made by Phe-D-Trp-Lys-Thr, which is crucial for the activity.

The current second “campaign” of research of subtype-specific receptors (Weckbecker et al., 1993) agonists and antagonists (Bass et al., 1996; Bass et al., 1997; Gazal et al., 2002; Hocart et al., 1999; Morgan and Sadat-Aalae, 2001) could lead to a better understanding of somatostatin roles and possibly much more specific treatments (Bousquet et al., 2001). A recent approach, replacing α -amino acids with β -amino acids, met success as a linear β -peptide (Fig. 4) was found to have a strong and specific affinity for somatostatin subtype 4 receptor (sst-4) (Gademann et al., 2001). Another noteworthy compound is a cyclic disulfide (Fig. 4) which was found to be an antagonist of somatostatin (Hocart et al., 1999).

Moreover, although somatostatin analogue therapy has its importance (Eriksson and Oberg, 1999), all tumour growth become eventually unresponsive to the treatment. Thus, further strategies based on specific targeting of toxic substances – possibly ^{90}Y terbium or other radio-labelled compounds (Albert et al., 1998; Heppeler et al., 2000; Kwekkeboom et al., 2000; Oberg, 2001; Szegedi et al., 1999) – may lead to new protocols (Eriksson and Oberg, 1999).

II.4 Less investigated examples

II.4.1 Growth hormone-releasing hormone. The growth hormone-releasing hormone (GH-RH) peptide belongs to a group of structurally related hormones that includes vasoactive intestinal peptide (VIP), pituitary adenylate cyclase-activating polypeptide, secretin and glucagon (Campbell and Scanes, 1992).

– From the structure of GH-RH, a 29-mer antagonist was found to have an *in vivo* antitumor effect on

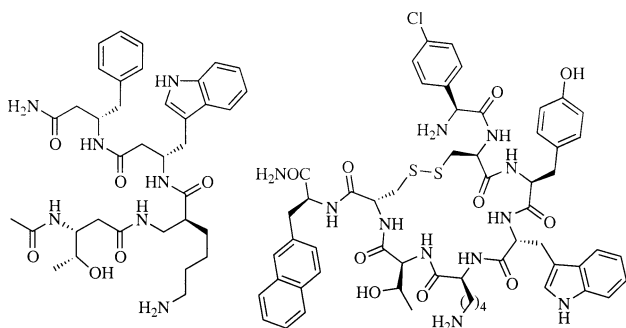


Fig. 4. The first β -peptide ligand of sst-4 and an antagonist of somatostatin

nude mice bearing xenografts (Kiaris et al., 1999; Varga et al., 1999). However, the actual mechanism of action of this antagonist may be more complex (Kineman, 2000). Concerning the 28-mer VIP, the presence of VIP receptor in non-small cell lung cancer led to the design of an antagonist. This peptide is made of a 22 amino acids-long segment of VIP linked with an N-terminal sequence (Lys-Pro-Arg-Arg-Pro-Tyr), added to increase its cell membrane permeability. This antagonist significantly inhibited xenograft formation in nude mice (Moody et al., 1993). More recently, longer unnatural peptides were patented for sub nanomolar cytotoxicities (Burman et al., 2001b).

– A 33-mer fragment of pituitary adenylate cyclase-activating polypeptide was also reported to inhibit the growth of prostate cancer cells bearing the corresponding receptor (Leyton et al., 1998). Recent works have focused on the preparation of more specific antagonists (Rekasi et al., 2000).

II.4.2 Vascular endothelial growth factor. Inhibition of the vascular endothelial growth factor (VEGF) signalling abrogates the development of many tumors (Neufeld et al., 1999). From a peptide library, some arginine-rich hexapeptides were found to inhibit the interaction between VEGF₁₆₅ and the VEGF receptor (Bae et al., 2000). For example, on a nude mice model, the peptide Arg-Arg-Lys-Arg-Arg-Arg prevented the growth and metastasis of the VEGF-secreting HM7 human colon carcinoma cells. Other peptides were also patented (Schatz et al., 2001). Another three different classes of disulfide-bridged peptide capable of inhibiting VEGF interaction with its receptor were found starting from a phage display method (Fairbrother et al., 1998). In a different approach, an inhibition of the interaction between VEGF and VEGF receptor-2 by the peptide Ala-Thr-Trp-Leu-Pro-Pro-Arg leads to a proliferation inhibition (Binétry-Tournaire et al., 2000). Moreover, the 18-mer Arg-Thr-Glu-Leu-Asn-Val-Gly-Ile-Asp-Phe-Asn-Trp-Glu-Tyr-Pro-Ala-Ser-Lys, also derived from VEGF receptor 2, inhibits proliferation and migration of microvascular endothelial cells at a micromolar concentration (Piossek et al., 1999). Other peptides, such as His-His-Glu-Val-Val-Lys-Phe-Met-Asp-Val-Tyr-Gln which is derived from the exon 6 sequence of VEGF, were also reported for their inhibition of endothelial cell responses (Jia et al., 2001). Finally, the small peptide Asn-Ile-Thr-Val-Thr-Leu-Lys-Lys-

Phe-Pro-Leu, derived from the sequence of VEGF receptor-1, was reported recently as an angiogenesis inhibitor although this peptide does not bind VEGF nor inhibits its binding to the corresponding receptor (Tan et al., 2001).

II.4.3 Epidermal growth factor. Some cyclic disulfide peptides, derived from the epidermal growth factor (EGF), such as Cys-His-Ser-Gly-Tyr-Val-Gly-Val-Arg-Cys were patented for their inhibitory activity of EGF-induced cell proliferation (Nestor et al., 1985). Moreover the peptidic motifs Leu-Gly-Leu-Arg-Ser-Leu-Arg-Glu or Leu-Gly-Leu-Arg-Ser-Leu-Lys-Glu were claimed to antagonise the EGF receptor (Lupu and Lippman, 1994). Fairly more complex compounds bearing three disulfide bridges (a T-knot feature) were also found to antagonise the EGF (Blanco-Aparicio et al., 1998). In a different approach, the product of HER2, which is also a member of the EGF family, was found to be inactivated by small peptides mimicking antibodies such as trastuzumab (Herceptin). Thus, the dodecapeptide Tyr-Cys-Asp-Gly-Phe-Tyr-Ala-Cys-Tyr-Met-Asp-Val-NH₂ binds to the HER2 receptor and antagonises its constitutive growth signalling properties *in vitro* (Berezov et al., 2001). A recent review of inhibitors of the EGF receptor as potential anticancer compounds describes other approaches (Woodburn, 1999).

II.4.4 Interleukin 6. Antagonists hindering the binding of interleukin 6 to its receptor were found using phage display techniques. Further modifications from the consensus sequence led to the cyclic disulfide peptide Gly-Gly-Cys-Lys-Leu-Trp-Thr-Ile-Pro-Glu-Cys-Gly-Gly which inhibited cellular growth (Mizuguchi et al., 2000).

II.4.5 Interleukin 8. A hexapeptide (Ac-Arg-Arg-Trp-Trp-Cys-Arg-NH₂) which inhibits the binding of interleukin 8 to neutrophils was found (Hayashi et al., 1997) to also inhibit the growth of melanoma cell lines stimulated by a 73-mer α -chemokine. This peptide is active *in vivo* as well (Fujisawa et al., 1999). On the other hand, the NH₂-terminal pentapeptide corresponding to endothelial interleukin 8 (Ala-Val-Leu-Pro-Arg) is responsible for apoptosis induction and has an antitumor effect *in vivo* (Terui et al., 1999).

II.4.6 Platelet-derived growth factor. The octadecapeptide Tyr-Gly-Arg-Pro-Arg-Glu-Ser-Gly-Lys-Lys-Arg-Lys-Arg-Lys-Arg-Leu-Lys-Pro-Thr is a fragment of the platelet-derived growth factor and was found to inhibit the growth of malignant glioma in athymic nude mice (Khachigian et al., 1995).

II.4.7 Tumour necrosis factor. Cyclic analogues of tumour necrosis factor, such as the cysteine-bridged Ac-Cys-Pro-Ser-Glu-Gly-Leu-Cys-NH₂ and Ac-Cys-Pro-Ser-Glu-Gly-Thr-Pro-Ser-Thr-His-Val-Leu-Cys-NH₂, were patented (Boehm et al., 1990c). The same research group also patented linear peptides such as Ac-Leu-Ala-Asn-Gly-Val-Glu or Pro-Gln-Ala-Glu-Gly-Gln-Leu-NH₂ for their tumour necrosis factor agonist or antagonist activities (Boehm et al., 1990a; Boehm et al., 1990b). The related sequence Val-Ala-Asn-Pro-Gln-Ala-Glu-Gly-Gln-Leu had actually been patented previously (Furuta and Hayashi, 1986). Moreover, cyclic peptides containing the murine tumour necrosis factor amino acids sequence 127–132 (cyclic Lys-Gly-Asp-Gln-Leu-Ser) or 59–66 (cyclic Tyr-Ser-Cln-Val-Leu-Phe-Lys-Gly) were found to have a weak cytotoxicity (Sheh et al., 1990; Sheh et al., 1993).

II.4.8 Alpha-feto protein. The octapeptide Glu-Met-Thr-Pro-Val-Asn-Pro-Gly, derived from the 590-mer alpha-feto protein, was shown to retain all the inhibition capacity against estrogen-dependant growth of human breast cells. Further work demonstrated that this peptide would undergo a time-dependant activity loss via a hydrophobic-based self aggregation phenomenon. The substitution of the two lipophilic prolines residues with the more polar 4-hydroxy prolines led to a stable analogue which is an inhibitor of the estrogen-dependant growth of MCF-7 human breast cancer (Mesfin et al., 2001). Remotely related to this approach is a recent patent which reports peptides, found by a phage-display technique, that mimic the biological activity of steroid hormones (Kohen et al., 2002).

II.4.9 Sialyl-Lewis mimics. Peptidic derivatives mimicking sialyl-Lewis carbohydrates were recently found using, respectively, combinatorial chemistry and phage display techniques (Fukuda et al., 2000; O et al., 1999). One of the peptide (Ile-Glu-Leu-Leu-Gln-Ala-Arg) inhibits the sialyl Lewis X-dependent lung colonisation of tumour cells *in vivo* by antagonising the selectin receptors (Fukuda et al., 2000).

III Inhibitors of protein-protein interaction

This section concerns compounds able to block cancer-related biological processes involving a protein-protein interaction (Huber et al., 1994). Many peptides should be placed in this section although either the actual interaction inhibited has not been

found or has not been recognised as such yet. Moreover the distinction between this class and the receptor-interacting class is somewhat arbitrary since what is the difference between two interacting proteins and the binding of a protein to its receptor?

III.1 Urokinase-type plasminogen activator interfering compounds

Cell migration and invasiveness are crucial steps in cancer where tissue remodelling, angiogenesis and metastasis and leading to critical situations. The serine protease urokinase-type plasminogen activator (uPA) and its cell surface urokinase plasminogen activator receptor (uPAR) are central to these processes (Andreasen et al., 1997; Mazar, 2001; Reuning et al., 1998; Weidle and Koenig, 1998). Interaction between the uPA and uPAR not only enables the activation of plasmin (via cleavage of the proenzyme plasminogen) but also focuses the plasmin proteases activity on the cell surface. A review further describes the complexity of this activation system in which some components have been found to be relevant indicators for patient prognosis in human cancer (Reuning et al., 1998).

Thus inhibition of the uPA/uPAR interaction was the focus of some research (Fazioli and Blasi, 1994; Frankenne et al., 1999). Peptides related to the 18–32 amino acids sequence of uPA, which is involved in the binding with uPAR (Bürgle et al., 1997), were found to have an effect on *in vitro* and *in vivo* invasiveness models (Kobayashi et al., 1994a; Kobayashi et al., 1993). The disulfide-bridged cyclic tridecapeptide uPA_{19–31} (Cys-Val-Ser-Asn-Lys-Tyr-Phe-Ser-Asn-Ile-His-Trp-Cys) has been suggested as a lead for further development of antagonists (Bürgle et al., 1997). On the other hand, phage display-based research led to the discovery of a group of short peptides (subsequence motifs Phe-X-X-Tyr-Lys-Trp or Lys-Trp-X-X-Ar; Ar being Tyr, Phe, His or Trp), unrelated to the above-mentioned uPAR binding sequence, that antagonise the uPA/uPAR interaction (Goodson et al., 1994). The same research group reported that compounds as short as the decapeptide Leu-Asn-Phe-Ser-Gln-Tyr-Leu-Trp-Tyr-Thr-NH₂ retained affinity in the nanomolar range for the uPAR (Tressler et al., 1999). It is noteworthy that the tumour growth inhibition observed in some cases may also be caused by the peptide promotion of the binding of uPAR cell-bearing to vitronectin (Tressler et al., 1999). Peptides containing Ser-Arg-Ser-Arg-Tyr, also corresponding

to a uPAR sequence, were patented (Blasi et al., 1998). And an octapeptide derived from another non receptor binding region of uPA (Ac-Lys-Pro-Ser-Ser-Pro-Pro-Glu-Glu-NH₂) was found to be a non competitive inhibitor of tumour progression and angiogenesis *in vivo* (Guo et al., 2000; Mishima et al., 2000).

III.2 p53 (Hdm2, p14^{ARF} and p53 C-terminal regulatory domain)

Mutations on the p53 gene or of the proteins that regulates p53 have been found in at least 80% of all human tumours (Helmreich, 2001). The key role of this protein is indeed the prevention, or the postponing, of the multiplication of DNA-damaged cells such as cancerous one. Accordingly, compounds able to restore p53 function – to rescue it pharmacologically (Foster et al., 1999) – could act alone, or in concert with classical DNA-damaging anticancer agent, as remarkably original approaches to anticancer treatment. Several research avenues involving p53 were recently reviewed (Hupp et al., 2000).

– The protein Hdm2 (Human double minute 2) or its murine equivalent Mdm2 were suggested to be responsible for the (necessary) cellular degradation of p53, via a binding to its N-terminus part, followed by ubiquitin-dependent proteolysis (Böttger et al., 1996; Lane and Hall, 1997). An overexpression of this protein would thus disrupt the normal function of p53. Accordingly, an approach started with the sequence of p53 Hdm2-binding site: Thr-Phe-Ser-Asp-Leu-Trp (Böttger et al., 1997). Since this peptide was only a very weak inhibitor of the interaction, a phage display-based search was conducted and led to the more active inhibitor Ac-Met-Pro-Arg-Phe-Met-Asp-Tyr-Trp-Glu-Gly-Leu-Asn-NH₂ (Böttger et al., 1996). From this, a feat of synthesis, quite driven by X-ray structure-based optimisation, led to very efficient *ex vivo* antagonists (García-Echeverría et al., 2000; Luke et al., 1999). One of the most active octapeptide (Fig. 5) was shown to stimulate, albeit – because of a low cell permeability – at a high concentrations, the p53 pathway in tumour cell lines (Chène et al., 2000). Curiously, two facts probably open new avenues of research. The first is a patent which claims antitumor peptides, derived from the N-terminal part of p53, that are able to block a p53-retinoblastoma protein interaction

(Kouzarides, 1998). The second is that the first 25 N-terminal amino acids of p53 were recently proven to interact with tubulin (Giannakakou et al., 2000). It is thus tempting to suggest that peptides mimicking the N-terminal moiety of p53 have at least three proteins as potential targets. Moreover, the naturally occurring chlorofusin (Fig. 5), isolated from *Fusarium sp.*, was found to be an antagonist of the p53-Mdm2 interaction at micromolar concentration (Duncan et al., 2001).

- One step removed from p53 is the p14^{ARF} protein which binds to Mdm2 and thus indirectly prevents the p53 inactivation. A 20 amino acid-long peptide (Met-Val-Arg-Arg-Phe-Leu-Val-Thr-Leu-Arg-Ile-Arg-Arg-Ala-Cys-Gly-Pro-Pro-Arg-Val) corresponding to the beginning of the p14^{ARF} sequence was recently demonstrated to induce p53 protein and prevent its ubiquitination (Midgley et al., 2000).
- Peptides corresponding to the carboxy-terminal sequence of p53 do restore its growth suppression function (Selivanova et al., 1997). Indeed, peptides derived from the C-terminal amino acids sequence 361–383 (Gly-Ser-Arg-Ala-His-Ser-Ser-His-Leu-Lys-Ser-Lys-Lys-Gly-Gln-Ser-Thr-Ser-Arg-His-Lys-Lys-Leu) of p53 have been patented for their ability to activate p53 function (Halazonetis and Hartwig, 1996; Halazonetis and Hartwig, 2001; Shibata et al., 1999). More is said about this case in the conclusion. These finding could be explained by the proposition that these peptides antagonise proteins which have an affinity for the p53 C-terminal negative regulatory domain (Hupp et al., 2000).

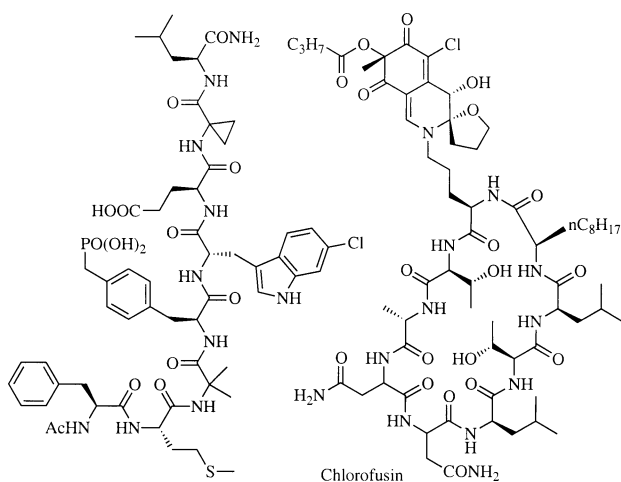


Fig. 5. A man-made Hdm2 antagonist and chlorofusin, a naturally occurring one

III.3 Inhibitors based on p34^{cdc2} / p33^{cdk2}, p21^{WAF1} and p16^{INK4}

It would be beyond the scope of this review to attempt to explain the role of the retinoblastoma related proteins such as p107 and pRb2/p130 in cell multiplication and cancer (Nevins, 2001). These proteins are transcriptional regulators that control the genes involved in the cellular cycling from G1 to S phase (Weinberg, 1995). The assembly of a protein complex made of a histone deacetylase-E2F and pRb is an example of how this control takes place. This trimeric structure binds DNA and thus represses its expression. However, if phosphorylation of pRb (by the many cyclin dependant kinase/cyclin complexes) takes place, pRb loses its affinity for E2F and the gene repression is lifted. Thus, these phosphorylation events can be considered as a ternary association of the cyclin, the cyclin dependant kinase and their protein substrates. These associations could be attractive targets for a selective protein-protein inhibition. A recent review further describes the system that regulates cyclin-dependent kinase (Cdk) and its importance for new anticancer drug design (McDonald III and El-Deiry, 2000). As far as we could tell, there are three different avenues of such research that led to peptides which are based either on the serine/threonine kinases p34^{cdc2} / p33^{cdk2}, the protein p21^{WAF1} or the protein p16^{INK4}.

- Serine/threonine kinases p34^{cdc2} / p33^{cdk2}. A patent describes inhibitors of the cell cycle regulatory serine/threonine kinases p33^{cdk2} and p34^{cdc2}. These two related kinases are associating with many proteins, including pRb2, p107 and the human papillomavirus oncoprotein E7. Cyclins binding to p34 or p33 is of course required for their kinase activity. The peptides Cys-Ala-Phe-Tyr-Ile, a longer homologue Leu-Cys-Ala-Phe-Tyr-Ile-Met-Ala-Lys, and Met-Cys-Ser-Met-Tyr-Gly-Ile-Cys-Lys, which are derive from the p34 binding domain of pRb2, p107 and cyclin E, were demonstrated to inhibit the interaction of p34 or p33 with these proteins. Hopefully, further work concerning their antitumor potential or their use as biological tools will be reported in the future (Webster and Coleman, 1994).

Another approach led to the anti-Cdk2 aptamer Tyr-ser-Phe-Val-His-His-Gly-Phe-Phe-Asn-Phe-Arg-Val-Ser-Trp-Arg-Glu-Met-Leu-Ala using an *Escherichia coli*-based display method along with the two

hybrids method and was found to inhibit the interaction of Cdk2 with histone H1 but not with pRb (Colas et al., 1996). As expression of this sequence led to the cell-cycle arrest, this work demonstrates a role for histone H1 phosphorylation (Cohen et al., 1998).

- p21^{WAF1}. The cdk inhibitor protein p21 is actually one of the gene products, mediating cell growth arrest, that are controlled by p53. This inhibition is achieved via protein-protein interaction between p21 and G1 cyclin-Cdk complexes (Chen et al., 1996). Using a series of 20-mer synthetic peptides that spanned the entire sequence of p21, the peptide Lys-Arg-Arg-Gln-Thr-Ser-Met-Thr-Ala-Phe-Tyr-His-Ser-Lys-Arg-Arg-Leu-Ile-Phe-Ser, closely related to the p21_{141–160} carboxy-terminal domain, was found to inhibit the cyclin D1-Cdk4 at nanomolar level and induced a G1/S growth arrest (Ball et al., 1996). The smaller fragment Lys-Arg-Arg-Leu-Ile-Phe-Ser-Lys did retain some activity. Expression of such peptide as a GFP miniprotein was also used to demonstrate a cell proliferation inhibition (Mattock et al., 2001). Other peptides, derived from two different regions of p21 (but one of them containing an Arg-Arg-Leu-Phe motif), were linked to the carrier/internalising hexadecapeptide sequence found in *Antennapedia* (Arg-Gln-Ile-Lys-Ile-Trp-Phe-Gln-Asn-Arg-Arg-Met-Lys-Trp-Lys-Lys) and were shown to inhibit human cancer cells growth (Bonfanti et al., 1997; Fischer et al., 2000). Further work demonstrated that an improved Cdk2 inhibition could be achieved by small structural changes. An example from the extensive structure-activity relationship study undertaken, is the alanine scan of p21 sequence 149–160 that led to the inhibitor Asp-Phe-Tyr-His-Ala-Lys-Arg-Arg-Leu-Ile-Phe-Ser-NH₂. This peptide is a 100 fold more active on Cdk2 cyclin E complex for the phosphorylation of GST-Rb (Zheleva et al., 2002).
- p16^{INK4}. The protein p16 is the only member of the INK4/CDKN2 gene products that is linked to tumour suppression. It acts at the level of Cdk-cyclin D interaction, possibly by binding to Cdk4 or in a more complex manner involving another protein (Fåhræus et al., 1998). A strategy, related to the one described above, led to the preparation of a 20-mer peptide, derived from p16 sequence, which is able to inhibit Cdk4-cyclin D1 kinase *in vitro* and block cell cycle progression through G1 (Fåhræus et al., 1996). Further studies led to a much smaller

peptide (Phe-Leu-Asp-Thr-Leu-Val-Val-Leu-His-Arg) which is still a kinase inhibitor although, contrary to its parent peptides, does not bind to the Cdk4 anymore. Moreover, when linked to the *Antennapedia* carrier sequence, it also stopped cell cycle progression (Fåhræus et al., 1998).

III.4 Inhibitors of E2F/DP and E2F/DNA interactions

Further down in the signalling cascade involving p16 is the heterodimeric association between the transcription factor E2F and another protein (of the DP family) prior to a binding to DNA. The E2F/DP transcription factor seems to have a role as both a tumour suppressor and an oncogene in mice (Yamasaki, 1999). A two hybrids strategy search was devised and led to aptamer and unconstrained peptidic compounds able to prevent this association (Fabbrizio et al., 1999). Moreover, one of the 20-mer peptide (Arg-Cys-Val-Arg-Cys-Arg-Phe-Val-Val-Trp-Ile-Gly-Leu-Arg-Val-Arg-Cys-Leu-Val) also inhibited E2F function *in vitro*. It is noteworthy that this peptide contains the sequence Trp-Ile-Gly-Leu which corresponds to a highly conserved motif present in the protein DP. Since a previous study (Bandara et al., 1997) showed that a 15-mer peptide containing this sequence had no significant effect – much longer one had – on E2F heterodimerisation, further structural requirement must be involved.

The naturally occurring hexadepsipeptide GE3 (Fig. 6) was found to be cytotoxic on tumour cell lines and, most remarkably, to probably interact with E2F (Sakai et al., 1997). It is noteworthy that many closely related depsipeptides such as azinotricin (Maehr et al., 1986), A83586C (Smitka et al., 1988), variapeptin and citropeptin (Nakagawa et al., 1990), aurantimycins (Gräfe et al., 1995), verucopeptin (Sugawara et al., 1993), polyoxypeptin (Umezawa et al., 1999) and pipalamycin (Uchihata et al., 2002) were also reported for their antitumor potential.

Another original avenue exists as the N-terminal dodecanoate of Leu-Asn-Trp-Ala-Trp-Ala-Ala-Glu-Val-Leu-Lys-Val-Gln-Lys-Arg-Arg-Ile-Tyr-Asp-Ile-Thr-Asn-Val-Leu-Glu-Gly-Ile-Gln-Leu-Ile-Ala-NH₂ is interfering with the E2F activity *via* a binding to its DNA recognition domain (Shibata et al., 1998). More recently, a patent claimed the three different short peptides Phe-Trp-Leu-Arg-Phe-Thr or Trp-Val-Arg-Trp-His-Phe and Trp-His-Phe-Ile-Phe-Trp as well

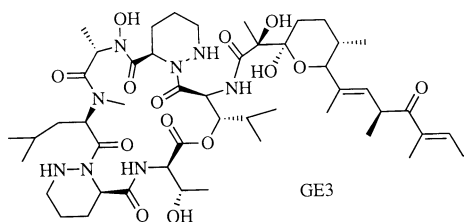


Fig. 6. A naturally occurring antagonist of E2F

as the two longer one Ile-Trp-Leu-Ser-Gly-Leu-Ser-Arg-Gly-Val-Trp-Val-Ser-Phe-Pro and Gly-Ser-Arg-Ile-Leu-Thr-Phe-Arg-Ser-Gly-Ser-Trp-Tyr-Ala-Ser as inhibitors of the E2F-DNA interaction and their use against cell proliferation (Muller et al., 2000). Moreover, it is noteworthy that the c-Myc-derived peptide Asp-Glu-Leu-Lys-Arg-Ala-Phe-Ala-Ala-Leu-Arg-Asp-Gln-Ile linked to an *Antennapedia* internalising sequence inhibits cancer cell growth via a similar mechanism (Draeger and Mullen, 1994; Giorello et al., 1998).

III.5 Src homology domains 2 and 3 and Bcl homology domain 3

Src homology 2 domains (SH2) are 100 amino acid-long sequences that bind phosphotyrosine-containing motifs in a sequence-specific manner. These SH2 domains were first found in the src family of cytoplasmic protein-kinases and exist in many adaptor proteins involved in intracellular signal transduction (Helmreich, 2001; Pawson and Schlessinger, 1993). On the other hand, the src homology domain 3 (SH3) are 55–70 amino acid-long sequences, that bind proline-rich peptides, that are also found in many proteins (Pawson and Schlessinger, 1993). Also, some or all of the four different Bcl homology domains (including Bcl homology domain 3 (BH3)) can be found in a still growing family of proteins involved in the regulation of apoptosis (Adams and Cory, 1998; Chao and Korsmeyer, 1998). Amongst the cancer-related signalling proteins, many interact via such domains. Recent reviews describe the potential targets and the compounds found (Sawyer, 1998; Shakespeare, 2001). But for few other cases (Bardelli et al., 1997; Bardelli et al., 1999; Kardinal et al., 2000), three type of proteins (Grb2, Src and Bcl2) are the main anticancer targets of research groups aiming at finding specific inhibitors of their SH2, SH3 or BH3-mediated interaction with other proteins.

– Grb2 (growth factor receptor-bound protein 2) is the most studied example. This is an adaptor protein involved, for example, in the Ras signalling pathway (Garbay et al., 2000). Inside the cell, the signal beginning from a tyrosine kinase receptor which, when phosphorylated, will bind to Grb2 via its SH2 domain. This protein will then bind, via its two SH3 domains, to Sos (son of sevenless) which in turns binds RasGDP. When this system loses some of its control, it leads to a cellular proliferation and differentiation. Thus, as for the interaction between Ras and Raf, inhibition of the tyrosine kinase-Grb2 or the Sos-Grb2 interactions might lead to antitumor agents, provided that the control loss has taken place upstream to the targeted interaction. Important effort have been made to find strong and selective Grb2 SH2-based ligands (Ettmayer et al., 1999; Garbay et al., 2000; Gay et al., 1999; Hart et al., 1999; Liu et al., 2000a; Yao et al., 1999). One of the problem is the apparent necessity to keep a phosphate group to have a strong *ex vivo* affinity although this very phosphate leads to compounds with a poor activity on cellular assays. One way to overcome this problem was to prepare prodrugs of these phosphate-containing compounds (Gay et al., 1999; Liu et al., 2000b). Another, to prepare phosphonate-derived compound such as the pentapeptide depicted in figure 7 which is, remarkably, a tubulogenesis inhibitor (Battistini et al., 1997). In a different approach, phosphorus-free ligands were designed (Fretz et al., 2000; Hart et al., 1999; Yao et al., 1999). In one instance, this last avenue was opened by a peptidic sequence, found by phage display technique (Oligino et al., 1997), and thus unrelated to the known Shc (another adaptor protein) binding motif pTyr-Val-Asn-Val for Grb2 (Comoglio and Ponzetto, 1995). Further synthetic refinement of the lead compound yielded a cyclic structure (Fig. 7), with submicromolar affinity for Grb2 SH2 domain, in which the γ -carboxyglutamate residue compensates for the lack of phosphate (Long et al., 1999; Lung et al., 2001). It is noteworthy that the same research group is now focusing on Grb7, another adapter-type signalling protein (Han et al., 2001), and has reported the discovery of Grb7 SH2-based peptidic ligands (Pero et al., 2002).

In an other approach, based on X-ray derived structures, very strong Grb2 SH3-based ligands were found

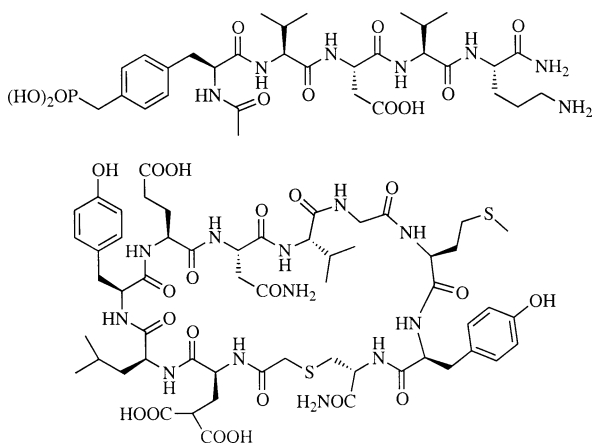


Fig. 7. A phosphonated and a phosphorus-free SH2-based ligand of Grb2

(Cussac et al., 1999; Nguyen et al., 1998). Attachment of one of these ligand (the “peptimer” [Val-Pro-Pro-Pro-Val-Pro-Pro-Arg-Arg-Arg]₂N α ,N ϵ Lys) to the *Antennapedia* cell internalising sequence enable to prove that an inhibition of NIH3T3 colonies formation could be achieved at micromolar concentration (Cussac et al., 1999; Garbay et al., 2000).

- The pp60^{c-src} (Src) protein is a nonreceptor tyrosine kinase containing an SH2 and an SH3 domain. This protein is associated with breast cancer (Luttrell et al., 1994). Accordingly research groups focused on the design of SH2-based ligand to Src in attempts to block its interaction with the protein involved in subsequent cellular proliferation (Gilmer et al., 1994; Lunney et al., 1997; Plummer et al., 1997; Waksman et al., 1993). Some very simple phosphorylated ligands, such as Ac(p)Tyr-Glu-Glu-Ile-NH₂, were found. Again a lack of *in vivo* activity is due to the necessary presence of a phosphate group on the tyrosine residue. This group is either the cause of a poor intracellular penetration or is hydrolysed inside the cell. Hopefully further research will lead to (probably) nonpeptidic analogues which will enable a proof of principle (Buchanan et al., 1999a; Buchanan et al., 1999b; Buchanan et al., 1999c; Pacofsky et al., 1998).
- Bcl2 and related cytoplasmic protein such as Bak, Bax and Bad play a central role in the regulation of apoptosis as “arbiters of cell survival” (Adams and Cory, 1998; Gross et al., 1999). A Bcl2 gene overexpression is observed in a wide number of cancers and this level of expression does correlate with cancer chemotherapeutic or radiation

resistances (Huang et al., 2000). As BH3-based heterodimerizations between Bcl2 and the apoptose-promoting proteins Bax, Bak, Bid and Bad were demonstrated (Chao and Korsmeyer, 1998; Diaz et al., 1997; Sattler et al., 1997), this became a target for anticancer research (Huang, 2000). Three different 16-mer peptides, mimicking the BH3 domain of Bax (Lys-Lys-Leu-Ser-Glu-Cys-Leu-Lys-Arg-Ile-Gly-Asp-Glu-Leu-Asp-Ser) Bak (Gly-Gln-Val-Gly-Arg-Gln-Leu-Ala-Ile-Ile-Gly-Asp-Asp-Ile-Asn-Arg) or Bid (Arg-Asn-Ile-Ala-Arg-His-Leu-Ala-Gln-Val-Gly-Asp-Ser-Met-Asp-Arg), were shown to bind the antiapoptotic Bcl2 and trigger apoptosis in a cell-free system (Cosulich et al., 1997). A peptide made of an *Antennapedia* internalisation sequence and a segment of Bak was proven to trigger apoptosis (Holinger et al., 1999). Moreover, a second proof of principle was achieved with a Bad-derived 25-mer peptide (Lys-Asn-Leu-Trp-Ala-Ala-Gln-Arg-Tyr-Gly-Arg-Glu-Leu-Arg-Arg-Met-Ser-Asp-Glu-Phe-Glu-Gly-Ser-Phe-Lys-Gly-Leu) which was acetylated with decanoic acid to increase its cell-membrane permeability. The pertinence of the target was then established on animal as mice, treated with this modified peptide, survived much longer than untreated mice (Wang et al., 2000). Interestingly a similar approach was undertaken and an “Antennapedia-derived” Bad 21-mer was shown to cause apoptosis but, independently of the Bcl-2 pathway (Schimmer et al., 2001). Current investigations still focus on peptides mimicking the BH3 domain of Bak (Finnegan et al., 2001) as well as on non-peptidic molecules binding the BH3 domain of Bcl2 (Enyedy et al., 2001).

III.6 Cell adhesion proteins binding agents, antimetastatics/antiangiogenics

This part concerns cell adhesion inhibitors that alter the properties of extracellular matrix proteins which play a central role in metastasis and angiogenesis. Cell adhesion proteins can specifically bind to peptidic motifs such as Arg-Gly-Asp (RGD) or Tyr-Ile-Gly-Ser-Arg (YIGSR) or other so far less defined. These are actually the anchoring zones of all the proteins involved in cellular adhesion (i.e.: thrombin B-chain or fibronectin contain this RGD sequence and YIGSR is found in laminin B1 chain). Many ligands analogues of the peptidic RGD or YIGSR motifs were made

(D'Souza et al., 1991) and their antimetastatic effect measured, as it was recognised (Humphries et al., 1986; Iwamoto et al., 1987; Terranova et al., 1984) that they were the underlying bases for cellular migration or metastasis. Further work on RGD-based peptides pointed out the importance to, out of the many integrins existing (Hynes, 1992), focus on selective antagonists for the $\alpha_v\beta_3$ integrin (Brooks et al., 1994; Haubner et al., 1987; Montgomery et al., 1994; Ruoslahti and Reed, 1994). Mentioning all the peptidic analogues would be beyond the scope of this review (Craig et al., 1995). One of the achievement of this avenue of research is the cyclic pentapeptide cilengitide (Fig. 8) which is currently undergoing clinical studies as an antiangiogenic drug (Dechantsreiter et al., 1999; Sorbera et al., 2001). Moreover, one recent report mentions the interesting fact that RGD-derived peptides may also induce apoptosis via a direct caspase-3 activation (Buckley et al., 1999). The clinical efficacy of $\alpha_v\beta_3$ antagonists (peptidic or not) is today the last step remaining to be demonstrated for this antitumor approach (Coleman and Le, 2002).

Comparatively less work was done on the laminin binding site Tyr-Ile-Gly-Ser-Arg, maybe because of its longer length. Here is a list of compounds made which illustrates some of the efforts deployed to transform such motif into a usable drug: Cys-Asp-Pro-Gly-Tyr-Ile-Gly-Ser-Arg-NH₂ (Martin et al., 1989); Ac-Tyr-Ile-Gly-Ser-Arg-NH₂ (Schasteen, 1991); Ac-Tyr-Ile-Gly-Ser-Arg-NHCH₃ (McKelvey et al., 1991); Ac-Tyr-Ile-Gly-Ser-Arg-N(CH₃)₂ (Mori et al., 1994); Phe(pNH₂)-Ile-Gly-Ser-Arg-NH₂ (Zhao et al., 1994); Ac-Tyr-Ile-Gly-Ser-Arg-NHCH(CH₃)₂ (Mori et al., 1995b); CO(Asp-Tyr-Ile-Gly-Ser-Arg-NHPr)₂ (Mori et al., 1995a).

Other peptides, with sequences differing from RGD or YIGSR were also found to be antimetastatic. A recent paper describes efforts to find such sequence that led for instance to the disulfide-bridged Gys-Trp-Asp-Asp-Gly-Trp-Leu-Cys, via a phage display library (Pasqualini et al., 1995).

The laminin receptor-derived compound Ile-Pro-Cys-Asn-Asn-Lys-Gly-Ala-His-Ser-Val-Gly-Leu-Met-Trp-Trp-Met-Leu-Ala-Arg was reported to inhibit cancer cell attachment to endothelium (Castronovo et al., 1991; Magnifico et al., 1996). Moreover, as a study demonstrated that there are many peptide fragments of laminin that have an affinity for cells, it is likely that some new antimetastatic

targets may be better defined in the future (Nomizu et al., 1997).

The tumstatin fragment Gln-Arg-Phe-Thr-Thr-Met-Pro-Phe-Leu-Phe-Cys-Asn-Val-Asn-Asp-Val-Cys-Asn-Phe and longer one containing this sequence are binding $\alpha_v\beta_3$ integrin and have an antiangiogenic potential (Maeshima et al., 2001).

Another phage display-based search, focusing on angiogenic vessels, led to the pentadecapeptide Ala-Ser-Ser-Ser-Tyr-Pro-Leu-Ile-His-Trp-Arg-Pro-Trp-Ala-Arg, which significantly suppress tumour growth. A noteworthy features is that fragment peptides containing the motif Trp-Arg-Pro retained some activity (Asai et al., 2002).

Thrombospondin-1 mimics. Peptides and analogues bearing the type I repeat (Lys-Arg-Phe-Lys-Gln-Asp-Gly-Gly-Trp-Ser-His-Trp-Ser-Pro-Trp-Ser-Ser-Cys) of the extracellular matrix glycoprotein thrombospondin-1, which is involved in antiangiogenesis, have been reported for antiproliferative and antitumor activities (Guo et al., 1997). Further research, based on D-amino acid substitutions, notably led to the heptapeptide Ac-Gly-Val-D-Ile-Thr-Arg-Ile-Arg-NHEt which also corresponds to a segment of the type I repeat sequence but one following the 18-mer described above (Dawson et al., 1999). Moreover, the sequence Ac-Gly-Val-D-Ile-Thr-Nva-Ile-Arg-Pro-NHEt was reported for its antitumor properties (Reiher et al., 2002) and related peptides such as Ac-Sar-Gly-Lys(Ac)-D-Leu-Thr-Nva-Ile-Arg-Pro-NHEt and Ac-Sar-Gly-Val-D-Ile-Thr-NMeNva-Ile-Arg-Pro-NHEt were patented (Haviv et al., 2001a; Haviv et al., 2001b).

Angiostatin analogues. Following the discovery of angiostatin, a naturally occurring fragment of plasminogen, some of its krings domains were demonstrated to be important (Cao et al., 1997; Cao et al., 1996). However, much shorter peptides, derived from the kringle domains or present in the sequence of endostatin, were found quite active and were patented. The following peptides, "bracketed" by prolines residues, are amongst the noteworthy compounds: Ser-Pro-His-Arg-Pro-Arg-Phe-Ser-Pro-Ala; Ser-Pro-His-Ala-His-Gly-Tyr-Ile-Pro-Ser and Thr-Pro-His-Thr-His-Asn-Arg-Thr-Pro-Glu or Thr-Pro-His-Arg-His-Gln-Lys-Thr-Pro-Glu and Glu-Pro-His-Arg-His-Ser-Ile-Phe-Thr-Pro-Glu (Ge and Kini, 2001). These prolines residues may actually provide an inherent structural requirement since such residues are quite frequently found near motifs responsible

for a protein-protein interaction (Kini and Evans, 1995).

Cadherins His-Ala-Val – derived motifs. Short cyclic peptides, containing the highly conserved cadherins motif His-Ala-Val, such as Ac-Cys-His-Ala-Val-Cys-NH₂ were recently patented for their angiogenesis inhibition potential (Blaschuk et al., 2002). This work probably opens a new avenue for this approach as cadherins are calcium-binding membrane proteins enabling cell processes such as segregation and regulation (Williams et al., 2000).

It is quite possible that the mechanism of action of some of the following peptides, reported for their antimetastatic potential, has nothing to do with an interaction with extra cellular matrix proteins: a fragment of the high mobility group 17 Ala-Glu-Asp-Gly-Asp-Ala-Lys-Thr-Asp-Glu-Ala-Gln-Lys-Ala-Glu-Gly-Ala-Gly-Asp-Ala-Lys (Akedo et al., 1991; Isoai et al., 1992); analogues of Glu-Ile-Leu-Asp-Val containing D amino-acids (Kaneda et al., 1997) or polyethyleneglycol derivatives of such motif (Kawasaki et al., 1996); kininogen L fragments and peptides containing Trp-Gly-His-Glu-Lys-Gln-Arg or Lys-Gly-Lys-Lys-Asn-Gly-Lys-His (Matsuda et al., 1996); thrombin fragment Tyr-Pro-Pro-Trp-Asn-Lys-Asn-Phe-Thr-Glu-Asn-Asp-Leu-Leu or shorter one (Packard, 1987). Moreover, a quite simple tripeptide (Fig. 8), of natural origin, was claimed to have a metastasis suppressing activity (Terano et al., 1986).

III.7 Antimitotic peptides, tubulin or actin interfering compounds

Microtubules and actin filaments are the cytoskeletal protein polymers involved in cell growth and division. The polymerisation of $\alpha\beta$ -tubulin dimers in microtubules is the target of many important antitumor drugs. It is the drug-induced alteration of tubulin polymerisation or depolymerisation dynamics which is hampering the correct occurrence of microtubules and is the underlying mechanism of the antitumor action

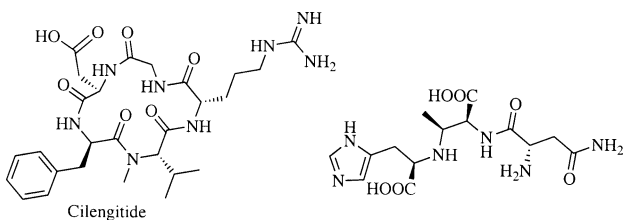


Fig. 8. Cilengitide and a naturally occurring metastasis inhibitor

(Correia, 1991; Jordan and Wilson, 1998). Moreover, other compounds owe their cytotoxicity to the disorganisation of actin filaments that are made from the polymerisation of actin monomers. We chose to class these two types of compounds as inhibitors of protein-protein interactions albeit they are multimeric interactions in the present cases. In the following are actually listed a great number of antimitotics of natural origin (Hamel, 1992; von Angerer, 2000).

III.7.1 Tubulin-interacting peptides. The weakly cytotoxic phomopsisin A (Fig. 9) is isolated from the fungus *Phomopsis leptostromiformi* and act at the level of tubulin (Hamel, 1992; Lacey et al., 1987). It is noteworthy that the related ustiloxins A–D (Fig. 9) were extracted from the pathogen *Ustilagoidea virens* and are also inhibitors of tubulin polymerisation (Li et al., 1995b; Ludueña et al., 1994).

The discovery of the cytotoxic 16-membered cyclic epoxyde-containing cryptophycins was reviewed recently (Corbett et al., 1997). They were isolated from blue-green algae and found to be antimitotic and act on tubulin functions (Mooberry et al., 1996; Moore et al., 1996; Smith and Zhang, 1996). A review of analogues of this structure has been published (Moore et al., 1996) and more compounds have been reported since (Georg et al., 1998; Norman and Shih, 1998; Patel et al., 1999; Patel et al., 1998; Shih et al., 1999; Shih and Williams, 1998; White et al., 1999). The arenastatins (Fig. 10), which have very similar structure, were isolated from the marine sponge *Dysidea arenaria* and were also leads for structure-activity studies (Kitagawa and Kobayashi, 1996; Kobayashi et al., 1994b; Kobayashi et al., 1995). The hindered synthetic dimethyl analogue cryptophycin 52 (Fig. 10) was found more stable toward hydrolysis and is the main candidate for a clinical development (Eggen and Georg, 2002).

A series of cytotoxic tryptophan-derived compounds have been identified so far in four sponge

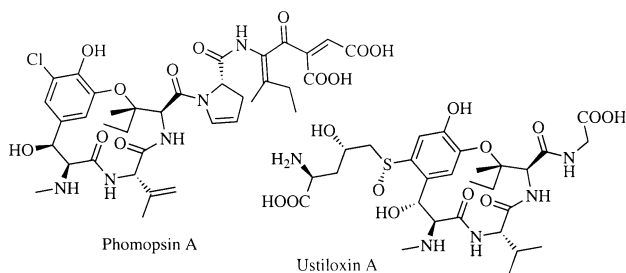


Fig. 9. Structures of phomopsisin A and ustiloxin A

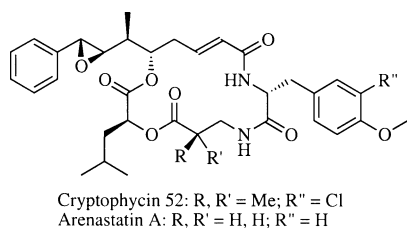


Fig. 10. Structures of cryptophycin 52 and arenastatin A

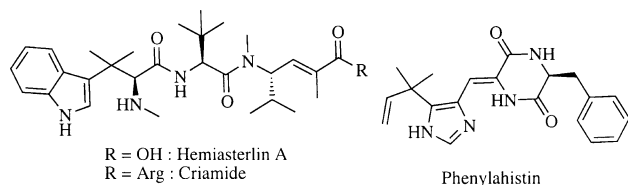


Fig. 11. Hemiasterlin A, criamide and phenylahistin

genera (Gamble et al., 1999). Hemiasterlin A (Fig. 11) was isolated from *Hemiasterella minor* and, along with arginine amides criamide A, from *Cymbastela* (Andersen et al., 1997; Coleman et al., 1995; Talpir et al., 1994) and was found to act via inhibition of microtubule dynamics (Anderson et al., 1997; Bai et al., 1999). It is noteworthy that the more rigid milnamide, which has a tetrahydropyridoindeole ("Tpi"; see Fig. 2) instead of the trimethyltryptophan moiety of hemiasterlin, is also cytotoxic (Crews et al., 1994). Moreover an analogue in which this indole is replaced by a phenyl is more active than the hemiasterlins (Andersen et al., 1999).

Phenylahistin (Fig. 11), which is barely a peptide, is a cytotoxic ketopiperazine derivative (Hayashi et al., 2000; Kanoh et al., 1999a; Kanoh et al., 1999c) that also inhibits tubulin polymerisation (Kanoh et al., 1999b). A remarkable number of other ketopiperazine derivatives such as tryprostatins (Cui et al., 1996a; Cui et al., 1995; Cui et al., 1996b; Cui et al., 1996c) have been reported for their cell cycle inhibition properties and were subsequently shown to inhibit microtubule assembly (Sanz-Cervera et al., 2000; Usui et al., 1998; Zhao et al., 2002). One can wonder at the mechanism of action of the many related compounds (Graz et al., 2000; Milne et al., 1998; Schiavi et al., 2002).

Two reviews report the nearly 25 year-long story of the isolation, identification and synthesis of the many cytotoxic substances found in the sea here *Dolabella* and named dolastatins (Pettit, 1997; Poncet, 1999). Actually, most of these metabolites are probably originated in the cyanobacteria, such as *Symploca*, that this

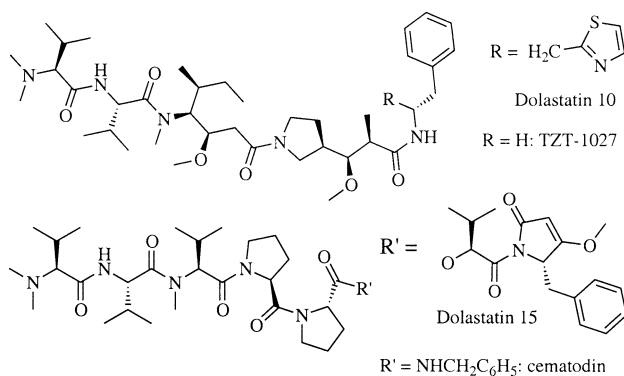


Fig. 12. Dolastatin 10 and 15 and two analogues

herbivorous sea hare ingests (Luesch et al., 2001a). Among them, the linear peptides dolastatin 10 (Pettit et al., 1987) and dolastatin 15 (Pettit et al., 1989a) were found to act on tubulin polymerisation. Their structures led to the synthesis of cematodin (de Arruda et al., 1995; Jordan et al., 1998) and TZT-1027 (Kobayashi et al., 1997) which are today, along with dolastatin 10, at different stages of clinical development (Madden et al., 2000; McElroy Jr. et al., 1997; Supko et al., 2000; Vaishampayan et al., 2000; Villalona-Calero et al., 1998). The naturally occurring linear peptides dolastatin C (Sone et al., 1993b), dolastatin H and isodolastatin H (Sone et al., 1996) also displays an N-terminal dimethylamine group. Moreover, the quite unrelated cytotoxic dolastatin 18 (Pettit et al., 1997b) or virenamide C (Carroll et al., 1996b) displays the same C-terminal thiooxazole-containing amide as dolastatin 10. These naturally originated compounds, the more recently isolated methyl homologue of dolastatin 10, symplostatins 1 (Luesch et al., 2001a) and the analogues synthesised, provide fascinating structure-activity relationships which were reviewed recently (Pettit et al., 1998a; Poncet, 1999).

Bicyclic peptides with structures bearing a central indole moiety such as moroidin and celogentin were reported for their tubulin polymerisation inhibition (Kobayashi et al., 2001; Morita et al., 2000). As a conclusion to this section, we should mention the recently isolated tubulysins which will probably be at the source of important structure-activity studies (Hoefle et al., 2002; Sasse et al., 2000). Tubulysin D (Fig. 13) displays a picomolar activity on mammalian cell lines and induces a complete disappearance of the cellular microtubules. One of the remarkable feature of these compounds (apart from a N-terminal tertiary amine

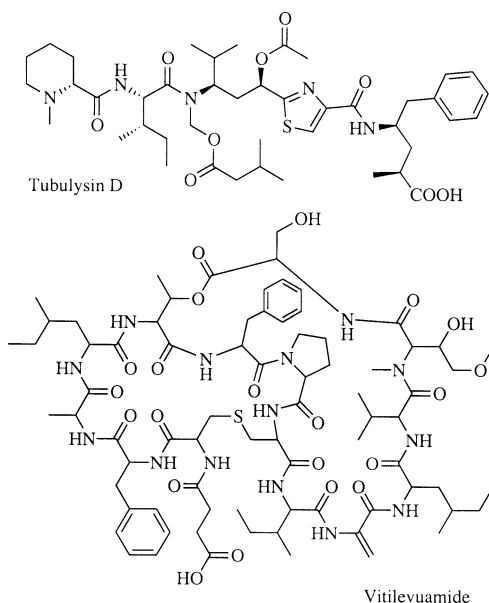


Fig. 13. Tubulyisin D and vitilevuamide

reminiscent of dolastatin 15 analogues (Janssen et al., 1998)) is an acyloxymethyl side chain that could make them actual lipophilic prodrugs, substrates of an *in vivo* hydrolysis. Even more recently, the remarkable bicyclic depsipeptide vitilevuamide (Fig. 13) was isolated from the ascidians *Didemnum cuculliferum* and *Polysyncranton lithostrotum*. The cytotoxicity of this compound is at least partly due to the inhibition of tubulin polymerisation via an interaction at a different site from the binding site of colchicine, the *vinca* alkaloids or dolastatin 10 (Edler et al., 2002).

III.7.2 Actin-interacting peptides. Jasplakinolide (Crews et al., 1986)/jaspamides (Zabriskie et al., 1986; Zampella et al., 1999) and chondamide D are related cytotoxic peptides (Breakman et al., 1987; Crews et al., 1994) that disrupt the proper function of actin (Bubb et al., 1994; Bubb et al., 2000). As depicted on Fig. 14, it is tempting to relate the jasplakinolide/jaspamides to the cytotoxic geodiamolides (Chan et al., 1987; Coleman et al., 1995; Coleman et al., 1999; de Silva et al., 1990). These compounds all display a 12-carbon polypropionate unit. Interestingly, the more structurally different dolicolide also bears a iodotyrosine residue as the geodiamolides (Ishiwata et al., 1994a; Ishiwata et al., 1994b; Ishiwata et al., 1994c) and was recently reported to enhance actin assembly (Bai et al., 2002).

The phallotoxins, such as phalloidin (Fig. 14) are toxic peptides isolated from the poisonous mushroom

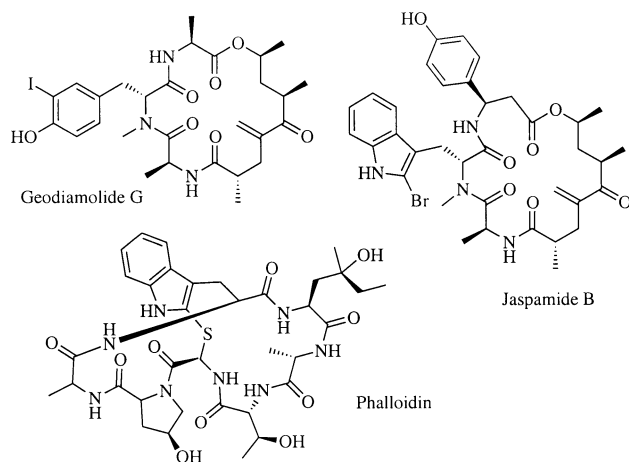
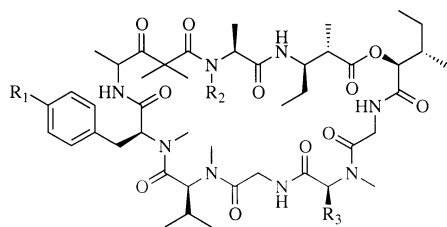


Fig. 14. Geodiamolide G, jaspamide B and phalloidin

Amanita phalloides (Wieland, 1968) that also interact with actin by preventing the depolymerisation of F-actin into G-actin (Miyamoto et al., 1986; Wieland, 1987). The noteworthy 4-hydroxyproline residue was actually found to be essential (Wieland and Faulstich, 1978). Remarkably, the amatoxins which are related to the phallotoxins, are “only” weak inhibitors of RNA polymerase B (Wieland and Faulstich, 1978). A structural common point for an affinity for actin and the inhibition of RNA polymerase remain to be described. This is actually reminiscent of the (non peptidic) etoposides analogues in which some compounds are inhibitors of topoisomerase II function while others are interfering with tubulin. The unrelated antitumor peptidic derivatives cupolamide A and astins, for which no mechanism of action has been suggested, also feature, respectively, a functionalised 4-sulfated or 3,4-bischlorinated proline residues (Bonnington et al., 1997; Morita et al., 1993; Morita et al., 1996c). Moreover 4-hydroxyproline-containing di and tripeptides have been reported for their cytotoxicity (Hall and Chen, 1999).

The total synthesis of dolastatin 11 allowed the elucidation of its mechanism of action (Bates et al., 1997). This compound, and (Fig. 15) the structurally related majusculamide C (Carter et al., 1984), lyngbyastatin and dolastatin 12 (Harrigan et al., 1998; Luesch et al., 1999), act on the cellular actin filament network and it was shown that dolastatin 11 induces hyperpolymerisation of purified actin. Dolastatin 11 is the most cytotoxic of the peptides/depsipeptides that induces the assembly of actin *in vitro* (Bai et al., 2001). However, pharmaceutical development of this class of



Dolastatin 11: $R_1 = \text{OMe}$, $R_2 = \text{H}$, $R_3 = \text{CH}_2\text{iPr}$
 Dolastatin 12: $R_1 = \text{H}$, $R_2 = \text{CH}_3$, $R_3 = \text{CH}_2\text{iPr}$
 Lyngbyastatin 1: $R_1 = \text{OMe}$, $R_2 = \text{CH}_3$, $R_3 = \text{CH}_2\text{iPr}$
 Majusculamide C: $R_1 = \text{OMe}$, $R_2 = \text{H}$, $R_3 = (S) \text{CH}_2\text{Bu}$

Fig. 15. Dolastatin 11 and related substances

compound may be discontinued as a pulmonary toxicity was found for jasplakinolide (Bai et al., 2001; Schindler-Horvat et al., 1998).

III.8 Less investigated examples

III.8.1 Ras-Raf interaction. The activated Raf kinases are the key to a typical cellular signalling pathway to, for instance, cell proliferation via mitogen-activated protein kinases (Helmreich, 2001). Activation of Raf kinase has been shown to be controlled by the interaction with the membrane-translocated protein Ras. *Ex vivo* inhibition of protein Ras and protein Raf interaction has been achieved by peptide sequences found in either Ras or Raf (Barnard et al., 1998; Niehof et al., 1995; Ohnishi et al., 1998). However, even for the short Raf-derived peptide Cys-Cys-Val-Ala-Phe-Arg-Leu, no *in vivo* efficiency of such compound could be demonstrated (Barnard et al., 1998).

III.8.2 Human papillomavirus oncoprotein E6 and E7. Human papillomavirus-associated carcinogenesis is linked to the coexpression of the two viral proteins E6 and E7 in the infected cells. As protein E7 was demonstrated to bind to the retinoblastoma protein, an inhibitor to this interaction could lead to an original therapy for papillomavirus-originated cancer. While a review (Huber et al., 1994) describes the very extensive efforts made to find such a peptide, it is only in a recent article that the hexapeptide Leu-Phe-Tyr-Lys-Lys-Val, actually corresponding to a fragment of the retinoblastoma protein, was reported to be cytotoxic against neoplastic cells containing protein E7 (Radulescu and Jacques, 2000). On the other hand, the E6 protein interacts with the p53 protein resulting in its ubiquitin-dependant degradation. Again, after an unsuccessful attempt to find short peptides (Huber et al., 1994), another approach led to many 20 amino

acids-long peptidic aptamers that binds E6. These peptides were found to cause the exclusive apoptotic elimination of the E6-bearing cancer cells (Butz et al., 2000).

III.8.3 Inhibition of mammalian ribonucleotide reductase assembly. Mammalian ribonucleotide reductase is a target for antitumor treatment (Szekeres et al., 1997). A new avenue could exist in peptide-based inhibitors as it was shown that a small compound (AcPhe-Thr-Leu-Asp-Ala-Asp-Phe) corresponding to the C-terminus of R2 subunit of ribonucleotide reductase does inhibit its assembly with the R1 subunit (Fisher et al., 1993). More recent work led to a series of lactam-bridged compounds with even better inhibitory effect (Liehr et al., 1999).

III.8.4 Restoration of Fas-induced apoptosis. An article describes derivatives of the tripeptide Ser-Leu-Val which are able to inhibit an association between Fas, a cell surface receptor, and FAP-1 a protein which is involved in acquisition of a resistance against anti-Fas antibody-mediated apoptosis. Thus such peptide may have the potential to restore the function of Fas (Sawa et al., 1999).

IV Enzymes inhibitors

IV.1 Ras farnesyl transferase

The antitumor potential of such inhibitors is due to the blocking of the carboxy-terminal cysteine prenylation of protein Ras. This prevents its anchorage to the inner side of cell membrane, which is crucial for the cell multiplication signals transmission (Crul et al., 2001). The first cysteine farnesyl transferase inhibitors found were based on the carboxy terminal part of protein Ras. Many peptides mimicking the last four amino acids of Ras with the general formula Cys-A-A-X are thus inhibiting the farnesyl transferase (Perrin et al., 1996). The thiol-free compound depicted on figure 16, is also derived from this CAAX motif and illustrates the intensive structure-activity studies undertaken (Anthony et al., 1996). All these compounds, including the fact that some entered clinical trials, were reviewed (Crul et al., 2001; Leonard and Sebolt-Leopold, 1999; Omer and Kohl, 1997; Perrin et al., 1996; Sebti and Hamilton, 1997). We should mention a second class of peptidic inhibitors (Fig. 16) which were found using peptide libraries. Their mechanism of action, as for other non peptidic compounds found (Aoyama et al., 1998), is based on an interference with

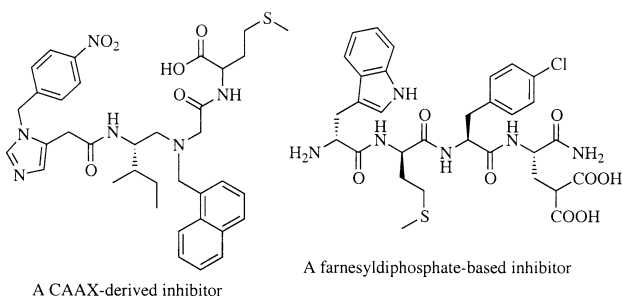


Fig. 16. Example of the two peptidic class of farnesyl transferase inhibitors

the farnesyl diphosphate binding site (Bogusky et al., 1999).

IV.2 Histone deacetylation

Histone deacetylase catalyses the removal of acetyl group from the ϵ -amine of lysine residues present near the amino terminus of nucleosomal histones. Inhibition of this process was shown to be a target for cytotoxic compounds, possibly by the prevention of DNA unwinding from around the histone prior to its transcription (Yoshida et al., 2001). Recent studies point out a much more elaborate role for post-translational modification of histones that would be hindered by deacetylation inhibitors (Strahl and Allis, 2000). Moreover, the histone deacetylase inhibitors effect on the up-regulation of p21^{WAF1/CIP1} was investigated (Burgess et al., 2001).

The two tetrapeptides trapoxins A and B (Fig. 17), containing an epoxyde moiety, were isolated from *Helicoma ambiens* (Itazaki et al., 1990). They are related to the many epoxyde-containing cyclic tetrapeptides known such as Cyl-2 (Hirota et al., 1973), chlamydocin (von Closse and Huguenin, 1974), HC-toxin (Liesch et al., 1982) and WF-3161 (Umehara et al., 1983). They were found to display detransforming activity against *v-sis* oncogene transformed NIH3T3 cells (Itazaki et al., 1990). The first structure-activity studies aiming at improving the poor stability of these compounds showed that the chemically reactive epoxyde moiety could be replaced with other functions, reactive as well (Bernadi et al., 1993; Shute et al., 1987). More recently, analogues, based on trapoxin B structure, not only pointed out the importance of the D-amino acids chirality but also confirmed (Tomizaki et al., 1999) that such compound have a capacity for histone deacetylase inhibition (Kijima et

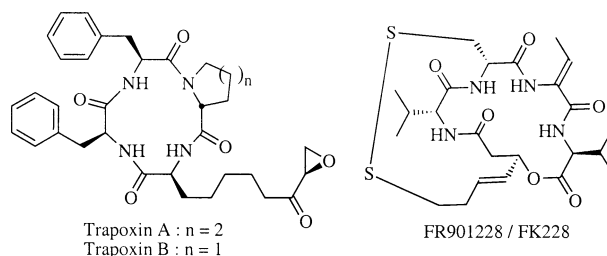


Fig. 17. Structures of trapoxins A, B and R901228 / FK228

al., 1993; Taunton et al., 1996). Recent work showed that the keto-epoxyde group could be replaced by a hydroxamate function (Komatsu et al., 2001; Yoshida et al., 2001). The related cyclic peptide such as apicidin (Singh et al., 1996) and diheteropeptin (Masuoka et al., 2000) display somewhat less reactive side chains (an N-methoxy indole or a diol) and retain biological activities. Moreover, a recent patent describes very active compounds only bearing an α -hydroxy ketone-containing side chain (Mori et al., 2000).

The antitumor peptide FR901228 (or FK 228) (Fig. 17) was isolated from *Chromobacterium violaceum* (Ueda et al., 1994a; Ueda et al., 1994b) and was later found to be also an inhibitor of histone deacetylase (Nakajima et al., 1998). Its pharmacological (Chan et al., 1997) and antiangiogenic (Kwon et al., 2002) properties made it a good candidate for clinical development (Piekarz et al., 2001). It is noteworthy that the less functionalised spiruchostatins were reported for a gene expression enhancement (Masuoka et al., 2001). Hopefully, analogue synthesis will enlighten the structure-activity relationship of this type of compounds that seem to have an effect on cellular signalling pathways (Yu et al., 2002).

Moreover, lunasin a remarkable soybean-extracted 43-mer peptide, or fragments as short as the 17-mer Cys-Glu-Lys-His-Ile-Met-Glu-Lys-Ile-Gln-Gly-Arg-Gly-Asp-Asp-Asp-Asp, were recently found to bind deacetylated histones and thus block cell proliferation by preventing their acetylation (Galvez, 2002; Galvez et al., 2001).

IV.3 Inhibitors of proteasome

The proteasome is an intracellular proteolytic system which displays the peptidase profile of chymotrypsin, trypsin and peptidylglutamyl-peptide protease. This system, in conjunction with ubiquitin, controls the level of many proteins involved in cell proliferation

(see above the case of p53). Potent inhibitors were isolated or prepared such as lactacystin (Fig. 18) (Corey and Wei-Dong, 1999; Omura et al., 1991a; Omura et al., 1991b), epoxyde-containing peptides (Elofson et al., 1999; Koguchi et al., 2000; Sin et al., 1999), the remarkable belactosin A (Fig. 18) or corresponding lipophilic prodrugs (Asai et al., 2000; Yamaguchi et al., 2000) and boron-derived peptides such as PS-341 (Fig. 18) (Adams et al., 1999). From the much more complicated structure of TMC-95A (Kohno et al., 2000), simpler compounds were recently reported to retain an inhibitory action (Kaiser et al., 2002). These inhibitors actually make remarkable tools to elucidate apoptotic processes (Delic et al., 1998; Gazos Lopes et al., 1997; You et al., 1999). Moreover, PS-341 is currently undergoing preclinical investigations (Teicher et al., 1999).

IV.4 Protein phosphatases

Interest in protein phosphatases inhibitor research (Sheppeck et al., 1997) for anticancer treatment could have been renewed by the fact that the strongly cytotoxic naturally-occurring motuporin (Fig. 19) was found to be one of the most potent inhibitor of protein phosphatase 1 (de Silva et al., 1992). The substituted phenyldecadienoic acid chain is also found in other naturally occurring phosphatase inhibiting peptides such as nodularin and microcystins (Goldberg et al., 1995; Namikoshi et al., 1992). This led to structure-activity investigations which showed that the dehydrobutyryl residue of motuporin was, surprisingly (Goldberg et al., 1995), non essential (Samy et al., 1999). More synthetic work was done on microcystin in order to improve the selectivity of its protein phosphatase inhibition (Aggen et al., 1999).

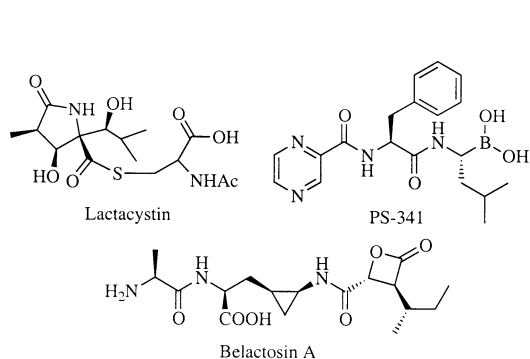


Fig. 18. Structures of lactacystin, PS-341 and belactosin A

IV.5 Ribosome interacting compounds

Two classes of ribosome-interacting compounds have been extensively studied.

- The antitumor properties of the many derivatives isolated from *Rubia akane*, such as RA-VII, and the related bouvardin (Fig. 20), were reviewed (Itokawa and Takeya, 1993). Their main mechanism of action was shown to be an inhibition of protein synthesis *via* a binding to eukaryotic ribosomes (Sirdeshpande and Toogood, 1995; Zalacain et al., 1982). However an effect on cyclin D1 protein level was reported recently for RA-VII (Wakita et al., 2001). Current research are focused on, for example, hemisynthetic dialkylaminated derivatives which display a higher water solubility (Hitotsuyanagi et al., 1997).
- The highly cytotoxic didemnin derivatives A-E, characterised by a 25-membered cyclic peptide, were found in the marine ascidian *Didemnum molle* (Rinehart Jr. et al., 1981a; Rinehart Jr. et al., 1981b; Rinehart Jr. et al., 1988; Toske and Fenical, 1995). This discovery was followed by much structure activity studies (Jouin et al., 1991) that were reviewed (Schmidt et al., 1999; Vera and Joullié, 2002). The isolation of the related cytotoxic

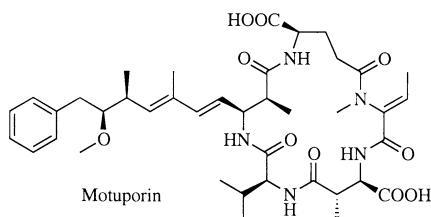


Fig. 19. Structure of motuporin

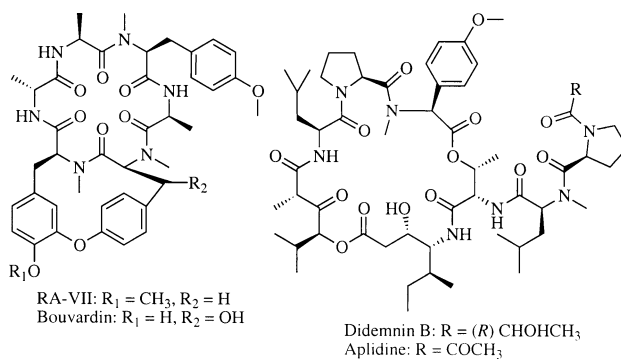


Fig. 20. Structure of bouvardin, RA-VII and didemnin B

peptides tamandarins were recently reported (Vervoort and Fenical, 2000). The mechanism of action of didemnins was shown to be based on the inhibition of protein synthesis (Ahuja et al., 2000). Concerning didemnin B, if the preclinical results were encouraging (Jiang et al., 1983), the poor results obtained in the course of phase II trials of didemnin B (Fig. 20) do not warrant any further clinical development of this compound (Hochster et al., 1999; Maroun et al., 1998; Mittelman et al., 1999; Sondak et al., 1994; Taylor et al., 1999). On the other hand, aplidine is still undergoing clinical trials (Jimeno, 2002; Nuijen et al., 1999; Raymond et al., 2000).

IV.6 Less investigated examples

IV.6.1 Inhibitors of proteases. There are four major classes of protease enzymes (aspartic, serine, cysteine and metallo) and peptidic inhibitors for these enzymes have been the subject of much research (Leung et al., 2000). On the oncology point of view, urokinase form of plasminogen activator (uPA), cathepsin B and D and various metalloproteases are involved in the metastasis process (Duffy, 1992).

Many non peptidic inhibitors of metalloproteases were found and some are undergoing clinical trials (Brown, 1999; De et al., 1999; Johnson et al., 1998; Whittaker et al., 1999). Recent work describes a phage display-based research which led to the disulfide-bridged cyclic Cys-Thr-Thr-His-Trp-Gly-Phe-Thr-Leu-Cys, a specific inhibitor of the matrix metalloproteinase 2 (gelatinase A) (Koivunen et al., 1999; Medina et al., 2001).

A typical example of the problematic encountered with the other proteases could be the *Actinomyces*-extracted tripeptide aldehyde leupeptin (Ac-Leu-Leu-ArgCHO) which is an inhibitor of carcinogenesis (Hozumi et al., 1972). This compound was shown to be a good inhibitor of various peptidases such as the serine proteases trypsin, plasmin and papain; the cysteine protease cathepsin B and the aspartic protease cathepsin D (Aoyagi et al., 1977). However even if all these proteases are potential anticancer targets, one of the remaining problem is obtaining a more selective inhibition, for example between serine and cysteine proteases (Leung et al., 2000).

– Recent studies focus on finding specific inhibitors of the serine protease uPA and the mechanism-based

peptidic inhibitors were reviewed recently (Tamura et al., 2000). Moreover, a recent article describes the synthesis of peptidic derivatives aiming at the inhibition of the serine protease plasmin (Abato et al., 2002).

- Many irreversible inhibitors of cysteine protease were also reviewed recently (Otto and Schirmeister, 1997). Inhibition of the cysteine protease, involved in the conversion of interleukin-1 β , could be beneficial in tumour treatment (Watanabe et al., 1998). Moreover, the peptidic derivative Z-Phe-Gly-NHO-Bz is an inhibitor of the cysteine protease cathepsins and induces apoptosis in human cancer cells (Zhu and Uckun, 2000).
- Relatively few inhibitors of the lysosomal aspartic protease cathepsin D have been synthesised and studied for their antitumor potential (Bessodes et al., 1999). A recent review describes a possible role for this protease as it is, unexpectedly, regulated by estrogens in breast cancer cell lines (Rochefort and Liaudet-Coopman, 1999).

IV.6.2 Glyoxalase inhibitors. Glyoxalase inhibitors research has been quite important and was reviewed (Creighton et al., 2000). However, a recent report does question the viability of glyoxalase as an anticancer target (Tew, 2000). Few peptides, derived from glutathione, were reported for their inhibition properties of glyoxalase (Kavarana et al., 1999; Vince et al., 1999).

V Nucleic acid-interacting agents

A nucleic acid interaction does not reflect the mechanism of action of these compounds past the fact that they have an affinity for DNA (or RNA). This binding is followed by other events such as chemical destruction of the DNA strand or inhibition of some of the enzymes involved in DNA chemistry (*i.e.*: maintenance and duplication) such as topoisomerases or polymerases. It is likely that some of the inhibitors of E2F/DNA interaction mentioned above could have been placed in this section.

V.1 Bleomycins and actinomycin D

Bleomycins and actinomycin D are highly active DNA-interacting compounds that are used in cancer chemotherapy (Larsen, 1996). Bleomycin A2, the main component of the bleomycins mixture used, is

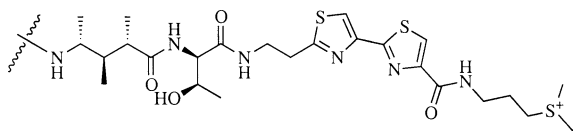


Fig. 21. Bleomycin A2 side chain

isolated from *Streptomyces verticillus*. It is a complex molecule made of a diholoside side chain, a metal chelating central part and a bithiazole-containing peptidic chain (Fig. 21). The latter is responsible for the interaction with DNA minor groove (Manderville et al., 1994). Hydrolysis of its last amide bond leads to an inactive compound (bleomycinic acid). Replacement of this terminal sulfonium part by aminated moiety (bleomycin B2 or peplomycin) restore the activity.

Actinomycin derivatives, which have been reviewed, are composed of a central aminophenoxazone and two cyclic peptidic side chains (for example: Val(NMe)Gly(NMe)-Pro-DVal) (Hollstein, 1974). These compounds were isolated from *Streptomyces* species. They bind to DNA *via* the intercalation of the central aminophenoxazone, and the side chains were shown to interact with the DNA minor groove. The relatively small alteration of the peptides made by the removal of the hydrophobic methyl of valine and glycine actually leads to a complete loss of cytotoxicity (Mosher and Goodman, 1972). It is noteworthy that peptide-anthraquinone derivatives have been prepared more recently and the specificity of their action studied (Ijaz et al., 2001; Takenaka et al., 1996).

V.2 Echinomycin and related compounds

In the last 40 years a remarkable number of related antitumor DNA-interacting (Quigley et al., 1986; Takusagawa, 1985; Waring and Wakelin, 1974; Zhang and Patel, 1991) peptidic compounds were isolated and characterised. They display a general structure made of a central depsipeptide ring flanked by two quinoline or quinoxaline moieties. It is beyond the scope of this review to depict and compare all these structures. Echinomycin was the first to be isolated (Keller-Schierlein et al., 1959) and its proposed structure revised later (Dell et al., 1975). This was followed by triostin (Fig. 22) (Otsuka and Shōji, 1965; Otsuka et al., 1976), the closely related quinomycins (Martin et al., 1975; Williamson et al., 1982), BBM-928 /

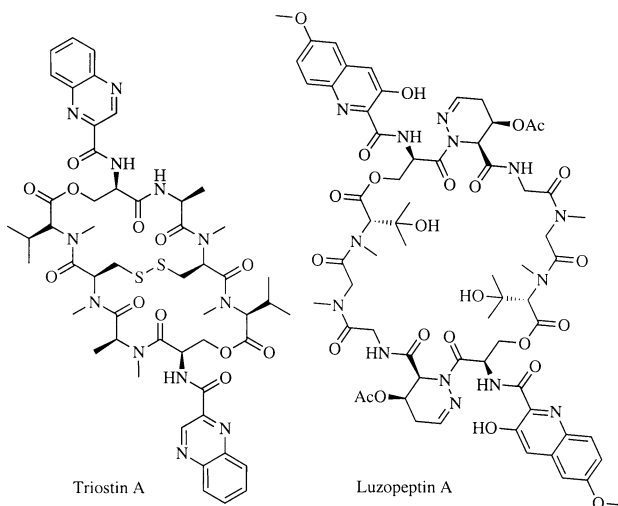


Fig. 22. Structures of triostin A and luzopeptin A

luzopeptin A (Fig. 22) (Arnold and Clardy, 1981; Konishi et al., 1981), quinaldopeptin (Toda et al., 1990), sandramycins (Matson and Bush, 1989; Matson et al., 1993), BE-22179 (Okada et al., 1994), quinoxapeptins (Lingham et al., 1996), thiocoraline (Pérez Baz et al., 1997), korkormicins (Lam et al., 1995) and SW-163C and E (Takahashi et al., 2001). Echinomycin as been the subject of clinical trials, without notable results (Chang et al., 1998; Gradishar et al., 1995; Wadler et al., 1994).

Structure-activity relationship studies were undertaken such as the synthesis of the bis-acridine analogue of des-N-tetramethyltriostin, which retained a high affinity for DNA (Helbecque et al., 1985). They were followed by much more extensive synthetic work on the luzopeptin-derived structures (Boger et al., 1998; Boger and Ichikawa, 2000; Boger et al., 1999; Boger and Saionz, 1999; Lam et al., 1995; Olsen et al., 1986). From them, some fascinating results were found for sandramycin analogues, such as an inhibition concentration for melanomas, carcinomas and adenocarcinomas in the picomolar range (Boger et al., 1998). On the mechanism of action point of view, quite remarkable specific effects were found. The depsipeptide BE-22179 was shown to preferentially inhibit topoisomerase II (Yoshinari et al., 1994), thiocoraline did not inhibit topoisomerases but DNA polymerase α (Erba et al., 1999) and quinoxapeptin did not inhibit any of the polymerase α , β , γ and δ but, as well as luzopeptin A, inhibited HIV-1 and HIV-2 reverse transcriptase (Lingham et al., 1996).

V.3 Other examples

Other examples of peptides that interact with DNA were found. Their structure is based on the sequences of transcriptional activators or other protein interacting with DNA. Remarkably simple peptides were studied such as the heptad repeat of polymerase II (Tyr-Ser-Pro-Thr-Ser-Pro-Ser-Tyr) (Khat et al., 1996; Suzuki, 1990). For what it is worth, statistically-wise, the naturally occurring cytotoxic heptapeptide yunnanin C featuring the motif Tyr-Ser-Pro, was also reported (Morita et al., 1996b). Remarkably, the anticancer peptide X-Met-Leu-Pro-Ser-Tyr-Ser-Pro-Tyr, was identified from a soy proteins extract (Kim et al., 2000). Moreover, the tetrapeptides Ser-Pro-X-X, derived from netropsin, were also studied (Hao et al., 1995).

Other compounds are the 60-mer peptide bZIP motif (Tabor, 1996; Talanian et al., 1992), or the AT-DNA binding motif of mammalian high mobility group I (Thr-Pro-Lys-Arg-Pro-Arg-Gly-Arg-Pro-Lys-Lys) (Reeves and Nissen, 1990). Moreover, peptides comprising the RNA-binding motif Leu-Asp-X-Arg (X = an alkyl residue), involved in polyadenylation, were patented for blocking a specific kinase activity (Richter and Mendez, 2001).

VI Other peptides

VI.1 Linear peptides

A remarkable number of endogenous pyroglutamate-starting growth regulatory peptides, such as pGlu-Glu-Asp-Ser-Gly and its analogues mentioned in the introduction were found (Balazs et al., 1992; Jensen et al., 1990; Laerum, 1990). The other peptides reported for a cellular growth inhibition are: pGlu-Phe-Gly-NH₂ (Gembistky et al., 2000), pGlu-Glu-Gly-Ser-Asp or pGlu-Glu-Gly-Ser-Asn (Paulsen et al., 1992), pGlu-Glu-Asp-Cys-Lys (Foa et al., 1987) or pGlu-His-Gly (Paulsen, 1993). The N-acetylated glutamate derivative Ac-Glu-Ser-Gly-NH₂ has also been found to inhibit lymphocyte growth (Liu et al., 2000c). Moreover the pineal-originated tetrapeptide Ala-Glu-Asp-Gly is reported to have a tumor suppressing effect (Khavinson and Anisimov, 2000). As a proper molecular mechanism of action of this class of peptide remain to be found, it is tempting to mention the dansylated octapeptide Dns-Glu-Asp-Asp-Ser-Asp-Glu-Glu-Asn reported for its antiproliferative action (Marsili et al., 1996). If this sequence is not exactly related to the

above pGlu-starting compounds, it is remarkable that many phosphorylated peptides such as pGlu-Ala-Glu-Ser-Asn or pGlu-Asp-Asp-Ser-Asp-Glu-Asn can bind DNA in the presence of divalent cations (Cardellini et al., 1999; Chillemi et al., 1991). Among the puzzling facts concerning this last octapeptide is that a) the phosphorylated form is able to inhibit RNA polymerase II b) its sequence actually corresponds to the carboxy terminus section of the largest RNA polymerase II subunit c) this sequence actually follows the DNA-interacting heptad repeat of polymerase II mentioned in the previous section (Angiolillo et al., 1993).

- The bone marrow-extracted peptides Ac-Ser-Asp-Lys-Pro and Ser-Asp-Lys-Ac (Lenfant et al., 1989; Ruhestroth-Bauer et al., 1993) or Phe-Arg-Pro-Arg-Ile-Met-Thr-Pro (Mikhailova et al., 1998; Strelkov et al., 2000) were reported for inhibition of cell proliferation. Structure-activity on the tetrapeptide Ac-Ser-Asp-Lys-Pro were undertaken in order to reduce its sensitivity toward proteases and to explore the respective importance of the peptide residues (Gaudron et al., 1999). They pointed out the importance of the central motif Ser-Asp-Lys (Thierry et al., 2001) which had actually been patented for its inhibition of the proliferation of liver cells (Ruhestroth-Bauer, 1994; Ruhestroth-Bauer, 1997).
- Tetrapeptides derived from phytotoxin AS-1, a severe toxin for plant leaves, such as Cys-Val-Gly-Glu; Tyr-Val-Gly-Glu and His-Val-Gly-Glu were cytotoxic to mouse fibroblast L929 (Liakopoulou-Kyriakides et al., 1998).
- Many other simple linear peptides listed here have been isolated or prepared and reported for an anticancer potential: copper complex of Gly-His-Lys (Pickart, 1989); Palmitoyl-Leu-Leu-Arg-OME (Ueda et al., 1986); enkephalin aldehydes H-Tyr-Gly-Gly-Phe-Met (or Val)H and enkephalin analogues (Daiichi and Yakuhin, 1985; Scholar et al., 1987); segments of Asn-Gln-Asn-Gly-Ser-Asn-Pro-Lys-The-Val-Lys-Gln-Ala, isolated from *Papaver somniferum* pollen (Xu and Jin, 1998); Leu-Ile-Glu-Asp-Asn-Glu-Tyr-Thr-Ala-Arg (Sagami, 1984) or the 16-mer sequence, pGlu-Leu-Lys-Cys-Tyr-Thr-Cys-Lys-Glu-Pro-Met-Thr-Ser-Ala-Ala-Cys, obtained from the amino terminal fragment of a urine-extracted antineoplastic protein (Ridge and Sloane, 1996; Sloane, 2002).

– More elaborate linear cytotoxic peptides were also characterised such as the two linear peptides, majusculamide D and deoxymajusculamide D (Fig. 23) (Moore and Entzeroth, 1988). The closely related microcolins were reported for their immunosuppressive properties, and their apoptosis induction (Koehn et al., 1992; Zhang and Longley, 1999). It is noteworthy that a mixture of, at least, 16 very lipophilic peptides named roseoferin was shown to have high antiproliferative effect (Degenkolb et al., 2000). The cytotoxic and quite lipophilic 18-mer AcPhe-Aib-Ala-Aib-Iva-Leu-Gln-Gly-Aib-Aib-Ala-Ala-Aib-Pro-Iva-Aib-Gln-Trp was isolated from *ascomycetes Apioera sp.* (Kim et al., 2002). Moreover adenopeptin, a fairly simple lipid-containing tridecapeptide was shown to induce apoptosis in transformed cells (Hayakawa et al., 1998).

VI.2 Cyclic peptides

The cyclodepsipeptide dolastatin 14 (Fig. 24), containing a 14 carbon long lipophilic hydroxy acid, was isolated from sea hare *Dolabella auricularia* and was found to be very active (Pettit et al., 1990). The lack of available sample is probably the reason for the ab-

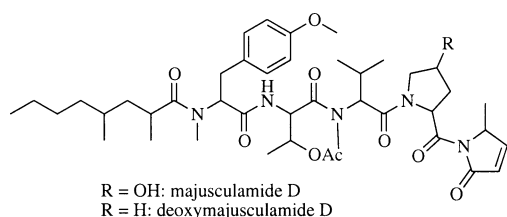


Fig. 23. Majusculamide D and deoxymajusculamide D

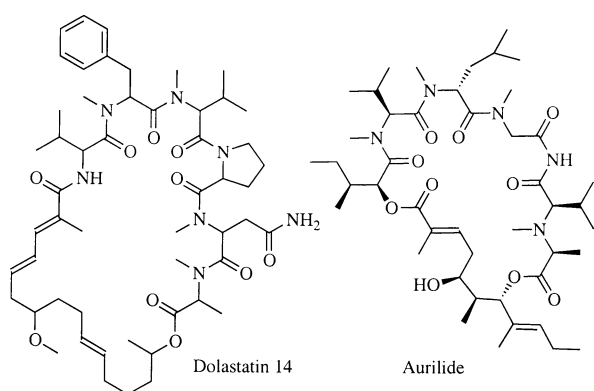


Fig. 24. Dolastatin 14 and aurilide

sence of experimental data on its mechanism of action. The remotely related aurilide bearing another lipophilic hydroxy acid was isolated from *Dolabella auricularia* and a synthetic sample is still very cytotoxic (Mutou et al., 1997). Dolastatin G (Mutou et al., 1996a; Mutou et al., 1996b) and lynchbyastatin 2 are compounds actually sharing some common components with these peptides. However, lynchbyastatin 2 was only found toxic on mice (Luesch et al., 1999).

Papuamides (Fig. 25) are depsipeptides displaying a lipophilic decanedienoic side chain that have been isolated from the sponges *Theonella mirabilis* and *Theonella swinhoei* and display powerful anti HIV and cytotoxic properties (Ford et al., 1999). Although it is 6-hydroxylated, an homoproline residue is also present in a number of much less cytotoxic depsipeptides such as microcystilide (Tsukamoto et al., 1993), dolastatin 13 (Pettit et al., 1989b), symplostatins 2 (Harrigan et al., 1999) or somamides (Nogle et al., 2001).

The strongly cytotoxic tetradecapeptide discodermin E (Fig. 26) was isolated from the marine sponge *Discodermia kiiensis* (Ryu et al., 1994a). Many related compounds have been reported such as: discodermins A-D, F-H (Matsunaga et al., 1984; Matsunaga et al., 1985a; Matsunaga et al., 1985b; Ryu et al., 1994b), polydiscamide A (Gulavita et al., 1992) and halicyclindramide (Li et al., 1995a; Li et al., 1996).

Kalahalide F (Fig. 27), is a potent antitumor cyclic depsipeptide isolated from the sarcoglossan mollusc *Elysia rufescens* and its diet, the green alga *Bryopsis sp.* (Goetz et al., 1999; Hamann and Scheuer, 1993). A recent patent describes the structure-activity studies that led to a lithocholoyl-containing analogue (Fig. 27) which is more potent than kahahalide F (Albericio et al., 2002; López-Macià et al., 2001). Their remark-

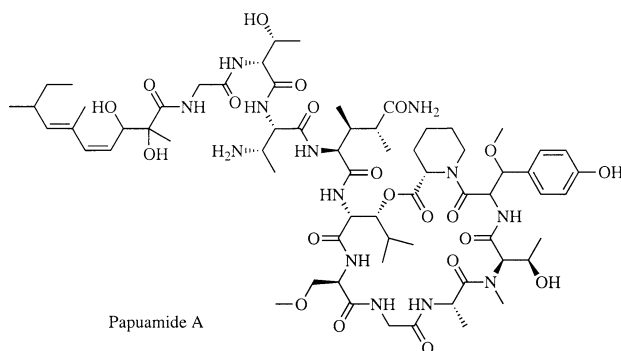


Fig. 25. Papuamide A

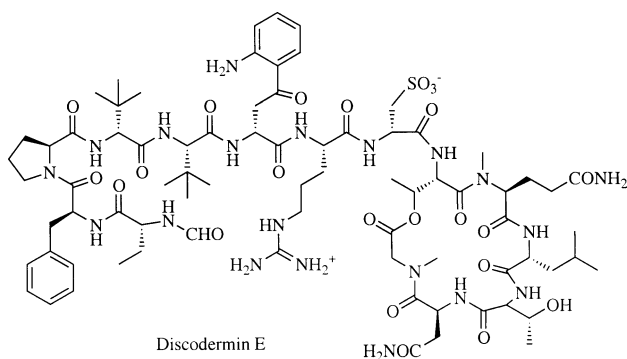


Fig. 26. Structure of discodermin E

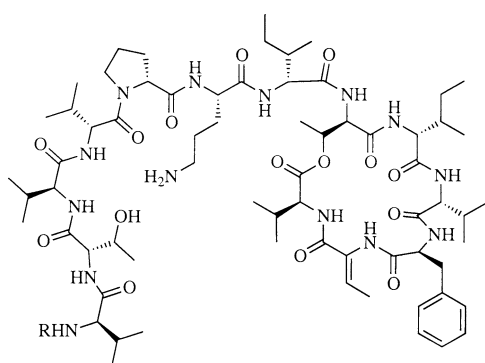


Fig. 27. Kahalalide F and its more potent analogue

able structural similarities with compounds such as discodermin E (Fig. 26) hold the promises of future synthetic studies. A dehydrobutyric residue has also been found in the structures of cytotoxic dolastatin 13 and other related compounds (Pettit et al., 1989b).

Many cytotoxic thiazole-containing cyclopeptidic compounds have been isolated. We will just depict trunkamide A (Caba et al., 2001; Carroll et al., 1996a; Wipf and Uto, 1999) and apratoxin A (Luesch et al., 2002; Luesch et al., 2001c) (Fig. 28) which are notable in terms of cytotoxicity. However, apratoxin A was found only marginally active *in vivo*. Other reported cytotoxic peptides containing thiazoles and/or oxazoles are: ulithiacyclamide (Ireland et al., 1982; Shioiri et al., 1987; Williams et al., 1989), dolastatin 3 (Pettit et al., 1987), ulicyclamide (Kohda et al., 1989), keramamide F (Itagaki et al., 1992), mollamide (Carroll et al., 1994), patellamide 6 (Carroll et al., 1996a), keenamide A (Wesson and Hamann, 1996), lissoclinamide 7 (Hawkins et al., 1990), orbiculamaide

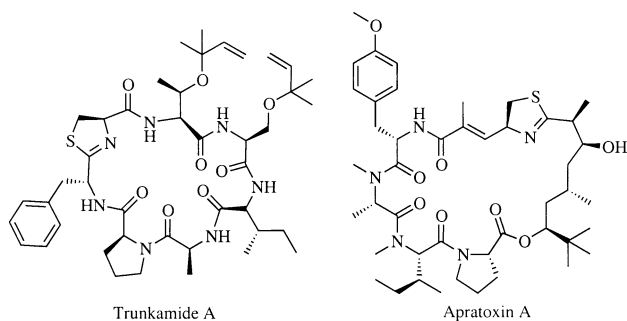


Fig. 28. Trunkamide A and apratoxin A

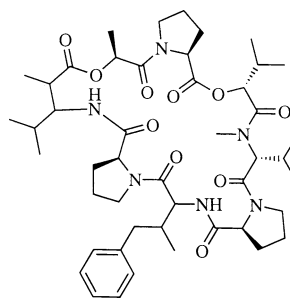


Fig. 29. Dolastatin 16

A (Fusetani et al., 1991), westiellamide (Wipf and Miller, 1992), cyclodidemnamide (Toske and Fenical, 1995), oriamide (Chill et al., 1997), keramamide K (Uemoto et al., 1998), keramamide M and N (Tsuda et al., 1999), microcyclamide (Ishida et al., 2000) and waiakeamide (Mau et al., 1996) or the related haligramides (Rashid et al., 2000).

Dolastatin 16 (Fig. 29) also isolated from *Dolabella auricularia*, but from another part of the sea world, was found very cytotoxic (Pettit et al., 1997a). Other depsipeptides, featuring a β -amino acid, were reported such as cyclic octapeptides made of Asn-Tyr-Asn-Gln-Pro-Asn-Ser and various fatty β -amino acids (Ikegami et al., 1987), dolastatin D (Sone et al., 1993a) and dolastatin 17 (Pettit et al., 1998b). A remarkable series of cyclo- β -tripeptides, bearing lipophilic sides chains, have been recently reported (Gademann and Seebach, 2001). Moreover the cytotoxic kailiuns (Harrigan et al., 1997) and pitipeptolides (Luesch et al., 2001b) display a fatty side chain as well as lipophilic residues. Also reported for their cytotoxicity are the lipophilic cyclic peptides axinellins (Randazzo et al., 1998), cycloleonuripeptides (Morita et al., 1996a), mollamide (Carroll et al., 1994) and onchidins (Fernández et al., 1996).

Many other cytotoxic peptides of natural origin could be depicted here either for their remarkable structure or their antitumor potential. As a chemist we would mention peptides bearing a challenging structure such as the kapakahines (Yeung et al., 1996), the himastatin (Kamenecka and Danishefsky, 1998; Mamber et al., 1994) and the related chloptosin (Umezawa et al., 2000), the theonellamides (Matsunaga and Fusetani, 1995), the aciculitins (Bewley et al., 1996), the microsclerodermins (Qureshi et al., 2000) and the cyclocinamide (Clark et al., 1997; Grieco and Reilly, 1998). Indeed all these cytotoxic peptides bear remarkable structures arising from further chemical reaction on the amino acid residues. Total synthesis of these compounds and their analogues, coupled with proper biological studies of their mechanism of action, could lead to original approaches to antitumor therapy.

VII Conclusions

This review is incomplete in at least two different ways. First, for all the relevant papers that were overlooked we have to apologise. The isolation, characterisation, design, synthesis/biosynthesis and biological testing of these compounds have been a tremendous task, undertook and achieved by many. Second, quite often, either when a common mechanism of action was known or not, some tantalising structural similarities between “unrelated” compounds were noticed. In this regard the present review is seriously faulty since only peptides were reviewed. Only one case of linear similarity such as the one described in the introduction was found in the course of this work (see the end of the nucleic acid-interacting agents section). However, a future achievement will be to be able to pinpoint three dimensional structural similarities for any type of compound and thus maybe, by analogy, suggest a common biochemical mechanism of action.

The remarkable phage-displaying method or the peptide aptamer screening and other related techniques have provided many peptides of potential interest. The most though provoking could be work describing tripeptides (derivatives of Pro-Thr-Trp, Pro-Tyr-Pro-NH₂ or Glu-Arg-Pro), found *via* a screening, which suppress cell proliferation (Hiwasa et al., 1996; Ike et al., 1997). One may forecast for the near future a landslide of new peptidic structures, targeting many original cellular mechanisms, which will have to be chemically “adapted” to the stringent re-

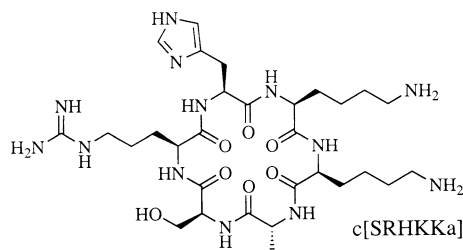


Fig. 30. A man-made activator of p53

quirement of cellular and mammalian pharmacology to yield marketable drugs. This is not a small task as its complexity is “exponentially more laborious with the starting peptide length” (Huber et al., 1994). On the other hand, nature also provides an access to a quite infinite number of secondary metabolites and, as analytical techniques have been improving enormously, it is a noteworthy source of original and already organism-adapted peptidic structures. As a final point to this review, the remarkable cyclic peptide Ser-Arg-His-Lys-Lys-D-Ala (Fig. 30) has been recently patented (Halazonetis and Hartwig, 2001) for its p53 activation properties. The “simplicity” of its structure is in itself an humbling lesson.

Acknowledgements

This work would have not been possible without the kind staff and the resources of the Bibliothèque de la Faculté de Pharmacie de la rue de l'Observatoire, Paris. I also wish to acknowledge Dr. René Pontikis, Dr. Jean-Pierre Buisson and Emmanuel Bouvier for their help in the proof-reading of this manuscript as well as Miles Neave, Callum Robertson and their customers. Moreover, Dr Emile Bisagni must also be mentioned here for many reasons, not the least for having been instrumental in my entrance into the C.N.R.S.

References

- Abato P, Yuen CM, Cubanski JY, Seto CT (2002) Inhibitors of plasmin that extend into both the S and S' binding sites: cooperative interactions between S1 and S2. *J Org Chem* 67: 1184–1191
- Adams JM, Cory S (1998) The Bcl-2 protein family: arbiters of cell survival. *Science* 281: 1322–1326
- Adams J, Palombella VJ, Sausville EA, Johnson J, Destree A, Lazarus DD, Maas J, Pien CS, Prakash S, Elliott PJ (1999) Proteasome inhibitors: a novel class of potent and effective antitumor agents. *Cancer Res* 59: 2615–2622
- Aggen JB, Humphrey JM, Gauss CM, Huang HB, Nairn AC, Chamberlin AR (1999) The design, synthesis, and biological evaluation of analogues of the serine-threonine protein phosphatase 1 and 2A selective inhibitor microcystin LA: rational modifications imparting PP1 selectivity. *Bioorg Med Chem* 7: 543–564
- Ahuja D, Vera MD, SirDeshpande BV, Morimoto H, Williams PG, Joulié MM, Toogood PL (2000) Inhibition of protein synthesis

- by didemnin B: how EF-1 α mediates inhibition of translocation. *Biochemistry* 39: 4339–4346
- Akedo H, Isoai A, Hama Y, Kumagai H (1991) Novel polypeptides as cancer metastasis inhibitors. Patent JP 03,34,993. See Chem Abstr 115: 22221t
- Albericio F, Giralt E, Jimenez JC, Lopez A, Manzanera I, Rodrigues I, Royo M (2002) Preparation of kahalalide compounds. Patent WO 01 58,934. See Chem Abstr 135: 167039n
- Albert R, Smith-Jones P, Stolz B, Simeon C, Knecht H, Bruns C, Pless J (1998) Direct synthesis of [DOTA-DPhe1]-octreotide and [DOTA-DPhe1, Tyr3]-octreotide (SMT487): two conjugates for systemic delivery of radiotherapeutic nucleides to somatostatin receptor positive tumors in man. *Bioorg Med Chem Lett* 8: 1207–1210
- Andersen RJ, Coleman JE, Piers E, Wallace D (1997) Total synthesis of (–)-hemiasterlin, a structurally novel tripeptide that exhibits potent cytotoxic activity. *Tetrahedron Lett* 38: 317–320
- Andersen R, Piers E, Nieman J, Coleman J, Roberge M (1999) Hemiasterlin analogs. Patent WO 99 32,509. See Chem Abstr 131: 59140X
- Anderson HJ, Coleman JE, Andersen RJ, Roberge M (1997) Cytotoxic peptides hemiasterlin, hemiasterlin A and hemiasterlin B induce mitotic arrest and abnormal spindle formation. *Cancer Chemother Pharmacol* 39: 223–226
- Andreassen PA, Kjoller L, Christense L, Duffy M (1997) The urokinase-type plasminogen activator system in cancer metastasis: a review. *Int J Cancer* 72: 1–22
- Angiolillo A, Desgro A, Marsili V, Panara F, Gianfranceschi GL (1993) Synthetic octapeptide pyroGLU-ASP-ASP-SER-ASP-GLU-GLU-ASN controls DNA transcription in vitro by RNA polymerase II. *Experientia* 49: 902–905
- Anthony NJ, Desholms SJ, Gomez RP, Graham SL, Hutchinson JH, Stokker GE (1996) Preparation of peptide analogs as thiol-free inhibitors of farnesyl-protein transferase. Patent WO 96 10,034. See Chem Abstr 125: 143311n
- Aoyagi T, Ishizuka M, Takeuchi T, Umezawa H (1977) Enzyme inhibitors in relation to cancer therapy. *Jpn J Antibiot* 30 [Suppl]: 122–132
- Aoyama T, Satoh T, Yonemoto M, Shibata J, Nonoshita K, Arai S, Kawakami K, Iwasawa Y, Sano H, Tanaka K, Monden Y, Kodaera T, Arakawa H, Suzuki-Takahashi I, Kamei T, Tomimoto K (1998) A new class of highly potent farnesyl diphosphate-competitive inhibitors of farnesyltransferase. *J Med Chem* 41: 143–147
- Arnold E, Clardy J (1981) Crystal and molecular structure of BBM-928 A, a novel antitumor antibiotic from *Actinomadura luzonensis*. *J Am Chem Soc* 103: 1243–1244
- Asai A, Hasegawa A, Ochiai K, Yamashita Y, Mizukami T (2000) Belactosin A, a novel antitumor antibiotic acting on cyclin/CDK mediated cell cycle regulation, produced by *Streptomyces sp.* *J Antibiot* 53: 81–83
- Asai T, Nagatsuka M, Kuromi K, Yamakawa S, Kurohane K, Ogino K, Tanaka M, Taki T, Oku N (2002) Suppression of tumor growth by novel peptides homing to tumor-derived new blood vessels. *FEBS Lett* 510: 206–210
- Bae D-G, Gho Y-S, Yoon W-H, Chae C-B (2000) Arginine-rich anti-vascular endothelial growth factor peptides inhibit tumor growth and metastasis by blocking angiogenesis. *J Biol Chem* 275: 13588–13596
- Bai R, Durso NA, Sackett DL, Hamel E (1999) Interactions of the sponge-derived antimitotic tripeptide hemiasterlin with tubulin: comparison with dolastatin 10 and cryptophycin 1. *Biochemistry* 38: 14302–14310
- Bai R, Verdier-Pinard P, Gangwar S, Stessman CC, McClure KJ, Sausville EA, Pettit GR, Bates RB, Hamel E (2001) Dolastatin 11, a marine depsipeptide, arrests cells at cytokinesis and induces hyperpolymerization of purified actin. *Mol Pharmacol* 59: 462–469
- Bai R, Covell DG, Liu C, Ghosh AK, Hamel E (2002) (–)-doliculide, a new macrocyclic depsipeptide enhancer of actin assembly. *J Biol Chem* 277: 32165–32171
- Balazs A, Szirtes T, Schon I, Kisfaludy L (1992) Preparation of pyroaminoadipate-containing peptide as an epidermal cell proliferation inhibitor. Patent EP 475,657. See Chem Abstr 117: 8496w
- Ball KL, Lain S, Fähræus R, Smythe C, Lane DP (1996) Cell-cycle arrest and inhibition of Cdk4 activity by small peptides based on the carboxy-terminal domain of p21WAF1. *Curr Biol* 7: 71–80
- Bandara L, Girling R, La Thangue N (1997) Apoptosis induce in mammalian cells by small peptides that functionally antagonize the Rb-regulated E2F transcription factor. *Nat Biotechnol* 15: 896–901
- Bardelli A, Longati P, Grmaglia D, Stella MC, Comoglio PM (1997) Gabl coupling to the HGF/Met receptor multifunctional docking site requires binding of Grb2 and correlates with the transforming potential. *Oncogene* 15: 3103–3111
- Bardelli A, Longati P, Williams TA, Benvenuti S, Comoglio PM (1999) A peptide representing the carboxyl-terminal tail of the Met receptor inhibits kinase activity and invasive growth. *J Biol Chem* 274: 29274–29281
- Barnard D, Sun H, Baker L, Marshall MS (1998) In vitro inhibition of Ras-Raf association by short peptides. *Biochem Biophys Res Commun* 247: 176–180
- Bass RT, Buckwalter BL, Patel BP, Pausch MH, Price LA, Strnad J, Hadcock JR (1996) Identification and characterization of a novel somatostatin antagonist. *Mol Pharmacol* 50: 709–715
- Bass RT, Buckwalter BL, Patel BP, Pausch MH, Price LA, Strnad J, Hadcock JR (1997) Identification and characterization of a novel somatostatin antagonist, erratum. *Mol Pharmacol* 51: 170
- Bates RB, Brusoe KG, Burns JJ, Caldera S, Cui W, Gangwar S, Gramme MR, McClure KJ, Rouen GP, Schadow H, Stessman CC, Taylor SR, Vu VH, Yarick GV, Zhang J, Pettit GR, Bontems R (1997) Dolastatins 26. Synthesis and stereochemistry of dolastatin 11. *J Am Chem Soc* 119: 2111–2113
- Battey J, Wada E (1991) Two distinct receptor subtypes for mammalian bombesin-like peptides. *Trends Neurosci* 14: 524–528
- Battistini C, Giordano P, De Rosa S, Coradi F, Comoglio P, Bardelli A (1997) Peptide antagonists of cellular mitogenesis and motogenesis and their therapeutic use. Patent WO 97 30,079. See Chem Abstr 127: 234615x
- Bauer W, Briner U, Doepfner W, Haller R, Huguenin R, Marbach P, Petchr TJ, Pless J (1982) SMS 201-995: a very potent and selective octapeptide analogue of somatostatin with prolonged action. *Life Sci* 31: 1133–1140
- Berezov A, Zhang H-T, Greene MI, Murali R (2001) Disabling ErbB receptors with rationally designed exocyclic mimetics of antibodies: structure-function analysis. *J Med Chem* 44: 2565–2574
- Bernadi E, Fauchere JL, Atassi G, Viallfont P, Lazaro R (1993) Antitumoral cyclic peptide analogues of chlamydocin. *Peptides* 14: 1091–1093
- Bessodes M, Antonakis K, Herscovici J, Garcia M, Rochefort H, Capony F, Lelievre Y, Scherman D (1999) Inhibition of cathepsin D by tripeptides containing statine analogs. *Biochem Pharmacol* 58: 329–333
- Bewley CA, He H, Williams DH, Faulkner DJ (1996) Aciculitins A–C: cytotoxic and antifungal cyclic peptides from the lithistid sponge *Aciculites orientalis*. *J Am Chem Soc* 118: 4314–4321
- Binétruy-Tournaire R, Demangel C, Malavaud B, Vassy R, Rouyre S, Kraemer M, Poluët J, Derbin C, Perret G, Mazié JC (2000)

- Identification of a peptide blocking vascular endothelial growth factor (VEGF)-mediated angiogenesis. *EMBO J* 19: 1525–1533
- Blanco-Aparicio C, Molina MA, Fernández-Salas E, Frazier ML, Mas JM, Querol E, Avilés FX, de Llorens R (1998) Potato carboxypeptidase inhibitors, a T-knot protein is an epidermal growth factor antagonist that inhibits tumor cell growth. *J Biol Chem* 273: 12370–12377
- Blaschuk OW, Gour BJ, Farookhi R (2002) Compounds and method for cancer therapy using cadherin cell recognition cyclic peptides. Patent US 6,417,325. See Chem Abstr 137: 88441a
- Blasi F, Fazioli F, Resnati M, Nicolai S (1998) Peptides mimicking uPAR and having chemotactic activity and therapeutic use of thereof. Patent WO 98 42,733. See Chem Abstr 129: 270635d
- Boehm HJ, Daum L, Haupt A, Schmied B, Walker N, Zechel JC (1990a) Preparation of low molecular weight peptides with tumor necrosis factor (TNF) activity. Patent DE 3,841,768. See Chem Abstr 113: 232082h
- Boehm HJ, Daum L, Haupt A, Schmied B, Walker N, Zechel JC (1990b) Preparation of peptides as tumor necrosis factor agonists-antagonists. Patent DE 3,841,767. See Chem Abstr 113: 232079n
- Boehm HJ, Daum L, Haupt A, Schmied B, Walker N, Zechel JC (1990c) Preparation of tumor necrosis factor analogs. Patent DE 3,841,763. See Chem Abstr 113: 232081g
- Bogden AE, Taylor JE, Moreau JP, Coy DH, LePage DJ (1990) Response of the human lung tumor xenografts to treatment with a somatostatin analogue (somatuline). *Cancer Res* 50: 4360–4365
- Boger DL, Ichikawa S (2000) Total syntheses of thiocoraline and BE-22179: establishment of relative and absolute stereochemistry. *J Am Chem Soc* 122: 2956–2957
- Boger DL, Saionz KW (1999) DNA binding properties of key sandramycin analogues: systematic examination of the intercalation chromophore. *Bioorg Med Chem* 7: 315–321
- Boger DL, Chen JH, Saionz KW, Jin Q (1998) Synthesis of key sandramycin analogs: systematic examination of the intercalation chromophore. *Bioorg Med Chem* 6: 85–102
- Boger DL, Ledebauer MW, Kume M, Searcey M, Jin Q (1999) Total synthesis and comparative evaluation of Luzopeptin A-C and Quinoxapeptin A-C. *J Am Chem Soc* 121: 11375–11388
- Bogusky MJ, Culbertson JC, Pitzberger SM, Garsky VM, Wallace A, Pessi A, Koblan KS (1999) Conformation of a novel tetrapeptide inhibitor NH₂-D-Trp-D-Met-Phe(pCl)-Gla-NH₂ bound to farnesyl-protein transferase. *J Pept Res* 54: 66–73
- Bonfanti M, Taverna S, Salmona M, D'Incalci M, Brogginini M (1997) P21WAF1 derived peptides linked to an internalization peptide inhibit human cancer cell growth. *Cancer Res* 57: 1442–1446
- Bonnington LS, Tanaka J, Higa T, Kimura J, Yoshimura Y, Nakao Y, Yoshida WY, Scheuer PJ (1997) Cupolamide A: a cytotoxic cyclic heptapeptide from two samples of the sponge *Theonella cupola*. *J Org Chem* 62: 7765–7767
- Böttger V, Böttger A, Howard SF, Picksley SM, Chène P, García-Echeverría C, Hochkeppel HK, Lane DP (1996) Identification of novel Mdm2 binding peptides by phage display. *Oncogene* 13: 2141–2147
- Böttger V, Böttger A, García-Echeverría C, Chène P, Hochkeppel HK, Sampson W, Ang K, Howard SF, Picksley SM, Lane DP (1997) Molecular characterization of the hdm2-p53 interaction. *J Mol Biol* 269: 744
- Bousquet C, Puente E, Buscail L, Vaysse N, Susini C (2001) Antiproliferative effect of somatostatin and analogs. *Chemotherapy* 47 [Suppl 2]: 30–39
- Brazeau P, Vale WW, Burgus R, Ling N, Butcher M, Rivier J, Guillemin R (1973) Hypothalamic polypeptide that inhibits the secretion of immunoreactive pituitary growth hormone. *Science* 179: 77–79
- Breakman JC, Daloze D, Moussiaux B, Riccio R (1987) Jaspamide from the marine sponge *Jaspis Johnstoni*. *J Nat Prod* 50: 994–995
- Brooks PC, Montgomery AMP, Rosenfeld M, Reisfeld RA, Hu T, Klier G, Cheresch DA (1994) Integrin $\alpha v \beta 3$ antagonists promote tumor-regression by inducing apoptosis of angiogenic blood-vessels. *Cell* 79: 1157–1164
- Brown PD (1999) Clinical studies with matrix metalloproteinase inhibitors. *APMIS* 107: 174–180
- Bubb MR, Senderowicz AMJ, Duncan IKK, Korn ED (1994) Jaspalinalide, a cytotoxic natural product induces actin polymerisation and completely inhibits the binding of phalloidin to F-actin. *J Biol Chem* 269: 14869–14871
- Bubb MR, Spector I, Beyer BB, Fosen KM (2000) Effects of jaspalinalide on the kinetics of actin polymerization. An explanation for certain *in vivo* observations. *J Biol Chem* 275: 5163–5170
- Buchanan J, Bohacek R, Vu CB, Luke GP (1999a) Preparation of heterocyclic phosphotyrosine derivatives as SH2-mediated signal transduction inhibitors. Patent WO 99 47,529. See Chem Abstr 131: 257875d
- Buchanan JL, Bohacek RS, Luke GP, Hatada M, Lu X, Dalgarno DC, Narula SS, Yuan R, Holt DA (1999b) Structure-based design and synthesis of a novel class of SRC SH2 inhibitors. *Bioorg Med Chem Lett* 9: 2359–2364
- Buchanan JL, Vu CB, Merry TJ, Corpuz EG, Pradeepan SG, Mani UN, Yang M, Plake HR, Varkhedhar VM, Lynch BA, MacNeil IA, Loiacono KA, Tiong CL, Holt DA (1999c) Structure-activity relationships of a novel class of SRC SH2 inhibitors. *Bioorg Med Chem Lett* 9: 2353–2358
- Buckley CD, Pilling D, Henriquez NV, Parsonage G, Threlfall K, Scheel-Toellner D, Simmons DL, Akbar AN, Lord JM, Salmon M (1999) RGD peptides induce apoptosis by direct caspase-3 activation. *Nature* 397: 534–539
- Burgess AJ, Pavey S, Warrener R, Hunter L-JK, Piva TJ, Musgrove EA, Saunders N, Parsons PG, Gabrielli BG (2001) Up-regulation of p2WAF/CIP1 by histone deacetylase inhibitors reduces their cytotoxicity. *Mol Pharmacol* 60: 828–837
- Bürge M, Koppitz M, Riemer C, König B, Weidle UH, Kellerman J, Lottspeich F, Graeff H, Schmitt M, Goretzki L, Reuning U, Wilhelm O, Magdolen V (1997) Inhibition of the interaction of urokinase-type plasminogen activator (uPA) with its receptor (uPAR) synthetic peptides. *Biol Chem* 378: 231–237
- Burman AC, Prasad S, Mukherjee R, Jaggi M, Singh AT, Mathur A (2001a) Synthesis of bombesin peptide analogs and their use in treatment of cancer. Patent WO 01 62,777. See Chem Abstr 135: 190403k
- Burman AC, Prasad S, Mukherjee R, Singh AT, Mathur A, Gupta N (2001b) Preparation of vasoactive intestinal peptide analogs as anticancer agents. Patent WO 01 60,862. See Chem Abstr 135: 195792w
- Burman AC, Prasad S, Mukherjee R, Jaggi M, Singh AT (2002) Preparation of substance P analogs for the treatment of cancer. Patent WO 02 10,194. See Chem Abstr 136: 151442y
- Butz K, Denk C, Ullman A, Schefner M, Hoppe-Seyler F (2000) Induction of apoptosis in human papillomavirus-positive cancer cells by peptide aptamers targeting the viral E6 oncoprotein. *Proc Natl Acad Sci USA* 97: 6693–6697
- Caba JM, Rodriguez IM, Manzanares I, Giralt E, Albericio F (2001) Solid-phase synthesis of trunkamide A. *J Org Chem* 66: 7568–7574
- Cai RZ, Szoke B, Lu R, Fu D, Redding TW, Schally AV (1986) Synthesis and biological activity of highly potent octapeptide analogs of somatostatin. *Proc Natl Acad Sci USA* 83: 1896–1900

- Campbell RM, Scanes CG (1992) Evolution of the growth hormone-releasing factor (GRF) family of peptides. *Growth Regul* 2: 175–191
- Cao Y, Ji R, Davidson D, Schaller J, Marti D, Sohndel S, McCance SG, O'Reilly MS, Llinas M, Folkman J (1996) Kringle domains of human angiostatin. Characterization of the anti-proliferative activity on endothelial cells. *J Biol Chem* 271: 29461–29467
- Cao Y, Chen A, An SS, Ji RW, Davidson D, Llinas M (1997) Kringle 5 of plasminogen is a novel inhibitor of endothelial cell growth. *J Biol Chem* 272: 22924–22928
- Cardellini E, Ponti D, Gianfranceschi GL (1999) Phosphorylation of the synthetic octapeptide pyroGlu-ASP-ASP-SER-ASP-GLU-GLU-ASN and binding to DNA in presence of divalent cations. *Mol Biol Rep* 26: 255–259
- Cardona C, Rabitts PH, Spindel ER, Ghatei MA, Blehen NM, Bloom SR, Reeve JG (1991) Production of neuromedin B and neuromedin B gene expression in human lung tumor cell lines. *Cancer Res* 51: 5205–5211
- Carroll AR, Bowden BF, Coll JC, Hockless CR, Skelton BW, White AH (1994) Studies of Australian ascidians. Mollamide, a cytotoxic cyclic heptapeptide from the compound ascidian *Didemnum molle*. *Aust J Chem* 47: 61–69
- Carroll AR, Coll JC, Bourne DJ, MacLeod JC, Zabriskie TM, Ireland CM, Bowden BF (1996a) Patellins 1–6 and trunkamide A: novel cyclic hexa-, hepta- and octa-peptides from colonial ascidians, *Lissoclinum* sp. *Aust J Chem* 49: 659–667
- Carroll AR, Feng Y, Bowden BF, Coll JC (1996b) Studies of Australian ascidians 5. Virenamidines A–C, new cytotoxic linear peptides from the colonial didemnid ascidian *Diplosoma virens*. *J Org Chem* 61: 4059–4061
- Carter DC, Moore RE, Mynderse JS, Niemezura WP, Todd JS (1984) Structure of majusculamide C, a cyclic depsipeptide from *Lyngbya majuscula*. *J Org Chem* 49: 236–241
- Castronovo V, Taraboletti G, Sobel ME (1991) Laminin receptor complementary DNA-deduced synthetic peptide inhibits cancer cell attachment to endothelium. *Cancer Res* 51: 5672–5678
- Chan WR, Tinto WF, Manchand PS, Todaro LJ (1987) Stereostructures of geodiamolides A and B, novel cyclodepsipeptides from the marine sponge *Geodia* sp. *J Org Chem* 52: 3091–3093
- Chan KK, Bakhtiar R, Jiang C (1997) Depsipeptide (FR901228, NSC-630176) pharmacokinetics in the rat by LC/MS/MS. *Invest New Drugs* 15: 195–206
- Chang AY, Kim K, Boucher H, Bonomi P, Stewart JA, Karp DD, Blum RH (1998) A randomized phase II trial of echinomycin, trimetrexate, and cisplatin plus etoposide in patients with metastatic non small cell lung carcinoma: an Eastern Cooperative Oncology Group Study (E1587). *Cancer* 15: 292–300
- Chao DT, Korsmeyer SJ (1998) Bcl-2 family: regulators of cell death. *Ann Rev Immunol* 16: 395–419
- Chen J, Saha P, Kornbluth S, Dynlacht BD, Dutta A (1996) Cyclin-binding motifs are essential for the function of p21Cipl. *Mol Cell Biol* 16: 4673–4682
- Chène P, Fuchs J, Bohn J, García-Echeverría C, Furet P, Fabbro D (2000) A small synthetic peptide, which inhibits the p53-hdm2 interaction, stimulates the p53 pathway in tumour cell lines. *J Mol Biol* 299: 245–253
- Chill L, Kashman Y, Schleyer M (1997) Oriamide, a new cytotoxic cyclic peptide containing a novel amino acid from the marine sponge *Theonella* sp. *Tetrahedron* 53: 16147–16152
- Chillemi F, Lugaro G, Boari D, Cardellini E, Bramucci M, Miano A, Amici D, Gianfranceschi GL, Durban E (1991) Acidic pentapeptide phosphorylated in vitro by calf thymus protein kinase NII binds to DNA in the presence of Mg²⁺ cations. *FEBS Lett* 291: 67–70
- Clark WD, Corbett T, Valeriote F, Crews P (1997) Cyclocinamide A. An unusual cytotoxic halogenated hexapeptide from the marine sponge *Psammocinia*. *J Am Chem Soc* 119: 9285–9286
- Cohen B, Colas P, Brent R (1998) An artificial cell-cycle inhibitor isolated from a combinatorial library. *Proc Natl Acad Sci USA* 95: 14272–14277
- Colas P, Cohen B, Jessen T, Grishina I, McCoy J, Brent R (1996) Genetic selection of peptide aptamers that recognize and inhibit cyclin-dependent kinase 2. *Nature* 380: 548–550
- Coleman JE, de Silva ED, Kong F, Andersen RJ, Allen TM (1995) Cytotoxic peptides from the marine sponge *Cymbastela* sp. *Tetrahedron* 51: 10653–10662
- Coleman JE, van Soest R, Andersen RJ (1999) New geodiamolides from the sponge *Cymbastela* sp. collected in Papua New Guinea. *J Nat Prod* 62: 1137–1141
- Coleman PJ, Le TD (2002) Ligands to the integrin receptor $\alpha v \beta 3$. *Expert Opin Ther Patents* 12: 1009–1021
- Comoglio P, Ponzetto C (1995) Peptide inhibitors of mitogenesis and motogenesis. Patent WO 9501376. See Chem Abstr 122: 282231n
- Corbett TH, Valeriote FA, Demchik L, Lowichik N, Polin L, Panchapor C, Pugh M, White K, Juiwanna RK, Wentland M, Golakoti T, Hetzel C, Ogino J, Patterson G, Moore R (1997) Discovery of cryptophycin-1 and BCN-183577: examples of strategies and problems in the detection of antitumor activity in mice. *Invest New Drugs* 15: 207–218
- Corey EJ, Wei-Dong ZL (1999) Total synthesis and biological activity of Lactacystin, Omuralide and analogs. *Chem Pharm Bull* 47: 1–10
- Correia JJ (1991) Effects of antimetabolic agents on tubulin-nucleotide interactions. *Pharmacol Ther* 52: 127–147
- Cosulich SC, Worrall V, Hedge PJ, Green S, Clarke PR (1997) Regulation of apoptosis by BH3 domains in a cell-free system. *Curr Biol* 7: 913–920
- Craig WS, Cheng S, Mullen DG, Blevitt J, Pierschbacher MD (1995) Concept and progress in the development of RGD-containing peptide pharmaceuticals. *Biopolymers* 37: 157–175
- Creighton DJ, Hamilton DS, Kavarana MJ, Sharkey E, Eiseman JL (2000) Glyoxalase enzyme system as a potential target for anti-tumor drug development. *Drugs Future* 25: 385–392
- Crews P, Manes LV, Boehler M (1986) Jaspilkinolide, a cyclodepsipeptide from the marine sponge *Jaspis* sp. *Tetrahedron Lett* 27: 2797–2800
- Crews P, Farias JJ, Emrich R, Kiefer PA (1994) Milnamide A, an unusual cytotoxic tripeptide from the marine sponge *Auletta cf. constricta*. *J Org Chem* 59: 2932–2934
- Crul M, de Klerk GJ, Beijnen JH, Schellens JH (2001) Ras biochemistry and farnesyl transferase inhibitors: a literature survey. *Anti Cancer Drugs* 12: 163–184
- Cui C-B, Kakeya H, Okada G, Onose R, Ubukata M, Takahashi I, Isono K, Osada H (1995) Tryprostatins A and B, novel mammalian cell cycle inhibitors produced by *Aspergillus fumigatus*. *J Antibiot* 48: 1382–1384
- Cui C-B, Kakeya H, Okada G, Onose R, Osada H (1996a) Novel mammalian cell cycle inhibitors, tryprostatins A, B and other diketopiperazines produced by *Aspergillus fumigatus*. In taxonomy, fermentation, isolation and biological properties. *J Antibiot* 49: 527–533
- Cui C-B, Kakeya H, Osada H (1996b) Novel mammalian cell cycle inhibitors, tryprostatins A, B and other diketopiperazines produced by *Aspergillus fumigatus*. II Physio-chemical properties and structures. *J Antibiot* 49: 534–540
- Cui CB, Kakeya H, Osada H (1996c) Novel mammalian cell cycle inhibitors, spirotrostatins A and B, produced by *Aspergillus fumigatus*, which inhibit mammalian cell cycle at G2/M phase. *Tetrahedron* 52: 12651–12666

- Cussac D, Vidal M, Leprince C, Liu WQ, Cornille F, Tiraboschi G, Roques BP, Garbay C (1999) A sos-derived peptimer blocks the ras signalling pathway by binding both Grb2 SH3 domains and displays antiproliferative activity. *FASEB J* 13: 31–38
- Cuttitta F, Carney DN, Mulshine J, Moody TW, Fedorko J, Fischler A, Minna J (1985) Bobesin-like peptides can function as autocrine growth factors in human small-cell lung cancer. *Nature* 316: 823–826
- D'Souza SE, Ginsberg MH, Plow EF (1991) Arginyl-glycyl-aspartic acid (RGD) a cell adhesion motif. *Trends Biochem Sci* 16: 246–250
- Daiichi K, Yakuhi KK (1985) Peptide aldehydes. Patent JP 58,164,563. See *Chem Abstr* 100: 139623g
- Dawson DW, Volpert OV, Pearce SF, Schneider AJ, Silverstein RL, Henkin J, Bouck NP (1999) Three distinct D-amino acid substitutions confer potent antiangiogenic activity on an inactive peptide derived from a thrombospondin-1 type 1 repeat. *Mol Pharmacol* 55: 332–338
- de Arruda M, Cocchiario CA, Nelson CM, Grinnell CM, Janssen B, Haupt A, Barlozzari T (1995) LU103793 (NSC D-669356): A synthetic peptide that interacts with microtubules and inhibits mitosis. *Cancer Res* 55: 3085–3092
- De B, Natchus MG, Cheng M, Pikul S, Almstead NG, Taiwo YO, Snider CE, Chen L, Barnett B, Gu F, Dowty M (1999) The next generation of MMP inhibitors: design and synthesis. *Ann N Y Acad Sci* 878: 40–60
- de Castiglione R, Gozzini L (1996) Bombesin receptor antagonists. *Crit Rev Oncology-Hematology* 24: 117–151
- de Silva ED, Andersen RJ, Allen TM (1990) Geodiamolides C to F, new cytotoxic cyclodepsipeptides from the marine sponge *Pseudaxynssa*. *Tetrahedron Lett* 31: 489–492
- de Silva ED, Williams dDE, Andersen RJ, Klix H, Holmes CFB, Allen TM (1992) Motuporin, a potent protein phosphatase inhibitor isolated from the Papua New Guinea sponge *Theonella swinhoei* gray. *Tetrahedron Lett* 33: 1561–1564
- Dechantsreiter MA, Planker E, Mathä B, Lohof E, Hölzemmann AJ, Goodman SL, Kessler H (1999) N-methylated cyclic RGD peptides as highly active and selective $\alpha\beta3$ integrin antagonist. *J Med Chem* 42: 3033–3040
- Degenkolb T, Heinze S, Schlegel B, Dornberger K, Mollmann U, Dahse H-M, Grafe U (2000) Roseoferin, a new aminolipopeptide antibiotic complex from *Mycogone rosea* DSM 12973, structures and biological activities. *J Antibiot* 53: 184–190
- Delic J, Masdehors P, Ömura S, Cosset J-M, Dumont J, Binet J-L, Magdelénat H (1998) The proteasome inhibitor lactacystin induces apoptosis and sensitizes chemo- and radioresistant human chronic lymphocytic leukaemia lymphocytes to TNF- α -initiated apoptosis. *Br J Cancer* 77: 1103–1107
- Dell A, Williams DH, Morris HR, Smith GA, Feeney J, Roberts CGK (1975) Structure revision of the antibiotics echinomycin. *J Am Chem Soc* 97: 2497–2502
- Diaz JL, Oltersdorf T, Horne W, McConnell M, Wilson G, Weeks S, Garcia T, Fritz LC (1997) A common binding site mediates heterodimerization and homodimerization of Bcl-2 family members. *J Biol Chem* 272: 11350–11355
- Draeger LJ, Mullen GP (1994) Interaction of the bHLH-zip domain of c-Myc with H1-type peptides. *J Biol Chem* 269: 1785–1793
- Duffy MJ (1992) The role of proteolytic enzymes in cancer invasion and metastasis. *Clin Exp Metastasis* 10: 145–155
- Duncan SJ, Grūshow S, William DH, McNicholas C, Purawal R, Hajek M, Gerlitz M, Martin S, Wrigley SK, Moore M (2001) Isolation and structure elucidation of chlorofusin a novel p53-MDM2 antagonist from *Fusarium sp.* *J Am Chem Soc* 123: 554–560
- Dutta AN (1988a) Luteinizing hormone-releasing hormone (LHRH) agonists. *Drugs Future* 13: 43–57
- Dutta AN (1988b) Luteinizing hormone-releasing hormone (LHRH) antagonists. *Drugs Future* 13: 761–787
- Eidler MC, Fernandez AM, Lassota P, Ireland CM, Barrows LR (2002) Inhibition of tubulin polymerization by vitilevuamide, a bicyclic marine peptide, at a site distinct from colchicine, the vinca alkaloids, and dolastatin 10. *Biochem Pharmacol* 63: 707–715
- Eggen M, Georg GI (2002) The cryptophycins: their synthesis and anticancer activity. *Med Res Rev* 22: 85–101
- Elofson M, Splitterger U, Myung J, Mohan R, Crews CM (1999) Towards subunit-specific proteasome inhibitors: synthesis and evaluation of peptide α',β' -epoxyketones. *Chemistry Biology* 6: 811–822
- Enyedy IJ, Ling Y, Nacro K, Tomita Y, Wu X, Cao Y, Guo R, Li B, Zhu X, Huang Y, Long YQ, Roller PP, Yang D, Wang S (2001) Discovery of small-molecule inhibitors of Bcl-2 through structure-based computer screening. *J Med Chem* 44: 4313–4324
- Erba E, Bergamaschi D, Ronzoni S, Faretta M, Taverna S, Bonfati M, Catapano CV, Faircloth G, Jimeno J, D'Incalci M (1999) Mode of action of thiocoraline, a natural marine compound with antitumour activity. *Br J Cancer* 80: 971–980
- Eriksson B, Oberg K (1999) Summing up 15 years of somatostatin analog therapy in neuroendocrine tumors: future outlook. *Ann Oncol* 10 [Suppl 2]: S31–38
- Ettmayer P, France D, Gounarides J, Jarosinski M, Martin M-S, Rondeau J-M, Sabio M, Topiol S, Weidmann M, Zurini M, Bair KW (1999) Structural and conformational requirements for high-affinity binding to the SH2 domain of grb2. *J Med Chem* 42: 971–980
- Everard MJ, Macaulay VM, Millar JL, Smith IE (1993) [D-Arg1, D-Phe5, D-Trp7,9, Leu11] substance P inhibits the growth of human small cell lung cancer xenografts in vivo. *Eur J Cancer* 29: 1450–1453
- Fabbrizio E, Le Cam L, Polanowska J, Kaczorek M, Lamb N, Brent R, Sardet C (1999) Inhibition of mammalian cell proliferation by genetically selected peptide aptamers that functionally antagonize E2F activity. *Oncogene* 18: 4357–4363
- Fähræus R, Lain S, Ball KL, Lane DP (1998) Characterization of the cyclin-dependent kinase inhibitory domain of the INK4 family as a model for a synthetic tumour suppressor molecule. *Oncogene* 16: 587–596
- Fähræus R, Paramio JM, Ball KL, Lain S, Lane DP (1996) Inhibition of pRb phosphorylation and cell-cycle progression by a 20-residue peptide derived from p16CDK2/INK4A. *Curr Biol* 6: 84–91
- Fairbrother WJ, Christinger HW, Cochran AG, Fuh G, Keenan CJ, Quan C, Shriver SK, Tom JY, Wells JA, Cunningham BC (1998) Novel peptides selected to bind vascular endothelial growth factor target the receptor-binding site. *Biochemistry* 37: 17754–17764
- Faulkner DJ (2002) Marine natural products. *Nat Prod Rep* 19: 1–48
- Fazioli F, Blasi F (1994) Urokinase-type plasminogen activator and its receptor: new targets for anti-metastatic therapy? *Trends Pharmacol Sci* 15: 25–29
- Fernández R, Rodríguez J, Quiñoá E, Riguerra R, Muñoz L, Fernández-Suárez M, Debitus C (1996) Onchidin B: a new cyclodepsipeptide from the mollusc *Onchidium sp.* *J Am Chem Soc* 118: 11635–11643
- Finnegan NM, Curtin JF, Prevost G, Morgan B, Cotter TG (2001) Induction of apoptosis in prostate carcinoma cells by BH3 peptides which inhibit Bak/Bcl-2 interactions. *Br J Cancer* 85: 115–121
- Fischer PM, Zhelev NZ, Wang S, Melville JE, Fähræus R, Lane DP (2000) Structure-activity relationship of truncated and substituted

- analogues of the intracellular delivery vector Penetratin. *J Pept Res* 55: 163–172
- Fisher A, Yang FD, Rubin H, Cooperman BS (1993) R2 C-terminal peptide inhibition of mammalian and yeast ribonucleotide reductase. *J Med Chem* 36: 3859–3862
- Foa P, Chillemi F, Lombardi L, Lonati S, Miaolo AT, Polli EE (1987) Inhibitory activity of synthetic pentapeptide on leukemic myelopoiesis both *in vitro* and *in vivo* in rats. *Eur J Haematol* 39: 399–403
- Ford PW, Gustafson KR, McKee TC, Shigematsu N, Maurizi LK, Pannell LK, Williams DE, de Silva ED, Lassota P, Allen TM, van Soest R, Andersen RJ, Boyd MR (1999) Papuamides A-D, HIV-inhibitory and cytotoxic depsipeptides from the sponges *Theonella mirabilis* and *Theonella swinhoei* collected in Papua New Guinea. *J Am Chem Soc* 121: 5899–5909
- Foster BA, Coffey HA, Morin MJ, Rastinjad F (1999) Pharmacological rescue of mutant p53 conformation and function. *Science* 286: 2507–2510
- Frankenne F, Noel A, Bajou K, Sounni NE, Goffin F, Mason V, Munaut C, Remacle A, Froidart JM (1999) Molecular interactions involving urokinase plasminogen activator (uPA), its receptor (uPAR) and its inhibitor, plasminogen activator inhibitor-1 (PAI-1), as new targets for tumour therapy. *Emerging Ther Targets* 3: 469–481
- Fretz H, Furet P, García-Echeverría C, Rahuel J, Schoepfer J (2000) Structure-based design of compounds inhibiting Grb2-SH2 mediated protein-protein interactions in signal transduction pathways. *Curr Pharm Des* 6: 1777–1796
- Fujisawa N, Hayashi S, Miller EJ (1999) A synthetic peptide inhibitor for alpha-chemokines inhibits the tumour growth and pulmonary metastasis of human melanoma cells in nude mice. *Melanoma Res* 9: 105–114
- Fukuda MN, Ohvama C, Lowitz K, Matsuo O, Pasqualini R, Ruoslati E, Fukuda M (2000) A peptide mimic of E-selectin ligand inhibits sialyl Lewis X-dependent lung colonisation of tumor cells. *Cancer Res* 60: 450–456
- Furuta T, Hayashi H (1986) Antitumor protein isolation from human cell line. Patent JP 60,226,816. See Chem Abstr 104: 180205t
- Fusetani N, Sugawara T, Matsunaga S, Hirota H (1991) Orbiculamide A: a novel cytotoxic cyclic peptide from a marine sponge *Theonella* sp. *J Am Chem Soc* 113: 7811–7812
- Gademann K, Kimmerlin T, Hoyer D, Seebach D (2001) Peptide folding induces high and selective affinity of a linear and small β -peptide to the human somatostatin receptor. *J Med Chem* 44: 2460–2468
- Gademann K, Seebach D (2001) Synthesis of cyclo- β -tripeptides and their biological *in vitro* evaluation as antiproliferative against the growth of human cancer cell lines. *Helv Chim Acta* 84: 2924–2937
- Galvez AF (2002) Therapeutic peptides having a motif that binds specifically to nonacetylated H3 and H4 histones for cancer therapy. Patent WO 01 72,784. See Chem Abstr 135: 283175c
- Galvez AF, Chen N, Macsaieb J, de Lumen BO (2001) Chemopreventive property of a soybean peptide (lunasin) that binds to deacetylated histones and inhibits their acetylation. *Cancer Res* 61: 7473–7478
- Gamble WR, Durso NA, Fuller RW, Westergaard CK, Johnson TR, Sackett DL, Hamel E, Cardellina JH, Boyd MR (1999) Cytotoxic and tubulin-interactive hemiasterlins from *Auletta* sp. and *Siphonochalina* spp. sponges. *Bioorg Med Chem* 7: 1611–1615
- Garbay C, Liu W-Q, Vidal M, Roques BP (2000) Inhibitors of RAS signal transduction as antitumor agents. *Biochem Pharmacol* 60: 1165–1169
- García-Echeverría C, Chène P, Blommers MJJ, Furet P (2000) Discovery of potent antagonists of the interaction between human double minute 2 and tumor suppressor p53. *J Med Chem* 43: 3205–3208
- Gaudron S, Grillon C, Thierry J, Riches A, Wierenga PK, Wdzieczak-Bakala J (1999) *In vitro* effect of acetyl-N-Ser-Asp-Lys-Pro (AcSDKP) analogs resistant to angiotensin I-converting enzyme on hematopoietic stem cell and progenitor cell proliferation. *Stem Cells* 17: 100–106
- Gay B, Suarez S, Caravatti G, Furet P, Meyer T, Schoepfer J (1999) Selective GRB2 SH2 inhibitors as anti-ras therapy. *Int J Cancer* 83: 235–241
- Gazal S, Gelerman G, Ziv O, Karpov O, Litman P, Bracha M, Afargan M, Gilon C (2002) Human somatostatin receptor specificity of backbone-cyclic analogues containing novel sulfur building units. *J Med Chem* 45: 1665–1671
- Gazos Lopes U, Erhardt P, Yao R, Cooper GM (1997) p53-dependent induction of apoptosis by proteasome inhibitors. *J Biol Chem* 272: 12893–12896
- Ge R, Kini RM (2001) Small peptides having potent anti-angiogenic activity. Patent WO 01 18,030. See Chem Abstr 134: 217187m
- Gembistky DS, De Angelis PM, Reichelt KL, Eljgo K (2000) An endogenous melanocyte-inhibiting tripeptide pyroGlu-Phe-GlyNH₂ delays *in vivo* growth of monoclonal experimental melanoma. *Cell Proliferation* 33: 91–99
- Georg GI, Ali SM, Stella VJ, Waugh WN, Himes RH (1998) Halohydrin analogues of cryptophycin 1: synthesis and biological activity. *Bioorg Med Chem Lett* 8: 1959–1962
- Giannakakou P, Sackett DL, Ward Y, Webster KR, Blagosklonny MV, Fojo T (2000) p53 is associated with cellular microtubules and is transported to the nucleus by dynein. *Nat Cell Biol* 2: 709–717
- Gilmer T, Rodriguez M, Jordan S, Crosby R, Alligood K, Green M, Kimery M, Wagner C, Kinder D, Charifson P, Hassel AM, Willard D, Luther M, Rusnak D, Sternbach DD, Mehrotra M, Peel M, Burkhart W, Moyer M, Brdshaw T, Berman J (1994) Peptide inhibitors of src SH3-SH2-phosphoprotein interaction. *J Biol Chem* 269: 31711–31719
- Giorello L, Clerico L, Pescarolo MP, Vikhanskaya F, Salmona M, Colella G, Bruno S, Mancuso T, Bagansco L, Russo P, Parodi S (1998) Inhibition of cancer cell growth and c-Myc transcriptional activity by a c-Myc helix 1-type peptide fused to an internalization sequence. *Cancer Res* 58: 3654–3659
- Goetz G, Yoshida WY, Scheuer PJ (1999) The absolute stereochemistry of Kahalalide F. *Tetrahedron* 55: 7739–7746
- Goldberg J, Huang H, Kwon Y, Greengard P, Nairn AC, Kuriyan J (1995) Three-dimensional structure of the catalytic subunit of protein serine/threonine phosphatase-1. *Nature* 376: 745–753
- Goodson RJ, Doyle MV, Kaufman SE, Rosenberg S (1994) High-affinity urokinase receptor antagonists identified with bacteriophage peptide display. *Proc Natl Acad Sci USA* 91: 7129–7133
- Gradishar WJ, Vogelzang NJ, Kilton LJ, Leibach SJ, Rademaker AW, French S, Benson AB (1995) A phase II clinical trial of echinomycin in metastatic soft tissue sarcoma. An Illinois Cancer Center Study. *Invest New Drugs* 13: 171–174
- Gräfe U, Schlegel R, Ritzau M, Ihn W, Dornberger K, Stengel C, Fleck WF, Gitsche W, Härtl A, Paulus EF (1995) Aurantimycins, new depsipeptide antibiotics from *Streptomyces aurantiacus* IMET 43917 production, isolation, structure elucidation, and biological activity. *J Antibiot* 48: 119–125
- Graz CJ, Grant GD, Brauns SC, Hunt A, Jamie H, Milne PJ (2000) Cyclic dipeptides in the induction of maturation for cancer therapy. *J Pharm Pharmacol* 52: 75–82

- Grieco PA, Reilly M (1998) Studies related to the absolute configuration of cyclocinamide A: total synthesis of 4(R),11(R)-cyclocinamide. *Tetrahedron Lett* 39: 8925–8928
- Gross A, McDonnell JM, Korsmeyer SJ (1999) BCL-2 family members and the mitochondria in apoptosis. *Genes Dev* 13: 1899–1911
- Gulavita NK, Gunasekera SA, Pomponi SA, Robinson EV (1992) Polydiscamide A: a new bioactive depsipeptide from the marine sponge *Discodermia* sp. *J Org Chem* 57: 1767–1772
- Guo NH, Krutzsch HC, Inman JK, Shannon CS, Roberts DD (1997) Antiproliferative and antitumor activities of D-reverse peptides derived from the second type-1 repeat of thrombospondin-1. *J Pept Res* 50: 210–221
- Guo Y, Higazi AA-R, Arakelian A, Sachais BS, Cines D, Goldfarb RH, Jones TR, Kwaan H, Mazar AP, Rabbani SA (2000) A peptide derived from the nonreceptor binding region of urokinase plasminogen activator (uPA) inhibits tumor progression and angiogenesis and induces tumor cell death in vivo. *FASEB J* 14: 1400–1410
- Halazonetis T, Hartwig W (1996) Peptides and peptidomimetics which activate human p53 DNA binding and their use as therapeutics. Patent WO 96 25,434. See Chem Abstr 125: 257158y
- Halazonetis T, Hartwig W (2001) Peptides and peptidomimetics with structural similarity to human p53 that activate p53 function. Patent US 6,245,886. See Chem Abstr 135: 51039h
- Hall I, Chen SY (1999) Substituted 4-hydroxyproline di- and tripeptides as cytotoxic agents. *Amino Acids* 16: 79–89
- Hamann MT, Scheuer PJ (1993) Kahalalide F: a bioactive depsipeptide from the sacoglossan mollusk *Elysia rufescens* and the green alga *Bryopsis* sp. *J Am Chem Soc* 115: 5825–5826
- Hamel E (1992) Natural products which interact with tubulin in the vinca domain: maytansine, rhizoxin, phomopsin A, dolastatin 10 and 12 and halichondrin B. *Pharmacol Ther* 55: 31–51
- Han DC, Shen TL, Guan JL (2001) The Grb7 family proteins: structure, interactions with other signalling molecules and potential cellular functions. *Oncogene* 20: 6315–6321
- Hao Y, Zhang R, Li C, Xu X (1995) Design and synthesis of antibacterial and antitumor peptides containing a SPXX motif. *Beijing Daxue Xuebao, Ziran Kexueban* 31: 703–710 See Chem Abstr 124: 331693s
- Harrigan GG, Harrigan BL, Davidson BS (1997) Kailuins A-D, new cyclic acyldepsipeptides from cultures of a marine-derived bacterium. *Tetrahedron* 53: 1577–1582
- Harrigan GG, Yoshida WY, Moore RE, Nagle DG, Park PU, Biggs J, Paul VJ, Mooberry SL, Corbett TH, Valeriote FA (1998) Isolation, structure determination, and biological activity of dolastatin 12 and linyngyastatin 1 from *Lynghya majuscula* / *Schizothrix calcicola* cyanobacterial assemblages. *J Nat Prod* 61: 1221–1225
- Harrigan GG, Luesch H, Yoshida WY, Moore RE, Nagle DG, Paul VJ (1999) Symplostatin 2: a dolastatin 13 analogue from marine cyanobacterium *Symploca hydnoidea*. *J Nat Prod* 62: 655–658
- Hart CP, Martin JE, Reed MA, Keval AA, Pustelnik MJ, Northrop JP, Patel VD, Grove JR (1999) Potent inhibitory ligands of the GRB2 SH2 domain from recombinant peptide libraries. *Cell Signal* 11: 453–464
- Haubner R, Finsinger D, Kessler H (1997) Stereoisomeric peptides libraries and peptidomimetics for designing selective inhibitors of the $\alpha\beta 3$ integrin for a new cancer therapy. *Angew Chem Int Ed Engl* 36: 1375–1389
- Haviv F, Henkin J, Bradley MF, Kalvin DM (2001a) Preparation of N-alkylated peptides having antiangiogenic activity. Patent WO 01 38,397. See Chem Abstr 135: 5822k
- Haviv F, Henkin J, Bradley MF, Kalvin DM, Schneider AJ (2001b) Preparation of peptides having antiangiogenic activity. Patent WO 01 38,347. See Chem Abstr 135: 5821j
- Hawkins CJ, Lavin MF, Marshall KA, van den Brenk AL, Watters DJ (1990) Structure-activity relationships of the lissoclinamides: cytotoxic cyclic peptides from the ascidian *Lissoclinum patella*. *J Med Chem* 33: 1634–1638
- Hayakawa Y, Adachi H, Kim JW, Shin-Ya K, Seto H (1998) Adenopeptin, a new apoptosis inducer in transformed cells from *Chrysosporium* sp. *Tetrahedron* 54: 15871–15878
- Hayashi S, Kurdowska A, Cohen AB, Stevens MD, Fujisawa N, Miller EJ (1997) A synthetic peptide inhibitor for alpha-chemokines inhibits the growth of melanoma cell lines. *J Clin Invest* 99: 2581–2587
- Hayashi Y, Orikasa S, Tanaka K, Kanoh K, Kiso Y (2000) Total synthesis of anti-microtubule diketopiperazine derivatives: Phenylahistin and aurantiamine. *J Org Chem* 65: 8402–8405
- Heasley LE (2001) Autocrine and paracrine signalling through neurotrophin receptors in human cancer. *Oncogene* 20: 1563–1569
- Helbecque N, Bernier JL, Hénichart JP (1985) Design of a new DNA-polyintercalating drug, a bisacridinyl peptidic analogue of triostin A. *Biochem J* 225: 829–832
- Helmreich EJM (2001) The biochemistry of cell signalling. New York, Oxford University Press
- Heppeler A, Froidevaux S, Eberle AN, Maecke HR (2000) Receptor targeting for tumor localisation and therapy with radiopeptides. *Curr Med Chem* 7: 971–994
- Hirota A, Suzuki A, Aizawa K, Tamura S (1973) Structure of cyl-2, a novel cyclotetrapeptide from *Cylindrocodium scoparium*. *Agric Biol Chem* 37: 955–956
- Hitotsuyanagi Y, Anazawa Y, Yamagishi T, Samata K, Ichihara T, Nanaumi K, Okado N, Nakaike S, Mizumura M, Takeya K, Itokawa H (1997) Novel water-soluble analogues retaining potent antitumor activity of RA-VII, a cyclic hexapeptide from *Rubia* plants. *Bioorg Med Chem Lett* 7: 3125–3128
- Hiwasa T, Soeda C, Takanohashi S, Kobayashi H, Suzuki A, Ueyama T, Ike Y (1996) Screening of peptides which suppress the proliferation of transformed cells. *Int J Oncol* 8: 125–129
- Hocart SJ, Jain R, Murphy WA, Taylor JE, Coy DH (1999) Highly potent cyclic disulfide antagonist of somatostatin. *J Med Chem* 42: 1863–1871
- Hochster H, Oratz R, Ettinger DS, Borden E (1999) A phase II study of didemnin B (NSC 325319) in advanced malignant melanoma: an eastern cooperative oncology group study (PB687). *Invest New Drugs* 16: 259–263
- Hoffken K, Kath R (2000) Peptides in Oncology III: Somatostatin and LH-RH analogues. Springer, Berlin Heidelberg New York Tokyo
- Hoefle G, Leibold T, Steinmetz H (2002) Preparation of stereospecific synthetic tubulysins and intermediate. Patent DE 10,008,089. See Chem Abstr 135: 331296s
- Holinger EP, Chittenden T, Lutz RJ (1999) Bak BH3 peptides antagonize Bcl-XL function and induce apoptosis through cytochrome c-independent activation of caspases. *J Biol Chem* 274: 13298–13304
- Hollstein V (1974) Actionmycin. Chemistry and mechanism of action. *Chem Rev* 74: 625–652
- Hozumi M, Ogawa M, Sugimura T, Takeuchi T, Umezawa H (1972) Inhibition of tumorigenesis in mouse skin by leupeptin, a protease inhibitor from *Actinomycetes*. *Cancer Res* 32: 1725–1728
- Huang Z (2000) Bcl-2 family proteins as targets for anticancer drug design. *Oncogene* 19: 6627–6631
- Huang Z, Lui D, Han X, Zhang Z, Wang J (2000) Small molecule inhibitors of Bcl-2 proteins for inducing apoptosis. Patent WO 00 04,901. See Chem Abstr 132: 117566z
- Huber HE, Koblan KS, Heimbrook DC (1994) Protein-protein interactions as therapeutic targets for cancer. *Curr Med Chem* 1: 13–34

- Huirne JA, Lambalk CB (2001) Gonadotropin-releasing-hormone-receptor antagonists. *Lancet* 358: 1793–1803
- Humphries MJ, Olden K, Yamada KM (1986) A synthetic peptide from fibronectin inhibits experimental metastasis of murine melanoma cells. *Science* 233: 467–470
- Hupp TR, Lane DP, Ball KL (2000) Strategies for manipulating the p53 pathway in the treatment of human cancer. *Biochem J* 352: 1–17
- Hynes RO (1992) Integrins: versatility, modulation, and signalling in cell adhesion. *Cell* 69: 11–25
- Ijaz T, Tran P, Ruparelia KC, Teesdale-Spittle PH, Orr S, Patterson LH (2001) Anthraquinonepeptides as inhibitors of AP-1 transcription factor. *Bioorg Med Chem Lett* 11: 351–353
- Ike Y, Kobayashi A, Suzuki A, Hiwasa S (1997) Antitumor agents containing tripeptides or dipeptides. Patent JP 09,40,577. See Chem Abstr 126: 233697t
- Ikegami S, Miyashiro S, Saki T, Shibai H (1987) Manufacture of antitumor polypeptides. Patent JP 61,93,125. See Chem Abstr 106: 31371h
- Ireland CM, Durso AR, Newman RA, Hacker MP (1982) Antineoplastic cyclic peptides from the marine tunicate *Lissoclinum patella*. *J Org Chem* 47: 1807–1811
- Ishida K, Nakagawa H, Murakami M (2000) Microcyclamide, a cytotoxic cyclic hexapeptide from the cyanobacterium *Microcystis aeruginosa*. *J Nat Prod* 63: 1315–1317
- Ishiwata H, Nemoto T, Ojika M, Yamada K (1994a) Isolation and stereostructure of dolicolide, a cytotoxic cyclodepsipeptide from the Japanese sea hare *Dolabella auricularia*. *J Org Chem* 59: 4710–4711
- Ishiwata H, Sone H, Kigoshi H, Yamada K (1994b) Enantioselective total synthesis of dolicolide a potent cytotoxic cyclodepsipeptide of marine origin and structure-cytotoxicity relationships of synthetic dolicolide congeners. *Tetrahedron* 50: 12853–12882
- Ishiwata H, Sone H, Kigoshi H, Yamada Y (1994c) Total synthesis of dolicolide, a potent cytotoxic cyclodepsipeptide from the Japanese sea hare *Dolabella auricularia*. *J Org Chem* 59: 4712–4713
- Isoai A, Giga-Hama Y, Shinkai K, Mukai M, Akedo H, Kumagai H (1992) Tumor invasion-inhibiting factor 2: primary structure and inhibitory effect on invasion in vitro and pulmonary metastasis of tumor cells. *Cancer Res* 52: 1422–1426
- Itagaki F, Shigemori H, Ishibashi M, Nakamura T, Sasaki T, Kobayashi J (1992) Keramamide F, a new thiazole-containing peptide from the okinawan marine sponge *Theonella sp.* *J Org Chem* 57: 5540–5542
- Itazaki H, Nagashima K, Sugita K, Yoshida H, Kawamura Y, Yasuda Y, Matsumoto K, Ishii K, Uotani N, Nakai I, Terui A, Yoshimatsu S, Ikenishi Y, Nakagawa Y (1990) Isolation and structural elucidation of new cyclotetrapeptides, trapoxins A and B, having detransformation activities as antitumor agents. *J Antibiot* 43: 1524–1532
- Itokawa H, Takeya K (1993) Antitumor substances from higher plants. *Heterocycles* 35: 1467–1487
- Iwamoto Y, Robey FA, Graf J, Sasaki M, Kleinman HK, Yamada Y, Martin GR (1987) YIGSR, a synthetic laminin pentapeptide, inhibits experimental metastasis formation. *Science* 238: 1132–1134
- Jacob L, Zasloff M (1994) Potential therapeutic applications of magainins and other microbial agents of animal origin. Ciba Foundation Symposium 186: 197–223
- Janssen B, Barlozzari T, Haupt A, Zierke T, Kling A (1998) Preparation of dolastatin 15 derivatives as antitumor agents. Patent WO 99 03,879. See Chem Abstr 130: 139656q
- Jensen PKA, Elgjo K, Laerum OD, Bolund L (1990) Synthetic epidermal pentapeptide and related growth regulatory peptides inhibit proliferation and enhance differentiation in primary and regenerating cultures of human epidermal keratinocytes. *J Cell Sci* 97: 51–58
- Jia H, Jezequel S, Lohr M, Shaikh S, Davis D, Soker S, Selwood D, Zachary I (2001) Peptides encoded by exon 6 of VEGF inhibit endothelial cell biological responses and angiogenesis induced by VEGF. *Biochem Biophys Res Commun* 283: 164–173
- Jiang TL, Liu RH, Salmon SE (1983) Antitumor activity of didemnin B in the human tumor stem cell assay. *Cancer Chemother Pharmacol* 11: 1–4
- Jiang G, Stalewski J, Galyean R, Dykert J, Schteingart C, Broqua P, Aebi A, Aubert ML, Semple G, Robson P, Akinsanya K, Haigh R, Riviere P, Trojnar J, Junien JL, Rivier JE (2001) GnRH antagonists: a new generation of long acting analogues incorporating p-ureido-phenylalanines at positions 5 and 6. *J Med Chem* 44: 453–467
- Jimeno JM (2002) A clinical armamentarium of marine-derived anti-cancer compounds. *Anti-Cancer Drugs* 13: S15–S19
- Johnson LL, Dyer R, Hupe DJ (1998) Matrix metalloproteinases. *Curr Opin Chem Biol* 2: 466–471
- Jordan MA, Wilson L (1998) Microtubules and actin filaments: dynamic targets for cancer chemotherapy. *Curr Opin Cell Biol* 10: 123–130
- Jordan MA, Walker D, De Arruda M, Barlozzari T, Panda D (1998) Suppression of microtubule dynamics by binding of cemadotin to tubulin: possible mechanism for its antitumor action. *Biochemistry* 37: 17571–17578
- Jouin P, Poncet J, Dufour M-N, Aumelas A, Pantaloni A, Cros S, François G (1991) Antineoplastic activity of didemnin congeners: nordidemnin and modified chain analogues. *J Med Chem* 34: 486–491
- Kaiser M, Groll M, Renner C, Huber R, Moroder L (2002) The core structure of TMC-95A is a promising lead for reversible proteasome inhibition. *Angew Chem Int Ed* 41: 780–783
- Kamenecka TM, Danishefsky SJ (1998) Total synthesis of himastatin: confirmation of the revised stereostructure. *Angew Chem Int Ed Engl* 37: 2995–2998
- Kaneda Y, Yamamoto Y, Okada N, Tsutsumi Y, Nakagawa S, Kakiuch M, Maeda M, Kawasaki K, Mayumi T (1997) Antimetastatic effect of synthetic Glu-Ile-Leu-Asp-Val peptide derivatives containing D-amino acids. *Anti-Cancer Drugs* 8: 702–707
- Kanoh K, Kohno S, Katada J, Hayashi Y, Muramatsu M, Uno I (1999a) Antitumor activity of phenylhistin *in vitro* and *in vivo*. *Biosci Biotechnol Biochem* 63: 1130–1133
- Kanoh K, Kohno S, Katada J, Takahashi J, Uno I (1999b) (-)-Phenylhistin arrests cells in mitosis by inhibiting tubulin polymerization. *J Antibiot* 52: 134–141
- Kanoh K, Kohno S, Katada J, Takahashi J, Uno I, Hayashi Y (1999c) Synthesis and biological activities of phenylhistin derivatives. *Bioorg Med Chem* 7: 1451–1457
- Kardinal C, Konkol B, Schulz A, Posern G, Lin H, Adermann K, Eulitz M, Estrov Z, Talpaz M, Arlinghaus RB, Feller SM (2000) Cell-penetrating SH3 domain blocker peptides inhibit proliferation of primary blast cells from CML patients. *FASEB J* 14: 1529–1538
- Kavarana MJ, Kovaleva EG, Creighton DJ, Wollman MB, Eisean JL (1999) Mechanism-based competitive inhibitors of glyoxalase I: intracellular delivery, *in vitro* antitumor activities and stabilities in human serum and mouse serum. *J Med Chem* 42: 221–228
- Kawasaki K, Maeda M, Inoue S, Yamashiro Y, Kaneda Y, Mu Y, Tsutsumi Y, Nakagawa S, Mayumi T (1996) Amino acids and peptides. XXIX. Synthesis and antimetastatic effects of peptides and peptide-poly(ethylene glycol) hybrids related to the core

- sequence of the type III connecting segment domain of fibronectin. *Biol Pharm Bull* 19: 1574–1579
- Keller-Schierlein W, Mihailovic ML, Prelog V (1959) Stoffwechselprodukte von Actinomyceten. *Helv Chem Acta* 42: 305–322
- Kéri G, Mező I, Vadsz Z, Horváth A, Idei M, Vántus T, Balogh A, Bökönyi G, Bajor T, Teplan I, Tamás J, Mák M, Horváth J, Csuka O (1993) Structure-activity relationship studies of novel somatostatin analogs with antitumor activity. *Pept Res* 6: 281–288
- Khachigian LM, Field SL, Crouch R, Chesterman CN (1995) Platelet-derived factor A-chain synthetic peptide inhibits human glioma xenograft proliferation in nude mice. *Anticancer Res* 15: 337–341
- Khavinson VK, Anisimov VN (2000) Synthetic pineal peptide increase a life span and inhibits a development of tumors in mice. *Dokl. Akad. Nauk.* 373: 567–569. See Chem Abstr 134: 95241b
- Khiat A, Lamoureux M, Boulanger Y (1996) Structural differences between the free and bound states of the DNA-bisintercalating peptide YSPTSPSY *J Med Chem* 39: 2492–2498
- Kiaris H, Schally AV, Sun B, Armatas P, Groot K (1999) Inhibition of growth of human malignant glioblastoma in nude mice by antagonists of bombesin/gastrin-releasing peptide. *Oncogene* 18: 7168–7173
- Kijima J, Yoshida M, Sugita K, Horinouchi S, Beppu T (1993) Trapoxin, an antitumor cyclic tetrapeptide is an irreversible inhibitors of mammalian histone deacetylase. *J Biol Chem* 268: 22429–22435
- Kim SE, Kim HH, J.Y. K, Kang YI, Woo HJ, Lee HJ (2000) Anticancer activity of hydrophobic peptides from soy proteins. *BioFactors* 12: 151–155
- Kim SS, Kim YH, Park MG, Park EG, Yeo UH, Lee SC, Lee YH, Chae SY (2002) Novel peptidyl antibiotic produced from ascomycetes *Apiocra* sp. 14t strain, preparation method and use thereof. Patent KR 2000 56,350. See Chem Abstr 136: 324169v
- Kineman RD (2000) Antitumorogenic actions of growth hormone-releasing hormone antagonists. *Proc Natl Acad Sci USA* 97: 532–534
- Kini RM, Evans HJ (1995) A hypothetical structural role for proline residues in the flanking segments of protein-protein interaction sites. *Biochem Biophys Res Commun* 212: 1115–1124
- Kitagawa I, Kobayashi S (1996) Isolation of arenastatin A having antitumor activity from *Dysidea arenaria* marine sponge and its total synthesis. patent JP 08,92,232. See Chem Abstr 125: 115144s
- Kobayashi H, Ohi H, Shinohara H, Sugimura M, Fujii T, Terao T, Schmitt M, Goretzki L, Chochołowski N, Jänicke F, Graeff H (1993) Saturation of tumor cell surface receptors for urokinase-type plasminogen activator by amino-terminal fragment of and subsequent effect on reconstituted basement membranes invasion. *Br J Cancer* 67: 537–544
- Kobayashi H, Gotoh J, Fujie M, Shinohara H, Moniwa N, Terao T (1994a) Inhibition of metastasis of lewis lung carcinoma by a synthetic peptide within growth factor-like domain of urokinase in the experimental and spontaneous metastasis mode. *Int J Cancer* 57: 727–733
- Kobayashi M, Kurosu M, Ohyabu N, Wang W, Fujii S, Kitagawa I (1994b) The absolute stereostructure of arenastatin A, a potent cytotoxic depsipeptide from the Okinawan marine sponge *Dysidea arenaria*. *Chem Pharm Bull* 42: 2196–2198
- Kobayashi M, Wang W, Ohyabu N, Kurosu M, Kitagawa I (1995) Improved total synthesis and structure-activity relationship of arenastatin A, a potent cytotoxic spongean depsipeptide. *Chem Pharm Bull* 43: 1598–1600
- Kobayashi M, Natsume T, Tamaoki S, Watanabe J, Asano H, Mikami T, Miyasaka K, Miyasaki K, Gondo M, Sakakibara K, Tsukagoshi S (1997) Antitumor activity of TZT-1027, a novel dolastatin 10 derivative. *Jpn J Cancer Res* 88: 316–327
- Kobayashi J, Suzuki H, Shimbo K, Takeya K, Morita H (2001) Celogentins A-C, new antimitotics bicyclic peptides from the seeds of *Celosia argentea*. *J Org Chem* 66: 6626–6633
- Koehn FE, Longley RE, Reed JK (1992) Microcolins A and B, new immunosuppressive peptides from the blue-green alga *Lyngbya majuscula*. *J Nat Prod* 55: 613–619
- Koguchi Y, Nishio M, Suzuki S-I, Takahashi K, Ohnuki T, Komatsubara S (2000) TMC-89A and B, new proteasome inhibitors from *Streptomyces* sp. TC 1087. *J Antibiot* 53: 967–972
- Kohda K, Ohta Y, Yokoyama Y, Kawazoe Y, Kato T, Suzumura Y, Hamada Y, Shioiri T (1989) Mechanistic aspects of the cytotoxic action of ulithiacyclamide on mouse leukemia L1210 cells in vitro. *Biochem Pharmacol* 38: 4497–4500
- Kohen F, Natarajan V, Kasher R, Fridkin M, Katchalskikatzir E (2002) Peptides mimicking the biological activity of steroids hormones, and their therapeutic uses. Patent WO 01 74,846. See Chem Abstr 135: 283229y
- Kohno J, Koguchi Y, Nishio M, Nakao K, Kuroda M, Shimizu R, Ohnuki T, Komatsubara S (2000) Structure of TMC-95A-D: novel proteasome inhibitors from *Apiospora montagnei* Sacc. TC 1093. *J Org Chem* 65: 990–995
- Koivunen E, Arap W, Valtanen H, Rainisalo A, Medina OP, Heikkilä P, Kantor C, Gahmberg C, Salo T, Kontinen YT, Sorsa T, Ruoslahti E, Pasqualini R (1999) Tumor targeting with a selective gelatinase inhibitor. *Nat Biotechnol* 17: 768–774
- Komatsu Y, Tomizaki KY, Tsukamoto M, Kato T, Nishino N, Sato S, Yamori T, Tsuruo T, Furumai R, Yoshida M, Horinouchi S, Hayashi H (2001) Cyclic hydroxamic-acid-containing peptide 31, a potent synthetic histone deacetylase inhibitor with antitumor activity. *Cancer Res* 61: 4459–4466
- Konishi M, Ohkuma H, Sakai F, Tsuno T, Koshiyama H, Naito T, Kawaguchi H (1981) BBM-928, a new antitumor antibiotic complex III. Structure determination of BBM-928 A, B and C. *J Antibiot* 34: 148–159
- Kouzarides T (1998) Method and means for disruption of p53 and RB interaction. Patent WO 97 41,433. See Chem Abstr 128: 18684x
- Krulich L, Dhariwal APS, McCann SM (1968) Stimulatory and inhibitory effects of purified hypothalamic extracts on growth hormone release from pituitary *in vitro*. *Endocrinology* 83: 783–790
- Kutscher B, Bernd M, Beckers T, Polymeropoulos EE, Engel J (1997) Chemistry and molecular biology in the search for new LHRH antagonists. *Angew. Chem Int Ed Engl* 36: 2149–2161
- Kwekkeboom D, Krenning EP, de Jong M (2000) Peptide receptor imaging and therapy. *J Nucl Med* 41: 1704–1713
- Kwon HJ, Kim MS, Kim MJ, Nakajima H, Kim KW (2002) Histone deacetylase inhibitor FK228 inhibits tumor angiogenesis. *Int J Cancer* 97: 290–296
- Lacey E, Edgar JA, Culvenor CCJ (1987) Interaction of phalloidin A and related compounds with purified sheep brain tubulin. *Biochem Pharmacol* 36: 2133–2138
- Laerum OD (1990) Preparation of pentapeptides and derivatives as stem cell proliferation inhibitors. Patent EP 359,338. See Chem Abstr 113: 115878m
- Lam KS, Gustavson DR, Hesler GA, Dabrah TT, Matson JA, Berry RL, Rose WC, Foreza S (1995) Korkomicins, novel depsipeptide antitumor antibiotics from *Micromonospora* sp C39500: fermentation, precursor directed biosynthesis and biological activities. *J Ind Microbiol* 15: 60–65
- Lamberts SW, van der Lely AJ, de Herder WW, Hofland LJ (1996) Octreotide. *New England J Med* 334: 246–254
- Lane DP, Hall PA (1997) MDM2-arbiter of p53's destruction. *Trends Biochem Sci* 22: 372–374

- Langdon S, Sethi T, Ritchie A, Muir M, Smyth J, Rozenfurt E (1992) Broad spectrum neuropeptide antagonists inhibit the growth of small cell lung cancer *in vivo*. *Cancer Res* 52: 4554–4557
- Larsen IK (1996) A Textbook of Drug Design and Development. In: Krosggaard-Larsen P, Liljefors T, Madsen U (eds) A Textbook of drug design and development. Harwood Academic, pp 460–506
- Lenfant M, Wdzieczak-Balaka J, Guittet E, Prome JC, Sotty D, Fringel E (1989) Inhibitor of hematopoietic pluripotent stem cell proliferation: purification and determination of its structure. *Proc Natl Acad Sci USA* 86: 779–782
- Leonard DM, Sebolt-Leopold JS (1999) Ras farnesyltransferase inhibitors. *Drugs Future* 24: 1099–1106
- Leung D, Abbenante G, Fairlie DP (2000) Protease inhibitors: current status and future prospect. *J Med Chem* 43: 305–341
- Leyton J, Coelho T, Coy DH, Jakowlew S, Birrer MJ, Moody TW (1998) PACAP(6-38) inhibits the growth of prostate cancer cells. *Cancer Lett* 125: 131–139
- Li H, Matsunaga S, Fusetani N (1995a) Halicyclindramides A-C, antifungal and cytotoxic depsipeptides from the marine sponge *Halichondria cylindrata*. *J Med Chem* 38: 338–343
- Li Y, Koiso Y, Kobayashi H, Hashimoto Y, Iwasaki S (1995b) Ustiloxins, new antimitotic cyclic peptides: interaction with porcine brain tubulin. *Biochem Pharmacol* 49: 1367–1372
- Li H, Matsunaga S, Fusetani N (1996) Halicyclindramides D and E, antifungal peptides from the marine sponge *Halichondria cylindrata*. *J Nat Prod* 59: 163–166
- Liakopoulou-Kyriakides M, Stavropoulos G, Geromichalos G, Papazisis KT, Kortsaris AH, Kyriakidis DA (1998) Antiproliferative activity of synthetic tetrapeptides, analogs of AS-I phytoalexin, towards cancer cell lines. *Anti-Cancer Drugs* 9: 175–179
- Liehr S, Barbosa J, Smith AB, Cooperman BS (1999) Synthesis and biological activity of cyclic peptide inhibitors of ribonucleotide reductase. *Org Lett* 1: 1201–1204
- Liesch JM, Sweeley CC, Staffeld GD, Anderson MS, Weber DJ, Scheffer RP (1982) Structure of HC-toxin, a cyclic tetrapeptide from *Helminthosporium carbonum*. *Tetrahedron* 38: 45–48
- Lin JT, Coy DH, Mantey SA, Jensen RT (1995) Comparison of the peptide structural requirements for high affinity interaction with bombesin receptors. *Eur J Pharmacol* 294: 55–69
- Lingham RB, Hsu AHM, O'Brien JA, Sigmund JM, Sanchez M, Gagliardi MM, Heimbuch BK, Genilloud O, Martin I, Diez MT, Hirsch CF, Zink DL, Liesch JM, Koch GE, Garter SE, Garrity GM, Tsou NN, Salituro GM (1996) Quinoxapeptins: Novel chromodepsipeptide inhibitors of HIV-1 and HIV-2 reverse transcriptase. *J Antibiot*. 49: 253–259
- Liu D-G, Yao Z-J, Gao Y, Burke TR (2000a) Large scale preparation of cell permeable nonphosphate-containing Grb2 SH2 domain inhibitors. *Org Prep Proced Int* 32: 197–201
- Liu W-Q, Vidal M, Mathé C, Périgaud C, Garbay C (2000b) Inhibition of the ras-dependant mitogenic pathway by phosphopeptide prodrugs with antiproliferative properties. *Bioorg Med Chem Lett* 10: 669–672
- Liu Y, Elgjo K, Wright M, Reichelt KL (2000c) Novel lymphocyte growth-inhibiting tripeptide: N-acetyl-Glu-Ser-Gly-NH₂. *Biochem Biophys Res Commun* 277: 562–567
- Loffet A (2002) Peptides as drugs: is there a market? *J Pept Sci* 8: 1–7
- Long Y-Q, Voigt JH, Lung F-D, King CR, Roller PP (1999) Significant compensatory role of position Y-2 conferring high affinity to non-phosphorylated inhibitors of GRB2-SH2 domain. *Bioorg Med Chem Lett* 9: 2267–2272
- López-Macià A, Jiménez JC, Royo M, Giralt E, Albericio F (2001) Synthesis and structure determination of kahalalide F (1,2). *J Am Chem Soc* 123: 11398–11401
- Ludueña RF, Roach MC, Prasad V, Banerjee M, Koiso Y, Li Y, Iwasaki S (1994) Interaction of ustiloxin A with bovine brain tubulin. *Biochem Pharmacol* 47: 1593–1599
- Luesch H, Yoshida WY, Moore RE, Paul VJ (1999) Lyngbyastatin 2 and norlungbyastatin 2, analogues of Dolastatin G and nordolastatin G from the marine cyanobacterium *Lyngbya majuscula*. *J Nat Prod* 62: 1702–1706
- Luesch H, Moore RE, Paul VJ, Mooberry SL, Corbett TH (2001a) Isolation of dolastatin 10 from the marine cyanobacterium *Symploca* species VP642 and total stereochemistry and biological evaluation of its analogue symplostatin 1. *J Nat Prod* 64: 907–910
- Luesch H, Pangilinan R, Yoshida WY, Moore RE, Paul VJ (2001b) Pitipeptolides A and B, new cyclodepsipeptides from the marine cyanobacterium *Lyngbya majuscula*. *J Nat Prod* 64: 304–307
- Luesch H, Yoshida WY, Moore RE, Paul VJ, Corbett TH (2001c) Total structure determination of apratoxin A, a potent novel cytotoxin from the marine cyanobacterium *Lyngbya majuscula*. *J Am Chem Soc* 123: 5418–5423
- Luesch H, Yoshida WY, Moore RE, Paul VJ (2002) New apratoxins of marine cyanobacterial origin from Guam and Palau. *Bioorg Med Chem* 10: 1973–1978
- Luke RWA, Hudson K, Hayward CF, Fiedling C, Cotton R, Best R, Giles MB, Veldman MH, Griffiths LA, Jewsbury PJ, Breeze AL, Embrey KJ (1999) Design and synthesis of small molecule inhibitors of the MDM2-p53 interaction as potential anti-tumor agents. *Proc Am Assoc Cancer Res* 40: 622
- Lung FD, Long YQ, Roller PP, King CR, Varady J, Wu XW, Wang S (2001) Functional preference of the constituent amino acid residues in a phage-library-based nonphosphorylated inhibitor of the Grb2-SH2 domain. *J Pept Res* 57: 447–454
- Lunney EA, Para KS, Rubin JR, Humblet C, Fergus JH, Marks JS, Sawyer TK (1997) Structure-based design of a novel series of nonpeptide ligands that bind to the pp60src SH2 domain. *J Am Chem Soc* 119: 12471–12476
- Lupu R, Lippman M (1994) Peptides that bind ligands of the epidermal growth factor receptor and erbB-2-receptor. Patent WO 93 22,339. See Chem Abstr 120: 209532r
- Luttrell DK, Lee A, Lansing TJ, Crosby RM, Jung KD, Willard D, Luther M, Rodriguez M, Berman J, Gilmer TM (1994) Involvement of pp60c-src with two major signalling pathways in human breast cancer. *Proc Natl Acad Sci USA* 91: 83–87
- Madden T, Tran HT, Beck D, Huies R, Newman RA, Pusztai L, Wright JJ, Abbruzzese JL (2000) Novel marine-derived anticancer agents: a phase I clinical, pharmacological, and pharmacodynamic study of dolastatin 10 (NSC 376128) in patients with advanced solid tumors. *Clin. Cancer Res* 6: 1293–1301
- Maehr H, Liu C, Palleroni NJ, Smallheer J, Todaro L, Williams TH, Blount JF (1986) Microbial Products VIII. Azinotricin, a novel hexadepsipeptide antibiotic. *J Antibiot* 39: 17–25
- Maeshima Y, Yerramalla UL, Dhanabal M, Holthaus KA, Barbashov S, Kharbanda S, Reimer C, Manfredi M, Dickerson WM, Kalluri R (2001) Extracellular matrix-derived peptide binds to $\alpha\beta 3$ integrin and inhibits angiogenesis. *J Biol Chem* 276: 31959–31968
- Magnifico A, Tagliabue E, Butó S, Ardini E, Castronovo V, Colnaghi MI, Ménard S (1996) Peptide G, containing the binding site of the 67-kDa laminin receptor, increases and stabilizes laminin binding to cancer cells. *J Biol Chem* 271: 31179–31184
- Maini A, Morse PD, Wang CY, Jones RF, Haas GP (1997) New developments in the use of cytokines for cancer therapy. *Anti-cancer Res* 17: 3803–3808

- Mamber SW, Brookshire KW, Dean BJ, Firestone RA, Leet JE, Matson JA, Forenza S (1994) Inhibition of antibacterial activity of himastatin, a new antitumor antibiotic from *Streptomyces hygroscopicus* by fatty acid sodium salts. *Antimicrob Agents Chemother* 38: 2633–2642
- Manderville RA, Ellena JF, Hecht SM (1994) Solution structure of a Zn(II)-bleomycin A5-d(CGCTAGCG)₂ complex. *J Am Chem Soc* 116: 10851–10852
- Maroun JA, Stewart D, Verma S, Eisenhauer E (1998) Phase I clinical study of didemnin B. *Invest New Drugs* 16: 51–56
- Marsili V, Nardicchi V, Lupidi G, Brozzetti A, Gianfranceschi GL (1996) Dansylated octapeptide Dns-Glu-Asp-Asp-Ser-Asp-Glu-Glu-Asn inhibits the proliferation rate of HL-60 cells. *Can J Physiol Pharmacol* 74: 1302–1307
- Martin DG, Mizsak SA, Biles C, Stewart JC, Baczynskyj L, Meulman PA (1975) Structure of quinomycin antibiotics. *J Antibiot* 28: 332–336
- Martin GR, Sasaki M, Yamada Y, Kleinman HK, Robey F, Iwamoto Y, Graf JO (1989) Peptides with laminin activity. Patent EP 278,781. See Chem Abstr 111: 109027a
- Masuoka Y, Shin-Ya K, Kim YB, Yoshida M, Nagai K, Suzuki K-I, Hayakawa Y, Seto H (2000) Diheteropeptin, a new substance with TGF- β -like activity, produced by a fungus, *Diheterospora chlamydosporia*. Part 1. Production, Isolation and biological activities. *J Antibiot* 53: 788–792
- Masuoka Y, Nagai A, Shin-Ya K, Furihata K, Nagai K, Suzuki KI, Hayakawa Y, Seto H (2001) Spiruchostatins A and B, novel gene expression-enhancing substances produced by *Pseudomonas sp.* *Tetrahedron Lett* 42: 41–44
- Matson JA, Bush JA (1989) Sandramycin, a novel antitumor antibiotic produced by a *Nocardioidea sp.* *J Antibiot* 42: 1763–1767
- Matson JA, Colson KL, Belofsky GN, Bleiberg BB (1993) Sandramycin, a novel antitumor antibiotic produced by a *Nocardioidea sp.* II. Structure determination. *J Antibiot* 46: 162–166
- Matsuda M, Asakura S, Myuura EU (1996) Cell adhesion inhibitory peptides derived from human high-molecular-weight kininogen L-chain. Patent JP 08,208,692. See Chem Abstr 125: 276595e
- Matsumoto Y, Kawatani M, Simizu S, Tanaka T, Takada M, Imoto M (2000) Bcl-2-independent induction of apoptosis by neuropeptide receptor antagonist in human small cell lung carcinoma cells. *Anticancer Res* 20: 3123–3129
- Matsunaga S, Fusetani N (1995) Theonellamides A-E, cytotoxic bicyclic peptides, from a marine sponge *Theonella sp.* *J Org Chem* 60: 1177–1181
- Matsunaga S, Fusetani N, Konosu S (1984) Bioactive marine metabolites VI. Structure elucidation of discodermin A, an antimicrobial peptide from the marine sponge *Discodermia kiiensis*. *Tetrahedron Lett* 25: 5165–5168
- Matsunaga S, Fusetani N, Konosu S (1985a) Bioactive marine metabolites VII. Structures of discodermins B, C and D antimicrobial peptides from the marine sponge *Discodermia kiiensis*. *Tetrahedron Lett* 26: 855–856
- Matsunaga S, Fusetani N, Konosu S (1985b) Bioactive marine metabolites IV. Isolation and the amino acid composition of discodermin A, an antimicrobial peptide, from the marine sponge *Discodermia kiiensis*. *J Nat Prod* 48: 236–241
- Mattock H, Lane DP, Warbrick E (2001) Inhibition of cell proliferation by the PCNA-binding region of p21 expressed as a GFP miniprotein. *Exp Cell Res* 265: 234–241
- Mau CMS, Nakao Y, Yoshida WY, Scheuer PJ, Kelly-Borges M (1996) Waiakeamide, a cyclic hexapeptide from the sponge *Ircinia dendroides*. *J Org Chem* 61: 6302–6304
- Mazar AP (2001) The urokinase plasminogen activator receptor (uPAR) as a target for the diagnosis and therapy of cancer. *Anti-cancer Drugs* 12: 387–400
- McDonald III ER, El-Deiry WS (2000) Cell cycle control as a basis for cancer drug development (review). *Int J Oncol* 16: 871–886
- McElroy Jr EA, Pitot HC, Erlichman C, Reid JM, Ames MM, Windebank AJ, Sloan JA, Rubin J (1997) Phase I trial of dolastatin 10 in patients with advanced solid tumors. *Proc Am Soc Clin Oncol* 16: 223a
- McKelvey DR, Brooks CL, Mokotoff M (1991) A CHARMM analysis of the conformations of the metastasis-inhibiting laminin pentapeptide. *J Protein Chem* 10: 265–271
- Medina OP, Söderlund T, Laakkonen LJ, Tuominen EK, Koivunen E, Kinnunen PK (2001) Binding of novel peptide inhibitors of type IV collagenases to phospholipid membranes and use in liposome targeting to tumor cells in vitro. *Cancer Res* 61: 3978–3985
- Mesfin FB, Andersen TT, Jacobson HI, Zhu S, Bennett JA (2001) Development of a synthetic cyclized peptide derived from α -fetoprotein that prevents the growth of human breast cancer. *J Pept Res* 58: 246–256
- Middley CA, Desterro JM, Saville MK, Howard S, Sparks A, Hay RT, Lane DP (2000) An N-terminal p14ARF peptide blocks Mdm2-dependent ubiquitination *in vitro* and can activate p53 *in vivo*. *Oncogene* 19: 2312–2323
- Mikhailova AA, Gerasimova GK, Treshchalina EM, Strelkov LA, Fonina LA, Gur'yanov SA (1998) Antitumor action of myelopoietin-2. *Russ Khim Zh* 42: 176–178. See Chem Abstr 132: 30429h
- Milne PJ, Hunt AL, Rostoll K, Van Der Walt JJ, Graz CJM (1998) The biological activity of selected cyclic dipeptides. *J Pharm Pharmacol* 50: 1331–1337
- Mishima K, Mazar AP, Gown A, Skelly M, Ji X-D, Wang X-D, Jones FR, Cavenee WK, Huang H-JS (2000) A peptide derived from the non-receptor binding region of urokinase plasminogen activator inhibits glioblastoma growth and angiogenesis *in vivo* in combination with cisplatin. *Proc Natl Acad Sci USA* 97: 8484–8489
- Mittelman A, Chun HG, Puccio C, Coombe N, Lansen T, Ahmed T (1999) Phase II clinical trial of didemnin in patients with recurrent or refractory anaplastic astrocytoma or glioblastoma multiform (NSC 325319). *Invest New Drugs* 17: 179–182
- Miyamoto Y, Kuroda M, Munekata E, Masaki T (1986) Stoichiometry of actin and phalloidin binding: one molecule of the toxin dominates two actin subunits. *J Biochem* 100: 1677–1680
- Mizejewski GJ (2001) Peptides as receptor ligand drugs and their relationship to G-coupled signal transduction. *Expert Opin Invest Drugs* 10: 1063–1073
- Mizuguchi H, Kubomi T, Nomura R, Yasukawa K, Imanaka T, Takagi M (2000) Screening of an oligopeptide antagonist for interleukin-6 from a random phage library. *Biotechnol Lett* 22: 1015–1020
- Molineaux CJ, Sluss PM, Bree MP, Gefter ML, Sullivan LM, Garnick MB (1998) Suppression of plasma gonadotropins by anarelix, a potent new LHRH antagonist. *Mol Urol* 2: 265–268
- Montgomery AMP, Reisfeld RA, Cheresch DA (1994) Integrin $\alpha v \beta 3$ rescues melanoma cells from apoptosis in 3-dimensional dermal collagen. *Proc Natl Acad Sci USA* 91: 8856–8860
- Mooberry SL, Taoka CR, Busquets L (1996) Cryptophycin 1 binds to tubulin at a site distinct from the colchicine binding site and at a site that may overlap the vinca binding site. *Cancer Lett* 107: 53–57
- Moody TW, Pert CB, Gazdar AF, Carney DN, Minna JD (1981) High levels of intracellular bombesin characterize human small-cell lung carcinoma. *Science* 214: 1246–1248

- Moody TW, Zia F, Draoui M, Brennemen D, Fridkin M, Davidson A, Gozes I (1993) A vasoactive intestinal peptide antagonist inhibits non small cell lung cancer growth. *Proc Natl Acad Sci USA* 90: 4345–4349
- Moore RE, Corbett TH, Patterson GML, Valeriote FA (1996) The search for new antitumor drugs from blue-green algae. *Curr Pharm Des* 2: 317–330
- Moore RE, Entzeroth M (1988) Majusculamide D and deoxymajusculamide D, two cytotoxins from *Lynngbya majuscula*. *Phytochemistry* 27: 3101–3103
- Morgan BA, Sadat-Aalae D (2001) Somatostatin receptor type 5 agonist cyclic peptide, their preparation, and their therapeutic use. Patent WO 01 00,676. See Chem Abstr 134: 80825w
- Mori H, Komazawa H, Saiki I, Azuma I (1994) Preparation of peptide derivatives for inhibition of cancer metastasis. Patent JP 05,97,890. See Chem Abstr 120: 192311t
- Mori H, Komazawa H, Kojima M, Saiki I, Azuma I (1995a) Preparation of peptide derivatives as cancer metastasis inhibitors. Patent JP 06,321,985 and 06,321,987. See Chem Abstr 122: 133866w and 133867x
- Mori H, Komazawa H, Saiki I, Azuma I (1995b) Preparation of pentapeptides for inhibitions of cancer metastasis. Patent JP 06,41,193. See Chem Abstr 122: 10684c
- Mori H, Sakamoto K, Tsurumi Y, Takase S, Hino M (2000) Cyclic tetrapeptide compounds as histone deacetylase and their use in therapy. Patent WO 00 21,979. See Chem Abstr 132: 288796d
- Morita H, Nagashima S, Takeya K, Itokawa H (1993) Astins A and B, antitumor cyclic pentapeptides from *Aster tataricus*. *Chem Pharm Bull* 41: 992–993
- Morita H, Gonda A, Takeya K, Itokawa H (1996a) Cycloleonoripeptide A, B and C, three new proline-rich cyclic nonapeptides from *Leonurus heterophyllus*. *Bioorg Med Chem Lett* 6: 767–770
- Morita H, Kayashita T, Shimomura M, Takeya K, Itokawa H (1996b) Cyclic peptide from higher plants. 24. Yunnanin C, a novel cyclic heptapeptide from *Stellaria yunnanensis*. *J Nat Prod* 59: 280–282
- Morita H, Nagashima S, Uchiumi Y, Kuroki O, Takeya K, Itokawa H (1996c) Cyclic peptides from higher plants. 28. Antitumor activity and hepatic microsomal biotransformation of cyclic pentapeptides, astins, from *Aster tataricus*. *Chem Pharm Bull* 44: 1026–1032
- Morita H, Shimbo K, Shigemori H, Kobayashi J (2000) Antimitotic activity of moroidin, a bicyclic peptide from the seeds of *Celosia argentea*. *Bioorg Med Chem Lett* 10: 469–471
- Mosher C, Goodman L (1972) Synthesis of [4',4'-bis(glycine),5',5'-bis(valine)]actinomycin D, a tetra-N-demethylactinomycin. *J Org Chem* 37: 2928–2933
- Mueller H (1998) Tumor necrosis factor as an antineoplastic agent: pitfalls and promises. *Cell Mol Life Sci* 54: 1291–1298
- Muller R, Kontermann RE, Montigiani S (2000) Transcription factor E2F DNA-binding domain inhibitor peptides and uses thereof. Patent WO 00 44,771. See Chem Abstr 133: 145915b
- Murphy WA, Heiman ML, Lance VA, Mezo I, Coy DH (1985) Octapeptide analogs of somatostatin exhibiting greatly enhanced *in vivo* and *in vitro* inhibition of growth hormone secretion in the rat. *Biochem Biophys Res Commun* 132: 922–928
- Mutou T, Kondo T, Ojika M, Yamada K (1996a) Isolation and stereostructures of dolastatin G and nordolastatin G, cytotoxic 35-membered cyclodepsipeptides from the Japanese sea hare *Dolabella auricularia*. *J Org Chem* 61: 6340–6345
- Mutou T, Kondo T, Shibata M, Ojika M, Kigoshi H, Yamada K (1996b) Synthesis of dolastatin G and nordolastatin G, cytotoxic 35-membered cyclodepsipeptides of marine origin. *Tetrahedron Lett* 37: 7299–7302
- Mutou T, Suenaga K, Fujita T, Itoh T, Takada N, Hayamizu K, Kigoshi H, Yamada K (1997) Enantioselective synthesis of aurilide, a cytotoxic 26-membered cyclodepsipeptide of marine origin. *Synlett* 199–201
- Nagalla SR, Barry BJ, Creswick KC, Eden P, Taylor JT, Spindel ER (1995) Cloning of a receptor for amphibian [Phe13]bombesin distinct from the receptor for gastrin-releasing peptide: identification of a fourth bombesin receptor subtype (BB4). *Proc Natl Acad Sci USA* 92: 6205–6209
- Nakagawa M, Hayakawa Y, Furihata K, Seto H (1990) Structural studies on new depsipeptide antibiotics, variapeptin and citropeptin. *J Antibiot* 43: 477–484
- Nakajima H, Kim YB, Terano H, Yoshida M, Horinouchi S (1998) FR901228, a potent antitumor antibiotic, is a novel histone deacetylase inhibitor. *Exp Cell Res* 241: 126–133
- Namikoshi M, Rinehart KL, Sakai R, Stotts RR, Dahlem AM, Beasley VR, Carmichael WW, Evans WR (1992) Identification of 12 hepatotoxins from a homer lake bloom of the cyanobacteria *Microcystis aeruginosa*, *Microcystis viridis* and *Microcystis wessenbergii*: nine new mycrocystins. *J Org Chem* 57: 866–872
- Nassar N, Horn G, Herrmann C, Scherer A, McCormick F, Wittinghofer A (1995) The 2.2 Å crystal structure of the Ras-binding domain of the Serine/threonine kinase c-Raf1 in complex with Rap1A and a GTP analogue. *Nature* 375: 554–560
- Nestor JJ, Moffatt JG, Sima JM (1985) Nona- and dodecapeptides for augmenting natural killer cell activity. Patent US 4,473,555. See Chem Abstr 102: 113959n
- Nestor JJJ, Tahilramani R, Ho TL, Goodpasture JC, Vickery BH, Ferrandon P (1992) Potent gonadotropin releasing hormone antagonists with low histamine-releasing activity. *J Med Chem* 35: 3942–3948
- Neufeld G, Cohen T, Gengrinovitch S, Poltorak Z (1999) Vascular endothelial growth factor (VEGF) and its receptors. *FASEB J* 13: 9–22
- Nevins JR (2001) The Rb/E2F pathway and cancer. *Human Mol Gen* 10: 699–703
- Nguyen JT, Turck CW, Cohen FE, Zuckermann RN, Lim WA (1998) Exploiting the basis of proline recognition by SH3 and WW domains: design of N-substituted inhibitors. *Science* 282: 2088–2092
- Niehof M, Radziwill G, Klausner S, Moelling K (1995) A small peptide derived from aminoterminal of c-Raf-1 inhibits c-Raf-1/Ras binding. *Biochem Biophys Res Commun* 206: 46–50
- Nogle LM, Williamson RT, Gerwick WH (2001) Somamides A and B, two new depsipeptide analogs of dolastatin 13 from a Fijian cyanobacterial assemblage of *Lynngbya majuscula* and *Schizothrix* species. *J Nat Prod* 64: 716–719
- Nomizu M, Kuratomi Y, Song SY, Ponce ML, Hoffman MP, Powell SK, Miyoshi K, Otake A, Kleinman HK, Yamada Y (1997) Identification of cell binding sequences in mouse laminin γ 1 chain by systematic peptide screening. *J Biol Chem* 272: 32198–32205
- Norman BH, Shih C (1998) Tripeptide and tetrapeptide pharmaceutical compounds. Patent WO 98 38,178. See Chem Abstr 129: 216922g
- Nuijen B, Bouma M, Henrar RE, Manada C, Bult A, Beijnen JH (1999) Compatibility and stability of apidine, a novel marine-derived depsipeptide antitumor agent, in infusion devices, and its hemolytic and precipitation potential upon i.v. administration. *Anti-Cancer Drugs* 10: 879–887
- Nyeki O, Rill A, Schon I, Orosz A, Schrett J, Bartha L, Nagy J (1998) Synthesis of peptide and pseudopeptide amides inhibiting the proliferation of small cell and epithelial types of lung carcinoma cells. *J Pept Sci* 4: 486–495

- O I, Kieber-Emmons T, Otvos L, Blaszyk-Thurin M (1999) Peptides mimicking sialyl-Lewis A isolated from a random peptide library and peptide array. *Ann NY Acad Sci* 886: 276–279
- Oberg K (2001) Established clinical use of octreotide and lanreotide in oncology. *Chemotherapy* 47 [Suppl 2]: 40–53
- Ohki-Hamazaki H (2000) Neuromedin B. *Prog Neurobiol (Oxford)* 62: 297–312
- Ohlsson B, Fredang N, Axelson J (1999) The effect of bombesin, cholecystokinin, gastrin, and their antagonists on proliferation of pancreatic cancer cell lines. *Scand J Gastroenterol* 34: 1224–1229
- Ohnishi M, Yamaki-Kataoka Y, Kariya K, Tamada M, Hu C-D, Kataoka T (1998) Selective inhibition of Ras interaction with its particular effector by synthetic peptides corresponding to the Ras effector region. *J Biol Chem* 273: 10210–10215
- Okada H, Suzuki H, Yoshinari T, Arakawa H, Okura A, Suda H (1994) A new topoisomerase II inhibitor, BE-22179, produced by a *Streptomyces* I. Producing strain, fermentation, isolation and biological activity. *J Antibiot* 47: 129–135
- Oligino L, Lung FD, Sastry L, Bigelow J, Cao T, Curran M, Burke TRJ, Wang S, Krag D, Roller PP, King CR (1997) Nonphosphorylated peptide ligands for the Grb2 Src homology 2 domain. *J Biol Chem* 272: 29046–29052
- Olsen RK, Apparao S, Bhat KL (1986) Synthesis of a model analogue of the cyclic decapeptide intercalating agent luzopeptin A (antibiotic BBM 928A) containing proline, valine and unsubstituted quinoline constituents. *J Org Chem* 51: 3079–3085
- Omer CA, Kohl NE (1997) CA1A2X-competitive inhibitors of farnesyltransferase as anti-cancer agents. *Trends Pharmacol Sci* 18: 437–444
- Omura S, Fujimoto T, Otoguro K, Matsuzaki K, Moriguchi R, Tanaka H, Sasaki Y (1991a) Lactacystin, a novel microbial metabolite, induces neurogenesis of neuroblastoma cells. *J Antibiot* 44: 113–116
- Omura S, Matsuzaki K, Fujimoto T, Kosuge K, Furuya T, Fujita S, Nakagawa A (1991b) Structure of lactacystin, a new microbial metabolite which induces differentiation of neuroblastoma cells. *J Antibiot* 44: 117–118
- Oppenheim JJ, Murphy WJ, Chertov O, Schirmacher V, Wang JM (1997) Prospects for cytokine and chemokine biotherapy. *Clin Cancer Res* 3: 2682–2686
- Orosz A (2001) Small peptide amides inhibiting the proliferation of cancer cells. Patent WO 01 85,195. See Chem Abstr 135: 366723f
- Orosz A, Nyeki O, Schon I, Kisfaludy L, Schrett J, Bartha L, Nagy J, Ril A, Balogh G (1994) Preparation of small peptide and pseudopeptide amides inhibiting the proliferation of small-cell and epithelial-cell lung cancer cell. Patent EP 556,962. See Chem Abstr 120: 77642q
- Orosz A, Schrett J, Nagy J, Bartha L, Schon I, Nyeki O (1995) New short-chain analogs of a substance-P antagonist inhibit proliferation of human small-cell lung-cancer cells *in vitro* and *in vivo*. *Int J Cancer* 60: 82–87
- Otsuka H, Shōji J (1965) The structure of triostin C. *Tetrahedron* 21: 2931–2938
- Otsuka H, Shoji J, Kawano K, Kyogoku Y (1976) Structure confirmation of triostin A by IH and ¹³C magnetic resonance. *J Antibiot* 29: 107–110
- Otto H-H, Schirmeister T (1997) Cysteine proteases and their inhibitors. *Chem Rev* 97: 133–171
- Packard BS (1987) Identification of a synthetic nonapeptide sequence that inhibits motility in culture of a melanoma subclone that possesses a high metastatic potential. *Proc Natl Acad Sci USA* 84: 9015–9019
- Pacofsky GJ, Lackey K, Alligood KJ, Berman J, Charifson PS, Crosby RM, Dorsey GFJ, Feldman PL, Gilmer TM, Hummel CW, Jordan SR, Mohr C, Shewchuk LM, Sternbach DD, Rodriguez M (1998) Potent dipeptide inhibitors of the pp60c-src SH2 domain. *J Med Chem* 41: 1894–1908
- Paqualini R, Koivunen E, Ruoslahti E (1995) A peptide isolated from phage display libraries is a structural and functional mimic of an RGD-binding site on integrins. *J Cell Biol* 130: 1189–1196
- Patel VF, Andis SL, Kennedy JH, Ray JE, Schultz RM (1999) Novel cryptophycin antitumor agents: synthesis and cytotoxicity of fragment “B” analogues. *J Med Chem* 42: 2588–2603
- Patel VF, Hoard DW, Moher ED, Norman BH (1998) Selective epoxidation process for preparation of cryptophycin compounds and intermediates. Patent EP 861,839. See Chem Abstr 129: 216920e
- Patel YC, Greenwood MT, Panetta R, Demchyshyn L, Niznik H, Srikant CB (1995) Minireview. The somatostatin receptor family. *Life Sci* 57: 1249–1265
- Paulsen JE (1993) The synthetic colon peptide pyroGlu-His-GlyOH inhibits growth of human colon carcinoma cells (HT-29) transplanted subcutaneously into athymic mice. *Carcinogenesis* 14: 1719–1721
- Paulsen JE, Hall KS, Rugstad HE, Reichelt KL, Elgo K (1992) The synthetic hepatic peptides Pyr-Glu-Gly-Ser-Asp and Pyr-Glu-Gly-Gly-Ser-Asp acid inhibit growth of MH1C1 rat hepatoma cells transplanted into buffalo rats or athymic mice. *Cancer Res* 53: 1218–1221
- Pawson T, Schlessinger J (1993) SH2 and SH3 domains. *Curr Biol* 3: 432–434
- Pérez Baz J, Cañedo LM, Fernández Puentes JL, Silva Elipe MV (1997) Thiocoraline, a novel depsipeptide with antitumor activity produced by a marine *Micromonospora* II. Physico-chemical properties and structure determination. *J Antibiot* 50: 738–741
- Pero SC, Oligino L, Daly RJ, Soden AL, Liu C, Roller PP, Li P, Krag DN (2002) Identification of novel non-phosphorylated ligands, which bind selectively to the SH2 domain of Grb7. *J Biol Chem* 277: 119–11926
- Perrin D, Halazy S, Hill BT (1996) Inhibitors of the Ras signal transduction pathway as potential antitumor agents. *J Enzyme Inhib* 11: 77–95
- Petit T, Davidson KK, Lawrence RA, von Hoff DD, Izbicka E (2001) Neuropeptide receptor status in human tumor cell lines. *Anti-Cancer Drugs* 12: 133–136
- Pettit GR (1997) The dolastatins. *Fortschr. Chem Org Naturst* 70: 1–79
- Pettit GR, Kamano Y, Holzapfel CW, van Zyl WJ, Tuinman AA, Herald CL, Baczynskyj L, Schmidt JM (1987) The structure and synthesis of dolastatin 3. *J Am Chem Soc* 109: 7581–7582
- Pettit GR, Kamano Y, Dufresne C, Cerny RL, Herald CL, Schmidt JM (1989a) Isolation and structure of the cytostatic linear depsipeptide dolastatin 15. *J Org Chem* 54: 6005–6006
- Pettit GR, Kamano Y, Herald CL, Dufresne C, Cerny RL, Herald DL, Schmidt JM, Kizu H (1989b) Isolation and structure of the cytostatic depsipeptide dolastatin 13 from the sea hare *Dolabella auricularia*. *J Am Chem Soc* 111: 5015–5017
- Pettit GR, Kamano Y, Herald CL, Dufresne C, Bates RB, Schmidt JM, Cerny RL, Kizu H (1990) Antineoplastic agents. 190. Isolation and structure of the cyclodepsipeptide dolastatin 14. *J Org Chem* 55: 2989–2990
- Pettit GR, Xu J-P, Hogan F, Williams MD, Doubek DL, Schmidt JM, Cerny RL, Boyd MR (1997a) Isolation and structure of the human cancer cell growth inhibitory cyclodepsipeptide dolastatin 16. *J Nat Prod* 60: 752–754
- Pettit GR, Xu J-P, Williams MD, Hogan F, Schmidt JM, Cerny RL (1997b) Antineoplastic agent 370. Isolation and structure of dolastatin 18. *Bioorg Med Chem Lett* 7: 827–832

- Pettit GR, Flahive EJ, Boyd MR, Bai R, Hamel E, Pettit RK, Schmidt JM (1998a) Antineoplastic agent 360. Synthesis and cancer cell growth inhibitory studies of dolastatin 15 structural modifications. *Anti-Cancer Drug Des* 13: 47–66
- Pettit GR, Xu J-P, Hogan F, Cerny RL (1998b) Isolation and structure of dolastatin 17. *Heterocycles* 47: 491–496
- Pickart LR (1989) Glycyl-L-histidyl-lysine copper complexes, their preparation and use as neoplasm inhibitors. Patent WO 88 08,715. See Chem Abstr 111: 70958c
- Piekarz RL, Robey R, Sandor V, Bakke S, Wilson WH, Dahmouh L, Kingma DM, Turner ML, Altemus R, Bates SE (2001) Inhibitor of histone deacetylation, depsipeptide (FR901228), in the treatment of peripheral and cutaneous T-cell lymphoma: a case report. *Blood* 98: 2865–2868
- Piossek C, Schneider-Mergener J, Schirner M, Vakalopoulou E, Germeroth L, Thierauch KH (1999) Vascular endothelial growth factor (VEGF) receptor II-derived peptides inhibit VEGF. *J Biol Chem* 274: 5612–5619
- Plummer MS, Holland DR, Sharipour A, Lunney EA, Fergus JH, Marks JS, McConnell P, Mueller WT, Sawyer TK (1997) Design, synthesis and cocrystal structure of a nonpeptide src SH2 domain ligand. *J Med Chem* 40: 3719–3725
- Poncet J (1999) The dolastatins, a family of promising antineoplastic agents. *Curr Pharm Des* 5: 139–162
- Preston SR, Miller GV, Primrose JN (1996) Bombesin-like peptides and cancer. *Crit Rev Oncology-Hematology* 23: 225–238
- Quigley GJ, Ughetto G, van der marel GA, van Boom JH, Wang AH-J, Rich A (1986) Non-Watson-Crick G.C and A.T base pairs in a DNA-antibiotic complex. *Science* 232: 1255–1259
- Qureshi A, Colin PL, Faulkner DJ (2000) Microsclerodermins F-I, antitumor and antifungal cyclic peptides from the lithistid sponge *Microscleroderma* sp. *Tetrahedron* 56: 3679–3685
- Radulescu RT, Jacques G (2000) Selective inhibition of human lung cancer cell growth by peptides derived from retinoblastoma protein. *Biochem Biophys Res Commun* 267: 71–76
- Radulovic S, Cai R-Z, Serfoso P, Groot K, Redding TW, Pinski J, Schally AV (1991) Biological effects and receptor binding affinities of new pseudonapeptide bombesin/GRP receptor antagonists with N-terminal D-Trp or D-Tpi. *Int J Pept Prot Res* 38: 593–600
- Randazzo A, Dal Piaz F, Orrù S, Debitus C, Roussakis C, Pucci P, Gomez-Paloma L (1998) Axinellins A and B: new proline-containing antiproliferative cyclopeptides from the Vanuatu sponge *Axinella carteri*. *Eur J Org Chem* 2659–2665
- Rashid MA, Gustafson KR, Boswell JL, Boyd MR (2000) Haligramides A and B, two new cytotoxic hexapeptides from the marine sponge *Haliclona nigra*. *J Nat Prod* 63: 956–959
- Raymond E, Ady-Vago N, Ribarg V, Faivres S, Lecot F, Wright T, Lopez-Lazaro L, Guzman C, Jimeno J, Armand JP (2000) Phase I and pharmacokinetics study of aplidine, a marine derived compound, given as a 24h infusion every 2 weeks in patients with advanced solid tumors and non-Hodgkin lymphoma (NHL). *Ann Oncol* 11 (S4): 134
- Raynor K, Murphy WA, Coy DH, Taylor JE, Moreau J-P, Yasuda K, Bell GI, Reisine T (1993) Cloned somatostatin receptor: identification of subtype-selective peptides and demonstration of high affinity binding of linear peptides. *Mol Pharmacol* 43: 838–844
- Reeves R, Nissen MS (1990) The AT-DNA-binding domain of mammalian high mobility group I chromosomal proteins. A novel peptide motif for recognizing DNA structure. *J Biol Chem* 265: 8573–8582
- Reichlin S (1983) Somatostatin. *N Engl J Med* 309: 1556–1563
- Reiher FK, Volpert OV, Jimenez B, Crawford SE, Dinney CP, Henkin J, Haviv F, Bouck NP, Campbell SC (2002) Inhibition of tumor growth by systemic treatment with thrombospondin-1 peptide mimetics. *Int J Cancer* 98: 682–689
- Reile H, R.-Z. C, Armatis P, Schally AV (1995) New antagonists of bombesin/gastrin-releasing peptide with C-terminal Leu (CH2N)Tac-NH2. *Int J Oncol* 7: 749–754
- Reissmann T, Engel J, Kuscher B, Bernd M, Hilgard P, Peukert M, Szeleny I, Reichert S, Gonzalez-Barcena D, Nieschlag E, Comaru-Schally AM, Schally AV (1994) Cetrorelix. *Drugs Future* 19: 228–237
- Rekasi Z, Varga JL, Schally AV, Halmos G, Groot K, Czompoly T (2000) Antagonistic actions of analogs related to growth hormone-releasing hormone (GHRH) on receptors for GHRH and vasoactive intestinal peptide on rat pituitary and pineal cells in vitro. *Proc Natl Acad Sci USA* 97: 1218–1223
- Reuning U, Magdolen V, Wilhelm O, Fisher K, Lutz VH, Schmitt M (1998) Multifunctional potential of the plasminogen activation system in tumor invasion and metastasis (review). *Int J Oncol* 13: 893–906
- Richter JD, Mendez R (2001) CPEB-derived peptides blocking Eg2 kinase activity and their therapeutic uses. Patent WO 01 07,466. See Chem Abstr 134: 143865g
- Ridge RJ, Sloane NH (1996) Partial N-terminal amino acid sequence of the anti-neoplastic urinary protein (ANUP) and the anti-tumour effect of the N-terminal nonapeptide of the unique cytokine present in human granulocytes. *Cytokine* 8: 1–5
- Rinehart Jr KL, Gloer JB, Cook JC, Miszak SA, Scahill TA (1981a) Structures of the didemnins, antiviral and cytotoxic depsipeptides from a Caribbean tunicate. *J Am Chem Soc* 103: 1857–1859
- Rinehart Jr KL, Gloer JB, Hughes Jr. RG, Renis HE, McGovern JP, Swynenberg EB, Stringfellow DA, Kuentzel SL, Li LH (1981b) Didemnins: antiviral and antitumor depsipeptides from a Caribbean tunicate. *Science* 212: 933–935
- Rinehart Jr KL, Kishore V, Bible KC, Sakai R, Sullins DW, Li K-M (1988) Didemnins and tunichlorin: novel natural products from the marine tunicate *Trididemnum solidum*. *J Nat Prod* 51: 1–21
- Rivier J, Porter J, Rivier C, Perrin M, Corrigan AZ, Hook WA, Siraganian RP, Vale WW (1986) New effective gonadotropin releasing hormone antagonists with minimal potency for histamine release in vitro. *J Med Chem* 29: 1846–1851
- Rocheffort H, Liaudet-Coopman E (1999) Cathepsin D in cancer metastasis: a protease and a ligand. *APMIS* 107: 86–95
- Ruhenstroth-Bauer G (1994) Peptide inhibitor of liver cell proliferation. Patent DE 4,224,509. See Chem Abstr 120: 183012u
- Ruhenstroth-Bauer G (1997) Tripeptide for inhibiting the proliferation rate of hepatocytes. Patent US 5,589,461. See Chem Abstr 126: 139869d
- Ruhenstroth-Bauer G, Vogl S, Voelter W, Goldberg M, Topic E (1993) Ser Asp Lys-Ac, a strong inhibitor of liver cell proliferation. *Naturwissenschaften* 80: 314–315
- Ruoslahti E, Reed JC (1994) Anchorage dependence, integrins and apoptosis. *Cell* 77: 477–478
- Ryan RR, Weber HC, Hou W, Sainz E, Mantey SA, Battey JF, Coy DH, Jensen RT (1998) Ability of various bombesin receptor agonists and antagonists to alter intracellular signalling of the human orphan receptor BRS-3. *J Biol Chem* 273: 13613–13624
- Ryu G, Matsunaga S, Fusetani N (1994a) Discodermins E, a cytotoxic and antimicrobial tetradecapeptide from the marine sponge *Discodermia kiiensis*. *Tetrahedron Lett* 35: 8251–8254
- Ryu G, Matsunaga S, Fusetani N (1994b) Discodermins F-H, cytotoxic and antimicrobial tetradecapeptides from the marine sponge *discodermia kiiensis*: structure revision of *Discodermins A-D*. *Tetrahedron* 50: 13409–13416
- Sagami (1984) Hexapeptides. Patent JP 84,152,356. See Chem Abstr 102: 25047h
- Sainz E, Akeson M, Mantey SA, Jensen RT, Battey JF (1998) Four amino acid residues are critical for high affinity binding of

- neuromedin B to the neuromedin B receptor. *J Biol Chem* 273: 15927–15932
- Sakai Y, Yoshida T, Tsujita T, Ochiai K, Agatsuma T, Saitoh Y, Tanaka F, Akiyama T, Akinaga S, Mizukami T (1997) GE3, a novel hexadepsipeptide antitumor antibiotic, produced by *Streptomyces sp.* I. Taxonomy, production, isolation, physico-chemical properties, and biological activities. *J Antibiot* 50: 659–664
- Samy R, Kim HY, Brady M, Toogood PL (1999) Total synthesis of motuporin and 5-[L-Ala]-motuporin. *J Org Chem* 64: 2711–2728
- Sanz-Cervera JF, Stocking EM, Usui T, Osada H, Williams RM (2000) Synthesis and evaluation of microtubule assembly inhibition and cytotoxicity of prenylated derivatives of cyclo-L-Trp-L-Pro. *Bioorg Med Chem* 8: 2407–2415
- Sasse F, Steinmetz H, Heil J, Hoeffle G (2000) Tubulysins, new cytostatic peptides from *Myxobacteria* acting of microtubuli. Production, isolation, physiochemical and biological properties. *J Antibiot* 53: 879–885
- Sattler M, Liang H, Nettessheim D, Meadows RP, Harlan JE, Eberstadt M, Yoon HS, Shuker SB, Chang BS, Minn AJ, Thompson CB, Fesik SW (1997) Structure of BCL-XL-Bak peptide complex: recognition between regulators of apoptosis. *Science* 275: 983–986
- Sawa E, Takahashi M, Kamishohara M, Tazunoki T, Kimura K, Arai M, Miyazaki T, Kataoka S, Nishitoba T (1999) Structural modification of Fas C-terminal tripeptide and its effects on the inhibitory activity of Fas/FAP-1 binding. *J Med Chem* 42: 3289–3299
- Sawyer TK (1998) Src homology-2 domains: structure, mechanisms, and drug discovery. *Biopolymers* 47: 243–261
- Schally AV (1999) Luteinizing hormone-releasing hormone analogs: their impact on the control of tumorigenesis. *Peptides* 20: 1247–1262
- Schasteen CS (1991) Preparation of a peptide with anti-metastasis activity. Patent EP 397, 635. See Chem Abstr 114: 186076e
- Schatz PJ, Chen M-J, Piplani S, Mozsgai CA, Balu P (2001) Peptide compounds having affinity for the vascular endothelial growth factor receptor-2 (VEGFR-2) and associated uses. Patent WO 01 83,693. See Chem Abstr 135: 366772w
- Schiavi B, Richard DJ, Jouillé MM (2002) Total synthesis of isoroquefortine C. *J Org Chem* 67: 620–624
- Schimmer AD, Hedley DW, Chow S, Pham NA, Chakrabarty A, Bouchard D, Mak TW, Trus MR, Minden MD (2001) The BH3 domain of BAD fused to the Antennapedia peptide induces apoptosis via its alpha helical structure and independent of Bcl-2. *Cell Death Differ* 8: 725–733
- Schindler-Horvat JE, Fairchild DG, Hasler C, Tamaszewski JE, Donohue SJ, Tyson CA (1998) Toxicity of jasplakinohide (NSC 613009) in rats and dogs. *Proc Am Assoc Cancer Res* 39: 597
- Schmidt U, Griesser H, Haas G, Kroner M, Riedl B, Schumacher A, Sutoris F, Haupt A, Emling F (1999) Synthesis and cytostatic activities of didemnin derivatives. *J Pept Res* 54: 146–161
- Scholar EM, Viola L, Hexum TD (1987) The antimetastatic activity of enkephalin-like peptides. *Cancer Lett* 35: 133–138
- Sebt SM, Hamilton AD (1997) Inhibition of Ras prenylation: a novel approach to cancer chemotherapy. *Pharmacol Ther* 74: 103–114
- Selivanova G, Iotsova V, Okan I, Fritsche M, Strom M, Groner B, Grafstrom RC, Wiman KG (1997) Restoration of the growth suppression function of mutant p53 by a synthetic peptide derived from p53 C-terminal domain. *Nat Med* 3: 632–638
- Sethi T, Langdon S, Smyth J, Rozengurt E (1992) Growth of small cell lung cancer cells: stimulation by multiple neuropeptides and inhibition by broad spectrum antagonists *in vitro* and *in vivo*. *Cancer Res* 52 (9, Suppl.): 2737s–2742s
- Shakespeare WC (2001) SH2 domain inhibition: a problem solved? *Curr Opin Chem Biol* 5: 409–415
- Sheh L, Cheng JY, Kuan YH, Chen CF (1990) Synthesis of a cyclic hexapeptide with sequence corresponding to murine tumor necrosis factor-(127-132) as a novel potential antitumor agent. *Int J Pept Prot Res* 36: 104–108
- Sheh L, Lin HH, Jeng KCG, Chen CF (1993) Studies of the synthesis, immunology and cytotoxicity of a cyclic octapeptide corresponding to TNF- α -(59-66). *J Med Chem* 36: 4302–4307
- Shepeck JE, Gauss C-M, Chamberlin AR (1997) Inhibition of the Ser-Thr phosphatase PP1 and PP2A by naturally occurring toxins. *Bioorg Med Chem* 5: 1739–1750
- Shibata K, Yamasaki M, Yoshida T, Mizukami T (1998) Preparation of E2F activity inhibitory peptide compounds. Patent WO.98 14,474. See Chem Abstr 128: 283089h
- Shibata K, Yamasaki M, Yoshida T, Mizukami T, Shinkai A, Anazawa H (1999) Preparation of peptides having cyclic structures and exerting p53 protein activity-restoring effect on p53 protein mutants. Patent WO 98 51,707. See Chem Abstr 130: 4096m
- Shih C, Williams DC (1998) Preparation of cryptophycin compounds in combination with synchronizing or activating agents for treating cancer. Patent EP 870,506. See Chem Abstr 129: 302893e
- Shih C, Gossett LS, Gruber JM, Grossman CS, Andis SL, Schultz RM, Worzalla JF, Corbett TH, Metz JT (1999) Synthesis and biological evaluation of novel cryptophycin analogs with modification in the β -alanine region. *Bioorg Med Chem Lett* 9: 69–74
- Shin SY, Kang JH, Hahm K-S (1999) Structure-antibacterial antitumor and hemolytic activity relationships of cecropin A-magainin 2 and cecropin A-melittin hybrid peptides. *J Pept Res* 53: 82–90
- Shioiri T, Hamada Y, Kato S, Shibata M, Kondo Y, Nakagawa H, Kohda K (1987) Cytotoxic activity of cyclic peptides of marine origin and their derivatives: importance of oxazoline functions. *Biochem Pharmacol* 36: 4181–4185
- Shute RE, Dunlap B, Rich DH (1987) Analogues of the cytostatic and antimetogenic agents chlamydocin and HC-toxin: synthesis and biological activity of chloromethyl ketone and diazomethyl ketone functionalized cyclic tetrapeptides. *J Med Chem* 30: 71–78
- Sin N, Kim KB, Elofsson M, Meng L, Auth H, Kwok BHB, Crews CM (1999) Total synthesis of the potent proteasome inhibitor epoxomicin: a useful tool for understanding proteasome biology. *Bioorg Med Chem Lett* 9: 2283–2288
- Singh SB, Zink DL, Polishook JD, Dombrowski AW, Darkin-Rattray SJ, Schmatz DM, Goetz MA (1996) Apicidins: novel tetrapeptides as coccidiostats and antimalarial agents from *Fusarium pallidoroseum*. *Tetrahedron Lett* 37: 8077–8080
- Sirdeshpande BV, Toogood PL (1995) Inhibition of protein synthesis by RA-VII. *Bioorg Chem* 23: 460–470
- Sloane NH (2002) Sixteen residue amino-terminal fragment of the antineoplastic protein (ANUP) as a pharmacologically active antitumor agent. Patent US 2002 61,851. See Chem Abstr 136: 380093c
- Smith CD, Zhang X (1996) Mechanism of action of cryptophycin. Interaction with the Vinca alkaloid domain of tubulin. *J Biol Chem* 271: 6192–6198
- Smitka TA, Deeter JB, Hunt AH, Mertz F, Ellis RM, Boeck LD, Yao RC (1988) A83586C, a new depsipeptide antibiotic. *J Antibiot* 41: 726–733
- Sondak VK, Kopecky KJ, Liu PY, Fletcher WS, Harvey WH, Laufman LR (1994) Didemnin B in metastatic malignant melanoma: a phase II trial of the Southwest Oncology Group. *Anti-Cancer Drugs* 5: 147–150
- Sone H, Nemoto T, Ishiwata H, Ojika M, Yamada K (1993a) Isolation, structures and synthesis of dolastatin D, a cytotoxic

- cyclic depsipeptide from the sea hare *Dolabella auricularia*. *Tetrahedron Lett* 34: 8449–8452
- Sone H, Nemoto T, Ojika M, Yamada K (1993b) Isolation, structure and synthesis of dolastatin C, a new depsipeptide from sea hare *Dolabella auricularia*. *Tetrahedron Lett* 34: 8445–8448
- Sone H, Shibata T, Fujita T, Ojika M, Yamada K (1996) Dolastatin H and isodolastatin H, potent cytotoxic peptides from the sea hare *Dolabella auricularia*: isolation, stereostructure and synthesis. *J Am Chem Soc* 118: 1874–1880
- Sorbera LA, Graul A, Castañer J (2001) Cilengitide. *Drugs Future* 25: 674–678
- Strahl BD, Allis CD (2000) The language of covalent histone modifications. *Nature* 403: 41–45
- Strelkov LA, Mikhailova AA, Fonina LA, Petrov RV (2000) A new endogenous differentiating factor (myeloepitope-4) for myeloid cells. *FEBS Lett* 470: 281–284
- Sugawara K, Toda S, Moriyama T, Konishi M, Oki T (1993) Verucopeptin, a new antitumor antibiotic active against B16 melanoma. Structure determination. *J Antibiot* 46: 928–935
- Supko JG, Lynch TJ, Clark JW, Fram R, Allen LF, Velagapudi R, Kufe DW, Eder JP (2000) A phase I clinical and pharmacokinetic study of the dolastatin analogue cemadotin administered as a 5-day continuous intravenous infusion. *Cancer Chemother Pharmacol* 46: 319–328
- Suzuki M (1990) The heptad repeat in the largest subunit of RNA polymerase II binds by intercalating into DNA. *Nature* 344: 562–565
- Szegedi Z, Takacs J, Szende B, Vadasz Z, Horvath A, Gulyas E, Toth G, Petak I, Bocsi J, Keri G (1999) A specifically radiolabelled somatostatin analog with strong antitumor activity induces apoptosis and accumulates in the cytosol and the nucleus of HT29 human colon carcinoma cells. *Endocrine* 10: 25–34
- Szekeresh T, Fritzer-Szekeresh M, Elford HL (1997) The enzyme ribonucleotide reductase: target for antitumor and anti-HIV therapy. *Crit Rev Clin Lab Sci* 34: 503–528
- Tabor AB (1996) Synthesis of a peptide-intercalator hybrid based on the bZIP motif from GCN4. *Tetrahedron* 52: 2229–2234
- Takahashi K, Koshino H, Esumi Y, Tsuda E, Kurosawa K (2001) Sw-163C and E, novel antitumor depsipeptides produced by *Streptomyces sp.* Part 2. Structure elucidation. *J Antibiot* 54: 622–627
- Takenaka S, Sato H, Itakura Y, Kondo H, Takagi M (1996) Construction of a dimeric DNA-binding peptide model by peptide-anthraquinone conjugation. *Int J Pept Prot Res* 48: 397–400
- Takusagawa F (1985) The role of the cyclic depsipeptide rings in antibiotics. *J Antibiot* 38: 1596–1604
- Talanian RV, McKnight J, Rutkowski R, Kim PS (1992) Minimum length of a sequence-specific DNA binding peptide. *Biochemistry* 31: 6871–6875
- Talpir R, Benayahu Y, Kashman Y, Pannell L, Schleyer M (1994) Hemiasterlin and geodiamolides TA; two new cytotoxic peptides from the marine sponge *Hemiasterella minor* (kirkpatrick). *Tetrahedron Lett* 35: 4453–4456
- Tamura SY, Weinhouse M, Roberts CA, Goldman EA, Masukawa K, Anderson SM, Cohen CR, Bradbury AE, Bernardino VT, Dixon SA, Ma MG, Nolan TB, Brunck TK (2000) Synthesis and biological activity of peptidyl aldehyde urokinase inhibitors. *Bioorg Med Chem Lett* 10: 983–987
- Tan DC, Kini RM, Jois SD, Lim DK, Xin L, Ge R (2001) A small peptide derived from Flt-1 (VEGFR-1) functions as an angiogenic inhibitor. *FEBS Lett* 494: 150–156
- Taunton J, Hassig CA, Schreiber SL (1996) A mammalian histone deacetylase related to the yeast transcriptional regulator Rpd3p. *Science* 272: 408–411
- Taylor JE, Bogden AE, Moreau J-P, Coy DH (1988) *In vitro* and *in vivo* inhibition of human small cell lung carcinoma (NCI-H69) growth by a somatostatin analogue. *Biochem Biophys Res Commun* 153: 81–86
- Taylor S, Giroux DJ, Jaeckle KA, Panella TJ, Dakhil SR, Schold SC (1999) Phase II study of didemnin B in central nervous system tumors: a Southwest oncology group study. *Invest New Drugs* 16: 331–332
- Teicher BA, Ara G, Herbst R, Palombella VJ, Adams J (1999) The proteasome inhibitor PS-341 in cancer therapy. *Clin Cancer Res* 5: 2638–2645
- Terano H, Tsurumi Y, Setoi H, Hashimoto M, Kohsaka M (1986) Amino acid derivatives. Patent EP 193,904. See Chem Abstr 105: 227347d
- Terranova VP, Williams JE, Liotta LA, Martin GR (1984) Modulation of the metastatic activity of melanoma cells by laminin and fibronectin. *Science* 226: 982–985
- Terui Y, Tomizuka H, Mishima Y, Ikeda M, Kasahara T, Uwai M, Mori M, Itoh T, Tanaka M, Yamada M, Shimamura S, Ishizaka Y, Ozawa K, Hatake K (1999) NH₂-terminal pentapeptide of endothelial interleukin 8 is responsible for the induction of apoptosis in leukemic cells and has antitumor effect *in vivo*. *Cancer Res* 59: 5651–5655
- Tew KD (2000) Is there a role for glyoxalase 1 inhibitors as antitumor drugs? *Drug Resist. Updates* 3: 263–264
- Thierry J, Grillon C, Gaudron S, Potier P, Riches A, Wdzieczak-Bakala J (2001) Synthesis and biological evaluation of analogues of the tetrapeptide N-Acetyl-Ser-Asp-Lys-Pro haematopoietic cell proliferation. *J Pept Sci* 7: 284–293
- Thomas F, Arvelo F, Antoine E, Jacrot M, Poupon MF (1992) Antitumor activity of bombesin analogues on small cell lung cancer xenografts: relationship with bombesin receptor expression. *Cancer Res* 52: 4872–4877
- Toda S, Sugawara K, Nishiyama Y, Ohbayashi M, Ohkusa N, Yamamoto H, Konishi M, Oki T (1990) Quinaldopeptin, a novel antibiotics of the quinomycin family. *J Antibiot* 43: 796–808
- Tokita K, Katsuno T, Hocart SJ, Coy DH, Llinares M, Martinez J, Jensen RT (2001) Molecular basis for selectivity of high affinity peptide antagonists for the gastrin-releasing peptide receptor. *J Biol Chem* 276: 36652–36663
- Tomizaki K-Y, Kate T, Nishino N, Yoshida M, Komatsu Y (1999) Histone deacetylase inhibitors based on trapoxin B. *Pept Sci* 35th (Pub. 1999): 181–184. See Chem Abstr 131: 130252t
- Toske SG, Fenical W (1995) Cyclodidemannamide: a new cyclic heptapeptide from the marine ascidian *Didemnum molle*. *Tetrahedron Lett* 36: 8355–8358
- Tressler RJ, Pitot PA, Stratton JRL, Forrest LD, Zhuo S, Drummond RJ, Fong S, Doyle MV, Doyle LV, Min HY, Rosenberg S (1999) Urokinase receptor antagonists: discovery and application to *in vivo* models of tumor growth. *APMIS* 107: 168–173
- Tsuda M, Ishiyama H, Masuko K, Takao T, Shimonishi Y, Kobayashi J (1999) Keramamides M and N, two new cyclic peptides with a sulfate ester from *Theonella sponge*. *Tetrahedron* 55: 12543–12548
- Tsukamoto S, Painuly P, Young KA, Yang X, Shimizu Y (1993) Microcystilide A: a novel cell-differentiation-promoting depsipeptide from *Microcystis aeruginosa* NO-15-1840. *J Am Chem Soc* 115: 11046–11047
- Uchihata Y, Ando N, Ikeda Y, Kondo S, Hamada M, Umezawa K (2002) Isolation of a novel cyclic hexadepsipeptide pipalamycin from *streptomyces* as an apoptosis-inducing agent. *J Antibiot* 55: 1–5
- Ueda I, Niwa M, Uchiyama F, Ono H (1986) Oligopeptides. Patent JP 60,214,768. See Chem Abstr 105: 43331n
- Ueda H, Manda T, Matsumoto S, Mukumoto S, Nishigaki F, Kawamura I, Shimomura K (1994a) FR901228, a novel antitumor

- bicyclic depsipeptide produced by *Chromobacterium violaceum* No. 968 III. Antitumor activities on experimental tumors in mice. *J Antibiot* 47: 315–323
- Ueda H, Nakajima H, Hori Y, Fujita T, Nishimura M, Goto T, Okuhara M (1994b) FR901228, a novel antitumor bicyclic depsipeptide produced by *Chromobacterium violaceum* No. 968 I. Taxonomy, fermentation, isolation, physicochemical and biological properties and antitumor activity. *J Antibiot* 47: 301–310
- Uemoto H, Yahiro Y, Shigemori H, Tsuda M, Takao T, Shimonishi Y, Kobayashi J (1998) Keramamides K and L, new cyclic peptides containing unusual tryptophan residue from *Theonella* sponge. *Tetrahedron* 54: 6719–6724
- Umehara K, Nakahara K, Kiyoto S, Iwami M, Okamoto M, Tanaka H, Kohsaka M, Aoki H, Imanaka H (1983) Studies on WF-3161, a new antitumor antibiotic. *J Antibiot* 36: 478–483
- Umezawa K, Nakazawa K, Ikeda Y, Naganawa H, Kondo S (1999) Polyoxypeptins A and B produced by *Streptomyces*: apoptosis-inducing cyclic depsipeptides containing the novel amino acid (2S,3R)-3-hydroxy-3-methylproline. *J Org Chem* 64: 3034–3038
- Umezawa K, Ikeda Y, Uchihata Y, Naganawa H, Kondo S (2000) Chloptosin, an apoptosis-inducing dimeric cyclohexapeptide produced by *Streptomyces*. *J Org Chem* 65: 459–463
- Usui T, Kondoh M, Cui CB, Mayumi T, Osada H (1998) Tryprostatin A, a specific and novel inhibitor of microtubule assembly. *Biochem J* 333: 543–548
- Vaishampayan U, Glode M, Du W, Kraft A, Hudes G, Wright J, Hussain M (2000) Phase II study of dolastatin-10 in patients with hormone-refractory metastatic prostate adenocarcinoma. *Clin Cancer Res* 6: 4205–4208
- Varga JL, Schally AV, Csernus VJ, Zarandi M, Halmos G, Groot K, Rekasi Z (1999) Synthesis and biological evaluation of antagonists of growth hormone-releasing hormone with high and protracted in vivo activities. *Proc Natl Acad Sci USA* 96: 692–697
- Veber DF, Freidinger RM, Perlow DS, Palaveda WJ, Holly FRW, Strachan G, Nutt RF, Arison BJC, Homnick WC, Randall MS, Glitzer R, Saperstein R, Hirschmann R (1981) A potent cyclic hexapeptide analogue of somatostatin. *Nature* 292: 55–58
- Velders MP, Nieland JD, Rudolf MP, Lovisecek K, Weijzen S, De Visser KE, Macedo MF, Carbone M, Kast WM (1998) Identification of peptides for immunotherapy of cancer. It is still worth the effort. *Crit Rev Immunol* 18: 7–27
- Vera MD, Joullié MM (2002) Natural products as probes of cell biology: 20 years of didemnin research. *Med Res Rev* 22: 102–145
- Vervoort H, Fenical W (2000) Tamandarins A and B: new cytotoxic depsipeptides from a Brazilian ascidian of the family *Didemnidae*. *J Org Chem* 65: 782–792
- Villalona-Calero MA, Baker SD, Hammond L, Aylesworth C, Eckhardt SG, Kraynak M, Fram R, Fischkoff S, Velagapudi R, Toppmeyer D, Razvillas B, Jakimowicz K, Van Hoff DD, Rowinsky E (1998) Phase I and pharmacokinetic study of the water-soluble dolastatin 15 analog LU103793 in patients with advanced solid malignancies. *J Clin Oncology* 16: 2770–2779
- Vince R, Brownell J, Akella LB (1999) Synthesis and activity of g-(L-g-azaglutamyl)-S-(p-bromobenzyl)-L-cysteinylglycine: a metabolically stable inhibitor of glyoxalase 1. *Bioorg Med Chem Lett* 9: 853–856
- von Angerer E (2000) Tubulin as a target for anticancer drugs. *Curr Opin Drugs Discovery Dev* 3: 575–584
- von Closse A, Huguenin R (1974) Isolierung und strukturaufklärung von chlamydocin. *Helv Chim Acta* 57: 533–545
- Wadler S, Tenteromano L, Cazenave L, Sparano JA, Greenwald ES, Rozenblit A, Kaleyra R, Wiernik PH (1994) Phase II trial of echinomycin in patients with advanced or recurrent colorectal cancer. *Cancer Chemother Pharmacol* 34: 266–269
- Wakita K-I, Minami M, Venkateswarlu A, Sharma VM, Ramesh M, Akahane K (2001) Antitumor bicyclic hexapeptide RA-VII modulates cyclin D1 protein level. *Anti-Cancer Drugs* 12: 433–439
- Waksman G, Shoelson SE, Pant N, Cowburn D, Kuriyan J (1993) Binding of a high affinity phosphotyrosyl peptide to the Src SH2 domain: crystal structures of the complexed and peptide-free forms. *Cell* 72: 779–790
- Wang J-L, Zhang Z-J, Choksi S, Shan S, Lu Z, Croce CM, Alnemri ES, Korngold R, Huang Z (2000) Cell permeable Bcl-2 binding peptides: a chemical approach to apoptosis induction in tumor cells. *Cancer Res* 60: 1498–1502
- Waring MJ, Wakelin LP (1974) Echinomycin: a bifunctional intercalating antibiotic. *Nature* 252: 653–657
- Watanabe H, Kamata S, Fukuda T (1998) Preparation of tripeptide analogs containing benzoxazepine derivatives as cysteine protease inhibitors. Patent JP 09,295,996. See Chem Abstr 128: 35022x
- Webster KR, Coleman KG (1994) Peptide inhibitors of the p33cdk2 and p34cdk2 cell cycle regulatory kinase and human papillomavirus E7 oncoprotein for inhibition of papillomavirus-associated neoplastic transformation. Patent EP 666,270. See Chem Abstr 123: 218395n
- Weckbecker G, Raulf F, Stolz B, Bruns C (1993) Somatostatin analogs for diagnosis and treatment of cancer. *Pharmacol Ther* 60: 245–264
- Weidle UH, Koenig B (1998) Urokinase receptor antagonists: novel agents for the treatment of cancer. *Expert Opin Invest Drugs* 7: 391–403
- Weinberg RA (1995) The retinoblastoma protein an cell cycle control. *Cell* 81: 323–330
- Wesson KJ, Hamann MT (1996) Keenamide A, a bioactive cyclic peptide from the marine mollusk *Pleurobranchus forskalii*. *J Nat Prod* 59: 629–631
- White JD, Hong J, Robarge LA (1999) Total Synthesis of Cryptophycins-1,-3-4-24 (Arenastatin A), and -29, cytotoxic depsipeptides from cyanobacteria of the *Nostocaceae*. *J Org Chem* 64: 6206–6216
- Whittaker M, Floyd CD, Brown P, Gearing AJH (1999) Design and therapeutic application of matrix metalloproteinase inhibitors. *Chem Rev* 99: 2735–2776
- Wieland T (1968) Poisonous principle of mushrooms of the genus *Amanita*. *Science* 159: 946–952
- Wieland T (1987) 50 Jahre Phalloidin. Seine Entdeckung, Charakterisierung sowie gegenwärtige und zukünftige Anwendung in der Zellforschung. *Naturwissenschaften* 74: 367–373
- Wieland T, Faulstich H (1978) Amatoxins, phallotoxins, phallolysin, and antamanide: the biologically active components of poisonous *Amanita* mushrooms. *CRC Crit Rev Biochem* 5: 185–260
- Williams DE, Moore RB, Paul VJ (1989) The structure of ulithiacyclamide B. Antitumor evaluation of cyclic peptides and macrolides from *Lissoclinum patella*. *J Nat Prod* 52: 732–739
- Williams E, Williams G, Gour BJ, Blaschuk OW, Doherty P (2000) A novel family of cyclic peptide antagonists suggests that N-cadherin specificity is determined by amino acids that flank the HAV motif. *J Biol Chem* 275: 4007–4012
- Williamson MP, Gauvreau D, Williams DH, Waring MJ (1982) Structure and conformation of fourteen antibiotics of the quinoxaline group determined by ¹H NMR. *J Antibiot* 35: 62–66
- Wipf P, Miller CP (1992) Total synthesis of westiellamide. *J Am Chem Soc* 114: 10975–10977
- Wipf P, Uto Y (1999) Total synthesis of the putative structure of the marine metabolite trunkamide A. *Tetrahedron Lett* 40: 5165–5169

- Woodburn JR (1999) The epidermal growth factor receptor and its inhibition in cancer therapy. *Pharmacol Ther* 82: 241–250
- Xu J-X, Jin S (1998) Synthesis and antitumor activities of analogs and segments of *Papaver somniferum pollen* tridecapeptide. *Pept.: Biol Chem., Proc Chin. Pept Symp.* 4th 71–74. See Chem Abstr 129: 230985m
- Yamaguchi H, Asai A, Mizukami T, Yamashita Y, Akinaga S, Ikeda S, Kanda Y (2000) Preparation of UCK 14A2 derivative as proteasome inhibitors. Patent WO 00 43,000. See Chem Abstr 133: 120677n
- Yamasaki L (1999) Balancing proliferation and apoptosis in vivo the goldilocks theory of E2F/DP action. *Biochim. Biophys Acta* 1423: M9–M15
- Yao Z-J, King CR, Kelley J, Milne GW, Voigt JH, Burke Jr TR (1999) Potent inhibition of grb2 SH2 domain binding by non-phosphate-containing ligands. *J Med Chem* 42: 25–35
- Yeung BKS, Nakao Y, Kinnel RB, Carney JR, Yoshida WY, Sceuer PJ, Kelly-Borges M (1996) The kapakahines, cyclic peptides from the marine sponge *Cribrorchalina olemda*. *J Org Chem* 61: 7168–7173
- Yoshida M, Furumai R, Nishiyama M, Komatsu Y, Nishino N, Horinouchi S (2001) Histone deacetylase as a new target for cancer chemotherapy. *Cancer Chemother Pharmacol* 48 [Suppl 1]: S20–S26
- Yoshinari T, Okada H, Yamada A, Uemura D, Oka H, Suda H, Okura A (1994) Inhibition of topoisomerase II by a novel antitumor cyclic depsipeptide, BE-22179. *Jpn. J Cancer Res* 85: 550–555
- You S-A, Basu A, Haldar S (1999) Potent antitumor agent proteasome inhibitors; a novel trigger for Bcl2 phosphorylation to induce apoptosis. *Int J Oncol* 15: 625–628
- Yu X, Guo ZS, Marcu MG, Neckers L, Nguyen DM, Chen GA, Schrupp DS (2002) Modulation of p53, ErbB1, ErbB2, and Raf-1 expression in lung cancer cells by depsipeptide FR901228. *J Natl Cancer Inst* 94: 504–513
- Zabriskie TM, Klocke JA, Ireland JM, Marcus AH, Molinski TF, Faulkner DJ, Xu C, Clardy J (1986) Jaspamide, a modified peptide from *Jaspis* sponge, with insecticidal and antifungal activity. *J Am. Chem Soc* 108: 3123–3124
- Zalacain M, Zaera E, Vázquez D, Jiménez A (1982) The mode of action of the antitumor drug bouvardin, an inhibitor of protein synthesis in eukaryotic cells. *FEBS Lett* 148: 95–97
- Zampella A, Giannini C, Debitus C, Roussakis C, D’Auria MV (1999) New jaspamide derivatives from the marine sponge *Jaspis splendans* collected in Vanuatu. *J Nat Prod* 62: 332–334
- Zhang LH, Longley RE (1999) Induction of apoptosis in mouse thymocytes by microcolin A and its synthetic analog. *Life Sci* 64: 1013–1028
- Zhang X, Patel DJ (1991) Solution structure of the luzopeptin-DNA complex. *Biochemistry* 30: 4026–4041
- Zhao M, Kleinman HK, Mokotoff M (1994) Synthetic laminin-like peptides and pseudopeptides as potential antimetastatic agents. *J Med Chem* 37: 3383–3388
- Zhao S, Smith KS, Deveau AM, Dieckhaus CM, Johnson MA, MacDonald TL, Cook JM (2002) Biological activity of the tryprostatins and their diastereoisomers on human carcinoma cell lines. *J Med Chem* 45: 1559–1562
- Zheleva DI, Fischer PM, McInnes C, Andrews MJI, Chan WC, Atkinson GE (2002) Cyclin-inhibiting p21 peptides. Patent WO 01 40,142. See Chem Abstr 135: 14300r
- Zhu D-M, Uckun FM (2000) Z-Phe-Gly-NHO-Bz, an inhibitor of cysteine cathepsins, induces apoptosis in human cancer cells. *Clin Cancer Res* 6: 2064–2069

Note added in proof

Concerning cell signalling, a remarkable book entitled “Signal Transduction” written by Gomperts BD, Kramer IM and Tatham PER (Academic Press) is a very good comprehensive contribution illustrating the complexity of the all the proteins involved.

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